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Patentanmeldung Nr. Patent application No. Demande de brevet n°

02077908.8

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R C van Dijk

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Expressive Research B.V.
Bornsesteeg 59
6708 PD Wageningen
PAYS-BAS

Bezeichnung der Erfindung/Title of the invention/Titre de l'invention:
(Falls die Bezeichnung der Erfindung nicht angegeben ist, siehe Beschreibung.
If no title is shown please refer to the description.
Si aucun titre n'est indiqué se referer à la description.)

Modulating developmental pathway in plants

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Title: Modulating developmental pathways in plants.

The invention relates to a method to modulate plant growth or development by modifying genes in plants. The invention among others relates to modifying RKS genes or gene products as found in Arabidopsis thaliana or other plants. The different domains of RKS gene products essentially have the following functions: The first domain of the predicted protein structure at the N-terminal end consists of a signal sequence, involved in targeting the protein towards the plasma membrane. Protein cleavage removes this sequence from the final mature protein product (Jain et al. 1994, J. Biol. Chemistry 269: 16306-16310). The second domain consists of different numbers of leucine zipper motifs, and is likely to be involved in protein protein dimerization. The next domain contains a conserved pair of cystein residues, involved in disulphate bridge formation. The next domain consists of 5 (or in the case of RKS3 only 4) leucine rich repeats (LRRs) shown in a gray colour, likely to be involved in ligand binding (Kobe and Deisenhofer 1994, TIBS 19: 415-420). This domain is again bordered by a domain containing a conserved pair of cystein residues involved in disulphate bridge formation often followed by a serine / proline rich region. The next domain displays all the characteristics of a simgle transmembrane domain. At the predicted cytoplasmic site of protein a domain is situated with unknown function, followed by a domain with serine /threonine kinase activity (Schmidt et al. 1997, Development 124: 2049-2062, WO 01/29240). The kinase domain is followed by a domain with unknown function whereas at the C-terminal end of the protein part of a leucine rich repeat is positioned, probably involved in protein-protein interactions.

Plant homologs of the Arabidopsis RKS genes can be found by comparison of various plant database (see also Table 2) and comprises amongst others:

Y14600|SBRLK1|Sorghum bicolor
BF004020|BF004020|EST432518 KV1 Medicago truncatata
AW934655|AW934655|EST353547 tomato
AW617954|AW617954|EST314028 L. pennellii
AA738544|AA738544|SbRLK2 Sorghum bicolor
AA738545|AA738545|SbRLK3 Sorghum bicolor
BG595415|BG595415|EST494093 cSTS Solanum tuberosa
AI896277|AI896277|EST265720 tomato
BF643238|BF643238|NF002H05EC1F1045
AA738546|AA738546|SbRLK4 Sorghum bicolor
BE658174|BE658174|GM700005A20D5 Gm-r1070 Glycine max
BF520845|BF520845|EST458318 DSIL Medicago truncata
AC069324|AC069324|Oryza sativa
AW761055|AW761055|sl70d06.y1 Gm-c1027 Glycine max
BE352622|BE352622|WHE0425_G11_M21ZS Wheat
BG647340|BG647340|EST508959 HOGA Medicago truncata
AY028699|AY028699|Brassica napus
AW666082|AW666082|sk31h04.y1 Gm-c1028 Glycine max
AA738547|AA738547|SbRLK5 Sorghum bicolor
BG127658|BG127658|EST473220 tomato
L27821|RICPRKI|Oryza sativa
BG238468|BG238468|sab51a09.y1 Gm-c1043 Glycine max
BG441204|BG441204|GA__Ea0012C15f Gossypium arbo.
AW667985|AW667985|GA__Ea0012C15 Gossypium arbore.
AW233982|AW233982|sf32g05.y1 Gm-c1028 Glycine max
AP003235|AP003235|Oryza sativa
BF460294|BF460294|074A05 Mature tuber
AY007545|AY007545|Brassica napus
AC087544|AC087544|Oryza sativa
AB041503|AB041503|Populus nigra

The invention furthermore relates to modifying ELS genes or gene products or functional equivalents thereof which are for example derived from at least three different genes in the Arabidopsis genome. They show high homology on protein level

with the corresponding transmembrane RKS gene products. However, they lack a transmembrane domain while they do contain a signalling sequence at the N-terminal end. Therefore these proteins are thought to be positioned within vesicles within the plant cell or at the outsite of the plasma membrane, within the cell wall of the plant cell. A number of homologs have been detected in other plant species, such as:

AF370543|AF370543|Arabidopsis thaliana
AF324989|AF324989|Arabidopsis thaliana
AV520367|AV520367|Arabidopsis thaliana
AV553051|AV553051|Arabidopsis thaliana
BF642233|BF642233|NF050C09IN1F1069
AW559436|AW559436|EST314484 DSIR Medicago truncata
BG456991|BG456991|NF099F02PL1F1025
AW622146|AW622146|EST312944 tomato
BF260895|BF260895|HVSMEf0023D15f Hordeum vulgare
BE322325|BE322325|NF022E12IN1F1088
BG414774|BG414774|HVSMEk0003K21f Hordeum vulgare
BE460627|BE460627|EST412046 tomato
BI204894|BI204894|EST522934 cTOS Lycopersicon esculentum
BI205306|BI205306|EST523346 cTOS Lycopersicon esculentum
BI204366|BI204366|EST522406 cTOS Lycopersicon esculentum
AW443205|AW443205|EST308135 tomato
AW031110|AW031110|EST274417 tomato
BI180080|BI180080|EST521025 cSTE Solanum tuberosa
BF644761|BF644761|NF015A11EC1F1084
AV526127|AV526127|Arabidopsis thaliana
AV556193|AV556193|Arabidopsis thaliana
BE203316|BE203316|EST403338 KV1 Medicago truncatata.
AW649615|AW649615|EST328069 tomato
BE512465|BE512465|946071E06
BI204917|BI204917|EST522957 cTOS Lycopersicon esculentum
BG590749|BG590749|EST498591
BG648725|BG648725|EST510344 HOGA Medicago truncata
BG648619|BG648619|EST510238 HOGA Medicago truncata
BG597757|BG597757|EST496435 cSTS Solanum tuberosa
AW221939|AW221939|EST298750 tomato
BE704836|BE704836|Sc01_
BG124409|BG124409|EST470055 tomato

BF051954|BF051954|EST437120 tomato
BG320355|BG320355|Zm03_05h01_zea mais
AV526624|AV526624|Arabidopsis thaliana
AW933960|AW933960|EST359803 tomato
AW221278|AW221278|EST297747 tomato
BE405514|BE405514|WHE1212_C01_F02ZS Wheat
BG314461|BG314461|WHE2495_A12_A23ZS Triticum
BF258673|BF258673|HVSMEf0016G01f Hordeum vulgare
BG262637|BG262637|WHE0938_E03_I06ZS Wheat
AW030188|AW030188|EST273443 tomato
BG653580|BG653580|sad76b11.y1 Gm-c1051 Glycine max
BG319729|BG319729|Zm03_05h01_A Zm03_zea mais
BF053590|BF053590|EST438820 potato
BE454808|BE454808|HVSMEh0095C03f Hordeum vulgare
BI075801|BI075801|IP1_21_D05.b1_A002
BE367593|BE367593|PI1_9_F02.b1_A002sorghum bicolor
2e-074 BF260080|BF260080|HVSMEf0021A22f Hordeum vulgare
BF627921|BF627921|HVSMEb0006I23f Hordeum vulgare
BG598491|BG598491|EST503391 cSTS Solanum tuberosa
AW038168|AW038168|EST279825 tomato
BG343258|BG343258|HVSMEg0005D23f Hordeum vulgare
AW925684|AW925684|HVSMEg0005D23 Hordeum vulgare
BG416093|BG416093|HVSMEk0009L18f Hordeum vulgare
AW683370|AW683370|NF011C09LF1F1069
BE420108|BE420108|WWS020.C1R000101 ITEC WWS Wheat
AW350720|AW350720|GM210009A10F4 Gm-r1021 Glycine max
AW616564|AW616564|EST322975 L. hirsutum trichome
AW011134|AW011134|ST17B03 Pine
BF630746|BF630746|HVSMEb0013N06f Hordeum vulgare
AW926045|AW926045|HVSMEg0006C10 Hordeum vulgare
BE519800|BE519800|HV_CEb0021E12f Hordeum vulgare
BG343657|BG343657|HVSMEg0006C10f Hordeum vulgare
BG933682|BG933682|OV1_16_C09.b1_A002
BE433368|BE433368|EST399897 tomato
AW219797|AW219797|EST302279 tomato
BF629324|BF629324|HVSMEb0010N06f Hordeum vulgare
BE597128|BE597128|PI1_71_A07.g1_A002
AW220075|AW220075|EST302558 tomato
AW616639|AW616639|EST323050 L. hirsutum trichome
BF645214|BF645214|NF032F11EC1F1094
AW924540|AW924540|WS1_70_H12.b1_A002

AI775448|AI775448|EST256548 tomato
AW983360|AW983360|HVSMEg0010F15f Hordeum vulgare
BF270171|BF270171|GA__Eb0007B13f Gossypium arbor.
BE919631|BE919631|EST423400 potato
AW037836|AW037836|EST279465 tomato
BF008781|BF008781|ss79h09.y1 Gm-c1064 Glycine max
BF254651|BF254651|HVSMEf0004K05f Hordeum vulgare
BE599797|BE599797|PI1_79_H01.g1_A002
BE599026|BE599026|PI1_86_E03.g1_A002
R89998|R89998|16353 Lambda-PRL2 Arabidopsis
BG841108|BG841108|MEST15-G02.T3 ISUM4-TN Zea maize
AW307218|AW307218|sf54c07.y1 Gm-c1009 Glycine max
AI496325|AI496325|sb05c09.y1 Gm-c1004 Glycine max
AJ277703|ZMA277703|Zea mays
AL375586|CNS0616P|Medicago truncatula EST
AW350549|AW350549|GM210009A10A12 Gm-r1021 Glycine max
BE125918|BE125918|DG1_59_F02.b1_A002
BF053901|BF053901|EST439131 potato
BE921389|BE921389|EST425266 potato
BE597551|BE597551|PI1_71_A07.b1_
BE360092|BE360092|DG1_61_C09.b1_A002
BE660084|BE660084|491 GmaxSC Glycine max
AJ277702|ZMA277702|Zea mays

The invention also relates to modifying SBP/SPL gene or products which represent a family of transcription factors with a bipartite nuclear localization signal (The SQUAMOSA PROMOTER-BINDING PROTEIN-LIKE (SBP/SPL) gene family of Arabidopsis thaliana, Columbia ecotype). Upon activation (probably by RKS mediated phosphorylation, the bipartite nuclear localization signal becomes linear and available for the nuclear translocation of the protein. Within the plant nucleus, the transcription factor regulates transcription by interaction with specific promoter elements. In Arabidopsis thaliana, this family is represented by at least 16 different members:

name	genetic code
ATSPL1	At2g47070*
ATSPL2	At5g43270
ATSPL3	At2g33810*

ATSPL4	At1g53160*
ATSPL5	At3g15270
ATSPL6	At1g69170
ATSPL7	At5g18830
ATSPL8	At1g02065
ATSPL9	At2g42200*
ATSPL10	At1g27370*
ATSPL11	At1g27360*
ATSPL12	At3g60030
ATSPL13	At5g50570
ATSPL14	At1g20980
ATSPL15	At3g57920
ATSPL16	At1g76580

* annotation in database not complete and/or correct

In many other plant species, we identified members of this transcription factor family, plant homologs of the Arabidopsis SBP/SPL proteins are for example:

AB023037|AB023037|Arabidopsis thaliana
BG789832|BG789832|sae56b07.y1 Gm-c1051 Glycine max
BG123992|BG123992|EST469638 tomato
BG595750|BG595750|EST494428 cSTS Solanum tuberosum
AF370612|AF370612|Arabidopsis thaliana
BF728335|BF728335|1000060H02.x1 1000 - zea mays
X92079|AMSBP2|A.majus
AW331087|AW331087|707047A12.x1 707 - Mixed adult... 128 zea mays
AJ011643|ATH011643|Arabidopsis thaliana
L34039|RICRMSOA|Oryza sativa
AJ011638|ATH011638|Arabidopsis thaliana
AJ011639|ATH011639|Arabidopsis thaliana
AJ132096|ATH132096|Arabidopsis thaliana
BF482644|BF482644|WHE2301-2304_A21_A21ZS Wheat
BF202242|BF202242|WHE0984_D01_G02ZS Wheat
BE057470|BE057470|sm58e10.y1 Gm-c1028 Glycine max
AJ011628|ATH011628|Arabidopsis thaliana
AJ011629|ATH011629|Arabidopsis thaliana
AJ011617|ZMA011617|Zea mays
AJ011637|ATH011637|Arabidopsis thaliana
AJ011622|AMA011622|Antirrhinum majus

AJ011621|AMA011621|Antirrhinum majus
AJ011635|ATH011635|Arabidopsis thaliana
AJ011623|AMA011623|Antirrhinum majus
BF650908|BF650908|NF098D09EC1F1076
AJ242959|ATH242959|Arabidopsis thaliana
Y09427|ATSPL3|A.thaliana mRNA
AJ011633|ATH011633|Arabidopsis thaliana
AW691786|AW691786|NF044B06ST1F1000
BE058432|BE058432|sn16a06.y1 Gm-c1016 Glycine max
AW728623|AW728623|GA__Ea0017G06 Gossypium arbore.
BG442540|BG442540|GA__Ea0017G06f Gossypium arbo.
AJ011626|ATH011626|Arabidopsis thaliana
AJ011625|ATH011625|Arabidopsis thaliana
AI993858|AI993858|701515182 A. thaliana
BG593787|BG593787|EST492465 cSTS Solanum tuberosum
BF634536|BF634536|NF060C08DT1F1065 Drought Medicago
BE806499|BE806499|ss59f10.y1 Gm-c1062 Glycine max
AW933950|AW933950|EST359793 tomato
AC008262|AC008262| Arabidopsis
B28493|B28493|T10A24TF TAMU Arabidopsis thaliana
AJ011644|ATH011644|Arabidopsis thaliana
AC018364|AC018364|Arabidopsis thaliana
AL092429|CNS00VLB|Arabidopsis thaliana
BE435668|BE435668|EST406746 tomato
BG097153|BG097153|EST461672 potato
BE440574|BE440574|sp47b09.y1 Gm-c1043 Glycine max
AI443033|AI443033|sa31a08.y1 Gm-c1004 Glycine max
U89496|ZMU89496|Zea mays liguleless1
AW433271|AW433271|sh54g07.y1 Gm-c1015 Glycine max
AW932595|AW932595|EST358438 tomato
AW096676|AW096676|EST289856 tomato
AJ011616|ZMA011616|Zea mays
AW036750|AW036750|EST252139 tomato
BF626329|BF626329|HVSMEa0018F24f Hordeum vulgare
AJ011614|ZMA011614|Zea mays
AJ011642|ATH011642|Arabidopsis thaliana
BE022435|BE022435|sm85h04.y1 Gm-c1015 Glycine max
X92369|AMSPB1|A.majus
AC015450|AC015450|Arabidopsis thaliana
AC079692|AC079692|Arabidopsis thaliana
AJ011632|ATH011632|Arabidopsis thaliana

AJ011631|ATH011631|Arabidopsis thaliana
BE455349|BE455349|HVSMEh0097E20f Hordeum vulgare
AJ242960|ATH242960|Arabidopsis thaliana
AJ011610|ATH011610|Arabidopsis thaliana
AJ132097|ATH132097|Arabidopsis thaliana
AL138658|ATT2O9|Arabidopsis thaliana
AJ011615|ZMA011615|Zea mays
BE499739|BE499739|WHE0975_ Wheat
AW398794|AW398794|EST309294 L. pennellii
AJ011618|ZMA011618|Zea mays
AW747167|AW747167|WS1_66_F11.b1_
AJ011577|ATH011577|Arabidopsis thaliana
AI992727|AI992727|701493410 A. thaliana
BE060783|BE060783|HVSMEg0013F15f Hordeum vulgare
BE804992|BE804992|ss34h10.y1 Gm-c1061 Glycine max
BE325341|BE325341|NF120H09ST1F1009
AC007369|AC007369|Arabidopsis thaliana
AJ011619|ZMA011619|Zea mays
BI099345|BI099345|IP1_37_H10.b1_A002
BI071295|BI071295|C054P79U Populus
AZ920400|AZ920400|1006019G01.y2 1006 -
AZ919034|AZ919034|1006013G02.x3 1006 -
BE805023|BE805023|ss35d09.y1 Gm-c1061 Glycine max
BG582086|BG582086|EST483824 GVN Medicago truncata
AJ011609|ATH011609|Arabidopsis thaliana
BE023083|BE023083|sm90e08.y1 Gm-c1015 Glycine max

Furthermore, the invention relates to modifying NDR-NHL- genes or gene products. All proteins belonging to this family contain one (and sometimes even more than one) transmembrane domain. Arabidopsis contains a large number of NDR-NHL genes, such as:
aad21459, aaf18257, aac36175, k10d20 (position 40852-41619), aad21460, cab78082, aad21461, aad42003, aaf02134, aaf187656, aaf02133, cab43430, cab88990, cab80950, aad25632, aaf23842, al163812, f20d21-35, t13m11-12, f1e22-7, t23g18, f5d14-4266, t32f12-16, f11f19-11, f11f19-12, f11f19-13, t20p8-13, f12k2, f23h14, k10d20-44043, k10d20-12, t19f11-6, t19f11-5, t10d17-10, f22o6-150, f3d13-5, m3e9-80, t25p22-30, mhf15-4, mhf15-5, mrn17-4, m1f18-9, mgn6-11994, mjj3-9667, f14f18-60, At1g17620 F11A6, At5g11890 , At2g27080 , At5g36970 ,

mlf18 , At1g65690 F1E22 , At4g01110 F2N1 , At2g35980 f11f19 , At4g01410 F3D13 , At1g54540 F20D21 , At2g46300 t3f17 , At5g21130 , At3g11650 T19F11 , At5g06320 MHF15 , At5g06330 MHF15 , At2g01080 f15b18 , At2g35460 t32f12 , At2g27260 f12k2 , At2g35970 f11f19 , At5g53730 MGN6 , At5g22870 MRN17 , At4g09590 , At3g54200 , At1g08160 T6D22 , At5g22200 , At3g52470 , At2g35960 f11f19 , At3g52460 , At5g56050 MDA7, At3g20590 K10D20 , At1g61760 T13M11 , At3g20600 K10D20 , At1g13050 F3F19 , At3g11660 T19F11 , At3g44220 , At1g64450 F1N19 , At3g26350 F20C19 C , At4g05220 , At5g45320 K9E15 , At4g23930 , At4g13270 , At4g39740 , At1g45688 F2G19 W , At5g42860 MBD2 , At1g32270 F27G20 , At4g30660 , At2g45430 f4123 , At4g30650 , At1g69500 F10D13

and

ndr2, At2g27080; T20P8.13, At5g21130, At1g65690, At5g36970, At1g54540, At5g06320, At5g11890, At1g17620, At3g11650, At2g22180, At5g22870, At2g35980, At2g46300, At4g05220, At2g35460, At2g27260, At4g01410, At5g22200, At1g61760, At3g52470, At5g53730, At4g01110, At2g35960, At3g52460, At4g09590, At2g35970, At3g26350, At3g11660, At3g44220, At1g08160, At2g01080, At5g06330, At5g56050, At3g20600, NDR1, At3g54200, At3g20590, At4g39740, At1g32270 syntaxin, putative, At1g13050, At5g45320, At3g20610, At4g26490, At5g42860, At1g45688, At4g26820

NDR-NHL genes belong to a large family of which one of the first identified is the defense-associated gene HIN1 (Harpin-induced gene). HIN1 is transcriptionally induced by harpins and bacteria, that elicit hypersensitive responses in tobacco. Other plant species also contain members of this large gene family, such as:

Plant homologs of the Arabidopsis NDR/NHL genes:

BG582276|BG582276|EST484016 GVN Medicago truncata
AV553539|AV553539|Arabidopsis thaliana
AC069325|AC069325|Arabidops
AV526693|AV526693|Arabidopsis thaliana
BG583456|BG583456|EST485208 GVN Medicago truncata

AW267833|AW267833|EST305961 DSIR Medicago truncata
BE997791|BE997791|EST429514 GVSN Medicago truncata
BG580928|BG580928|EST482657 GVN Medicago truncata
BF520916|BF520916|EST458389 DSIL Medicago truncata
AV544651|AV544651|Arabidopsis thaliana
AV543762|AV543762|Arabidopsis thaliana
AW559665|AW559665|EST314777 DSIR Medicago truncata
BG581012|BG581012|EST482741 GVN Medicago truncata
AV552164|AV552164|Arabidopsis thaliana
BE999881|BE999881|EST431604 GVSN Medicago truncata
AW031098|AW031098|EST274405 tomato
AI998763|AI998763|701546833 A. thaliana
AW219286|AW219286|EST301768 tomato
BE124562|BE124562|EST393597 GVN Medicago truncata
AV540371|AV540371|Arabidopsis thaliana
AV539549|AV539549|Arabidopsis thaliana
BG647432|BG647432|EST509051 HOGA Medicago truncata
BE434210|BE434210|EST405288 tomato
BG725849|BG725849|sae42g02.y1 Gm-c1051 Glycine max
AP003247|AP003247|Oryza sativa
BE348073|BE348073|sp11a11.y1 Gm-c1042 Glycine max
AW508383|AW508383|si40c06.y1 Gm-r1030 Glycine max
AI856504|AI856504|sb40b07.y1 Gm-c1014 Glycine max
BE556317|BE556317|sq01b07.y1 Gm-c1045 Glycine max
AA713120|AA713120|32681 Arabidopsis
AV541531|AV541531|Arabidopsis thaliana
AI894456|AI894456|EST263911 tomato
AW704493|AW704493|sk53g11.y1 Gm-c1019 Glycine max
AW219298|AW219298|EST301780 tomato
BF425685|BF425685|ss03c11.y1 Gm-c1047 Glycine max
AV422557|AV422557|Lotus japonicus
BE190816|BE190816|sn79a08.y1 Gm-c1038 Glycine max
BG580331|BG580331|EST482056 GVN Medicago truncata
AV423251|AV423251|Lotus japonicus
AI896088|AI896088|EST265531 tomato
AV413427|AV413427|Lotus japonicus
AV426656|AV426656|Lotus japonicus
AV416256|AV416256|Lotus japonicus
AL385732|CNS0690I|Medicago truncatula
AB016877|AB016877|Arabidopsis thaliana
AV419449|AV419449|Lotus japonicus

AI486269|AI486269|EST244590 tomato
AV411690|AV411690|Lotus japonicus
AV419925|AV419925|Lotus japonicus
AV418222|AV418222|Lotus japonicus
AV409427|AV409427|Lotus japonicus
AC005287|AC005287|Arabidopsis thaliana
AV426716|AV426716|Lotus japonicus
AV411791|AV411791|Lotus japonicus
BG351730|BG351730|131E12 Mature tuber
BG046452|BG046452|saa54b12.y1 Gm-c1060 Glycine max
AI781777|AI781777|EST262656 tomato
BE451428|BE451428|EST402316 tomato
AI772944|AI772944|EST254044 tomato
AI895510|AI895510|EST264953 tomato
AW030762|AW030762|EST274017 tomato
AW218859|AW218859|EST301341 tomato
BE203936|BE203936|EST396612 KV0 Medicago truncata
AV410289|AV410289|Lotus japonicus
AW032019|AW032019|EST275473 tomato
AW030868|AW030868|EST274158 tomato
AV421824|AV421824|Lotus japonicus
BG646408|BG646408|EST508027 HOGA Medicago truncata
AF325013|AF325013|Arabidopsis thaliana
AC007234|AC007234| Arabidops
AW217237|AW217237|EST295951 tomato
AC034257|AC034257|Arabidopsis thaliana
AW625608|AW625608|EST319515 tomato
AW031064|AW031064|EST274371 tomato
AF370332|AF370332|Arabidopsis thaliana
AB006700|AB006700|Arabidopsis thaliana
AW035467|AW035467|EST281205 tomato
AL163812|ATF14F18|Arabidopsis thaliana
AI896652|AI896652|EST266095 tomato
AI730803|AI730803|BNLGHi7970 Cotton
AW034775|AW034775|EST278811 tomato

The invention provides the insight that RKS proteins or functional equivalents thereof play part in a signaling complex (herein also called the RKS signaling complex) comprising molecules of RKS proteins, ELS (Extracellular Like

SERK) proteins, NDR/NHL proteins and SBP/SPL (Squamosa Binding Protein) proteins, and the corresponding protein ligands (see for example table 3) whereby each of these proteins interplay or act in such a way that modifying genes, or modifying expression of genes, encoding ELS, RKS, NDR/NHL or SBP/SPL, proteins or said ligands may lead to functionally equivalent results (Figure 5. Two-hybrid interaction experiments have for example shown *in vitro* interaction between RKS 0 and NDR0/NHL28 and members of the SBP/SPL family. Here we show that *in vivo* the individual components of this signaling complex are regulating identical processes, as based on functional genomics on transgenic plants, overexpressing or co-suppressing single components or combinations of components in this transmembrane signalling complex. ELS proteins are involved in the heterodimerizing complex with the RKS transmembrane receptor at the outer membrane site. ELS molecules are together with RKS molecules involved in the high affinity binding of the ligand. The signal transmitted from the ligand onto the ELS/RKS heterodimerizing protein complex is then transporter over the membrane towards the N-terminal site of RKS protein, located on the other site of the membrane. The activation stage of the RKS molecule is changed, likely as a result of autophosphorylation at specific residues. Subsequently the signal is transmitted to other proteins, one family of such proteins is defined as the SBP/SPL family of transcription factors, the other family of proteins is represented by the NDR/NHL members.

"Functionally equivalent" as used herein is not only used to identify the functional equivalence of otherwise not so homologous genes encoding ELS, RKS, NDR/NHL or SBP/SPL proteins, but also means an equivalent gene or gene product of genes encoding ELS, RKS, NDR/NHL or SBP/SPL proteins in Arabidopsis Thaliana, e.g. identifying a homologue found in nature in other plants or a homologue comprising a deliberate nucleic acid modification, such as a deletion, truncation, insertion, or deliberate codon substitution which may be made on the basis of similarity in polarity, charge, solubility,

hydrophobicity, and/or the amphipathetic nature of the residues as long as the biological activity of the polypeptide is retained. Homology is generally over at least 50% of the full-length of the relevant sequence shown herein. As is well-understood, homology at the amino acid level is generally in terms of amino acid similarity or identity. Similarity allows for "conservative variation", i. e. substitution of one hydrophobic residue such as isoleucine, valine, leucine or methionine for another, or the substitution of one polar residue for another, such as arginine for lysine, glutamic for aspartic acid, or glutamine for asparagine. Deliberate amino acid substitution may be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, and/or the amphipathetic nature of the residues as long as the biological activity of the polypeptide is retained. In a preferred embodiment, all percentage homologies referred to herein refer to percentage sequence identity, e.g. percent (%) amino acid sequence identity with respect to a particular reference sequence can be the percentage of amino acid residues in a candidate sequence that are identical with the amino acid residues in the reference sequence, after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity, without considering any conservative substitutions as part of the sequence identity. Amino acid similarity or identity can be determined by genetic programs known in the art.

'Plant cell', as used herein, amongst others comprises seeds, suspension cultures, embryos, meristematic regions, callous tissues, protoplasts, leaves, roots, shoots, bulbs, gametophytes, sporophytes, pollen and microspores. A target plant to be modified according to the invention may be selected from any monocotyledonous or dicotyledonous plant species, such as for example ornamental plants, vegetables, arable crops etc. 'Dicotyledons' form one of the two divisions of the flowering plants or angiospermae in which the embryo has two or more free or fused cotyledons. 'Monocotyledons' form one of the two divisions of the flowering plants or angiospermae in which the

embryo has one cotyledon. 'Angiospermae' or flowering plants are seed plants characterized by flowers as specialized organs of plant reproduction and by carpels covering the ovaries. Also included are gymnospermae. Gymnospermae are seed plants characterized by strobili as specialized organs for plant reproduction and by naked sporophylls bearing the male or female reproductive organs, for example woody plants. 'Ornamental' plants are plants that are primarily in cultivation for their habitus, special shape, (flower, foliage or otherwise) colour or other characteristics which contribute to human well being indoor as cut flowers or pot plants or outdoors in the man made landscape, for example bulbous plant species like *Tulipa, Freesia, Narcissus, Hyacinthus* etc. 'Vegetables' are plants that are purposely selected or bred for human consumption of foliage, tubers, stems, fruits, flowers or parts of them and that may need an intensive cultivation regime. 'Arable crops' are generally purposely bred or selected for human objectivity's (ranging from direct or indirect consumption, feed or industrial applications such as fibers) for example soybean, sunflower, corn, peanut, maize, wheat, cotton, safflower and rapeseed.

The invention provides a method for modulating a developmental pathway of a plant comprising modifying a gene encoding for a gene product or protein belonging to a developmental cascade or signaling complex comprising modifying at least one gene encoding a gene product belonging to the complex of RKS proteins, ELS proteins, NDR/NHL proteins, SBP/SPL proteins and ligand proteins. In one embodiment, the invention provides a method for modulating or modifying organ size. Plant or plant organ size is determined by both cell elongation and cell division rate. Modifying either one or both processes results in a change in final organ size. Increasing the level of specific members of the family of RKS genes results in an increase in organ size, growth rate and yield. Modulating plant growth, organ size and yield of plant organs is the most important process to be optimized in plant performance. Here we show that modulating the level of members of the family of the RKS signaling complex with a method according to the

invention is sufficient to modulate these processes. The invention provides herewith a method for modulating a developmental pathway of a plant or plant cell comprising modifying a gene or modifying expression of said gene, wherein said gene is encoding a protein belonging to a signaling complex comprising RKS protein, ELS protein, NDR/NHL protein, SBP/SPL protein and RKS/ELS ligand protein allowing modulating cellular division during plant growth or organ formation, in particular wherein said gene comprises an RKS4 or RKS10 gene or functional equivalent thereof. Inactivation of endogenous RKS gene product results in a decrease in plant growth, proving that the normal function of these endogenous RKS gene products is the regulation of growth and organ size. Use of a method according to invention for elevation of the levels of the regulating of the RKS signaling complex in plant cells is provided in order to increase for example the size of plant organs, the growth rate, the yield of harvested crop, the yield of total plant material or the total plant size. Decreasing the levels of endogenous RKS gene product is provided in order to decrease the size of plant organs, the growth rate, or the total plant size.

In another embodiment, the invention relates to cell division. The mitotic cell cycle in eukaryotes determines the total number of cells within the organism and the number of cells within individual organs. The links between cell proliferation, cell differentiation and cell-cycle machinery are of primary importance for eucaryotes, and regulation of these processes allows modifications during every single stage of development. Here we show that modulating the level of members of the family of the RKS signaling complex is sufficient to modulate these processes. The invention provides herewith a method for modulating a developmental pathway of a plant or plant cell comprising modifying a gene or modifying expression of said gene, wherein said gene is encoding a protein belonging to a signaling complex comprising RKS protein, ELS protein, NDR/NHL protein, SBP/SPL protein and RKS/ELS ligand protein allowing modulating cellular division

during plant growth or organ formation, in particular wherein said gene comprises an RKS4 or RKJS 10 gene or functional equivalent Herewith the invention provides a method for modulating the number of cells to be formed within an eukaryotic organism as a whole or for modulating the cell number within individual organs is, which of primary importance in modulating plant developmental processes, especially of arable plants. Here we show that members of the RKS signaling complex are able to regulate the number of cellular divisions, thereby regulating the total number of cells within the organism or different organs.

In a further embodiment, the invention telates to the regeneration of apical meristem. Modification the levels of different RKS and ELS genes within plants allows the initiation and / or outgrowth of apical meristems, resulting in the formation of large numbers of plantlets from a single source. A number of gene products that is able to increase the regeneration potential of plants is known already. Examples of these are KNAT1, cycD3, CUC2 and IPT. Here we show that modulation of the endogenous levels of RKS genes results in the formation of new shoots and plantlets in different plant species like *Nicotiana tabacum* and *Arabidopsis thaliana*. Herewith the invention provides a method for modulating a developmental pathway of a plant or plant cell comprising modifying a gene or modifying expression of said gene, wherein said gene is encoding a protein belonging to a signaling complex comprising RKS protein, ELS protein, NDR/NHL protein, SBP/SPL protein and RKS/ELS ligand protein, allowing modulating apical meristem formation, in particular wherein said gene comprises an ELS1, RKS0, RKS3, RKS4, RKS8 or RKS10 gene or functional equivalent thereof. A direct application of such a method according to the invention is the stable or transient expression of RKS and ELS genes or gene products in order to initiate vegetative reproduction. Regeneration can be induced after overexpression of for example RKS0 and ELS1; or by co-suppression of for example the endogenous RKS3, RKS4,

RKS8 or RKS10 genes. Overexpression or co-suppression of these RKS and ELS gene products can be either transient, or stable by integration of the corresponding expression casettes in the plant genome. A further example of essentially identical functions for for example ELS1 and RKS0 overexpressing plants is for example shown in the detailed description, example 3, where both transgenic constructs are able to induce the regeneration capacity of in vitro cultured *Arabidopsis* callus. Another example comprises functional interaction between RKS and SBP proteins which was shown by studies in transgenic tobacco plants in which SBP5 and RKS0 were both overexpressed under the control of an enhanced 35S promoter. At the tip of double overexpressing plants, embryostructures appeared whereas in the SBP5 overexpressing plants alone or the RKS0 overexpressing plants alone no phenotype was detectable at the root tips of transgenic tobacco plants. These results show that both RKS and SBP proteins are involved together in a signaling cascade, resulting in the reprogramming of developmental fate of a determined meristem.

Furthermore, it is herein also shown that several RKS genes are able to regulate proper identity and development of meristems and primordia. The invention for example also relates to fasciation, Fasciation is normally a result from an increased size of the apical meristem in apical plant organs. Modulation of the number of cells within the proliferating zone of the shoot apical meristem results in an excess number of cellular divisions, giving rise to excess numbers of primordia formed or to stems in which the number of cells is increased. The invention herewith provides a method for modulating a developmental pathway of a plant or plant cell comprising modifying a gene or modifying expression of said gene, wherein said gene is encoding a protein belonging to a signaling complex comprising RKS protein, ELS protein, NDR/NHL protein, SBP/SPL protein and RKS/ELS ligand protein allowing modulating fasciation, in particular wherein said gene comprises an RKS0, RKS3, RKS8 or RKS10 gene or functional equivalent thereof.Here we for example show that modulation of

the levels of RKS gene products in plants like *Arabidopsis thaliana* can result in fasciated stems. A direct application as provided herein is the regulated formation of fasciation in plant species in which such a trait is desired like ornamental plants. Regulation of the initiation and extent of fasciation, either by placing the responsible RKS encoding DNA sequences under the control of stage or tissue specific promoters, constitutive promoters or inducible promoters results in plants with localized or consitutive fasciation of stem tissue. Another application is modulating the number of primordias by regulation of the process of fasciation. An example is provided by for example sprouts, in which an increased number of primordia will result in an increased numbers of sprouts to be harvested. Fasciation can also result in a strong modification in the structural architecture of the influorescence, resulting in a terminal group of flowers resembling the *Umbelliferae* type.

Identical phenotypes can be observed when transgenic plants are produced that contain the NHL10 cDNA under control of an enhanced 35S promoter. The resulting phenotype of the resulting flowers show that flower organ primordia are switched in identity, similar as observed for RKS10 and RKS13. These meristematic identity switches are normally never observed in Arabidopsis and the fact that two different classes of genes are able to display the same phenotypes in transgenic plants is a clear indication for a process in which both members of the RKS and the NDR/NHL families are involved. The invention also relates to root development. Fasciation is normally a result from an increased size of the apical meristem in apical plant organs. Modulation of the number of cells within the proliferating zone of the root apical meristem results in an excess number of cellular divisions, giving rise to excess numbers of primordia formed or to roots in which the number of cells is increased. Adaptation to soil conditions is possible by regulation of root development of plants. Here we describe several processes in root development that can me manipulated by modification of the levels of RKS

signaling complex within the root. The invention provides a method for modulating a developmental pathway of a plant or plant cell comprising modifying a gene or modifying expression of said gene, wherein said gene is encoding a protein belonging to a signaling complex comprising RKS protein, ELS protein, NDR/NHL protein, SBP/SPL protein and RKS/ELS ligand protein allowing modulating root development, in particular wherein said gene comprises an ELS1, ELS2, RKS1, RKS3, RKS4, RKS6, RKS8 or RKS10 gene or functional equivalent thereof. Root length, a result by either root cells proliferation or elongation, can for example be increased by overexpression of for example RKS3, RKS4, RKS6 and ELS2, or inactivation of the endogenous RKS10 gene product. Root length can also be decreased by decreasing of endogenous RKS1 levels or by strong overexpression of RKS10.The initiation of lateral roots is also regulated by RKS gene products. Overexpression of for example RKS10 can result in a strong increase in the initiation and outgrowth of lateral roots. Co-suppression of RKS1 also resulted in the initiation and outgrowth of large numbers of lateral roots. Root hair formation and elongation is important in determining the total contact surface between plant and soil. A strong increase of root hair length (elongation) can be obtained by overexpression of ELS1 and RKS3 gene products. As the roots of terrestrial plants are involved in the acquisition of water and nutrients, anchorage of the plant, synthesis of plant hormones, interaction with the rhizosphere and storage functions, increasing or decreasing root length, for example for flexable adaptations to different water levels, can be manipulated by overexpressing or cosuppressing RKS and / or ELS gene products. Modulation of the total contact surface between plant cells and the outside environment can be manipulated by regulation lateral root formation (increased by RKS10 overexpression and co-suppression of RKS1). Finally the contact surface between plant cells and the soil can be influenced by modulation of the number of root hairs formed or

the elongation of the root hairs, as mediated by ELS1 and RKS3.

In a further embodiment, the invention relates to apical meristem identity. All parts of the plant above the ground are generally the result on one apical shoot meristem that has been initiated early at embryogenesis and that gives rise to all apical organs. This development of a single meristem into complex tissue and repeated patterns is the result of tissue and stage-dependent differentiation processes within the meristems and its resulting offspring cells. The control of meristem formation, meristem identity and meristem differentiation is therefore an important tool in regulating plant architecture and development. Here we present evidence the function of RKS and ELS gene products in regulation of the meristem identity and the formation and outgrowth of new apical meristems. The invention provides a method for modulating a developmental pathway of a plant or plant cell comprising modifying a gene or modifying expression of said gene, wherein said gene is encoding a protein belonging to a signaling complex comprising RKS protein, ELS protein, NDR/NHL protein, SBP/SPL protein and RKS/ELS ligand protein allowing modulating meristem identity, in particular wherein said gene comprises an ELS1, RKS8, RKS10 or RKS13 gene or functional equivalent thereof. Introduction of for example the RKS10 gene product or an other member of the RKS signaling complex under the control of a tissue and / or stage specific promoter as provided herein allows localized and time regulated increases in the levels of gene product. For example the meristematic identity in a determined meristem might thereby be switched back into an indetermined meristem, thereby changing for example a terminal flower into an indetermined generative meristem.

Another application might be found in changing the meristematic identity at an early time point, during early vegetative growth, thereby switching the vegetative meristem into a generative meristem, allowing early flowering. Modulation of meristem identity in terminal primordia, like

for example as shown in Figure 30, where flower organ primordia are converted into terminal flower primordia, allows the formation of completely new types of flowers and fused fruitstructures. Constitutive overexpression of RKS gene products results in plants with many apical meristems, as can clearly been seen in Figure 29, where RKS10 overexpression results in an extremely bushy phenotype.

In another embodiment, the invention relates to male sterility. Male sterility is a highly desired trait in many plant species. For example, manipulation of pollen development is crucial for F1 hybrid seed production, to reduce labour costs and for the production of low-envirnmental impanct genetically engineerded crops. In order to produce hybrid seed from inbred plant lines, the male organs are removed from each flower, and pollen from another parent is applied manually to produce the hybrid seed. This labour-intensive method is used with a number of vegetables (e.g. hybrid tomatoes) and with many ornamental plants. Transgenic appoaches, in which one or more introduced gene products interfere with normal pollen initiation and development is therefore highly desired. Expecially when the number of revertants (growing normal pollen) is extremely low.

Male sterility in plants is a desired trait that has been shown already in many plant species as a result of the inactivation of expression of a number of genes essential for proper stamen develpment, mitotic divisions in the pollen stem cells, or male gametogenesis. A method for modulating a developmental pathway of a plant or plant cell comprising modifying a gene or modifying expression of said gene, wherein said gene is encoding a protein belonging to a signaling complex comprising RKS protein, ELS protein, NDR/NHL protein, SBP/SPL protein and RKS/ELS ligand protein, allowing modulating pollen development, in particular wherein said gene comprises an ELS2 or RKS10 gene or functional equivalent thereof.

Here we present data that show that overexpression of gene products, like transmembrane receptor kinases (RKS) and extracellular proteins (ELS) can also result in the formation of male sterility. The ability to induce male sterility by overexpressing specific genes as provided herein allows the opportunity to produce transgenic overexpressing plants in which the pollen development is inhibited. Stable single copy homozygous integration of such overexpressing traits into the plant genome will render such plants completely sterile, making them excellent material for the production of F1 hybrid seed. Furthermore, the combined integration of a male sterility inducing overexpressing gene coupled directly with another desired transgene result in transgenic plants which are unable to produce transgenic seed, making these transgenic plants excellent material for outside growth without problems affecting transgenic pollen spreading throughout the environment, thereby eliminating possible crosses with wild plant species or other non-transgenic crops. The combination of a desired transgene flanked on both sites by different male-sterility inducing overexpressing genes would decrease the frequency of pollen formation to an extremely low level. An example is an overexpressing construct of RKS10 at the 5'end of integrated DNA fragment, the desired transgene expression casette in the middle and at the 3'end of the integrated DNA the ELS2 overexpressing construct. This complete DNA fragment is integrated into the genome by convential techniques, like particle bombardment, *Agrobacterium* transformation etc. Another possible application concerns the modification of pollen in ornamental plant species like lilly, where the release of pollen from cut flowers can be avoided by making transgenic plants in which pollen development is initiated by release from the stamen is prevented (a desired trait that can be obtained by overexpressing for example ELS2, resulting in partial pollen development). Hereby the ornamental value of the stamen with pollen is not lost, but release of pollen is inhibited.

Furthermore, surprisingly we observe that NDR NHL gene products share homology with the family of syntaxins, involved in vesicle transport, positioning of cell wall formation and cytokinesis.

Table 1

Homology between members of the syntaxin family and the NDR NHL family

NHL10= At2g35980

maaeqplnga fygpsvpppa pkgyyrrghg rgcgcclsl fvkviisliv ilgvaalifw
livrpraikf hvtdasltrf dhtspdnilr ynlaltvpvr npnkriglyy drieahayye
gkrfstitlt pfyqghkntt vltptfqgqn lvifnagqsr tlnaerisgv ynieikfrlr
vrfklgdlkf rrikpkvdcd dlrlplstsn gttttstvfp ikcdfdf

At1g32270 syntaxin,

MVRSNDVKFQ VYDAELTHFD LESNNNLQYS LSLNLSIRNS KSSIGIHYDR FEATVYYMNQ
RLGAVPMPLF YLGSKNTMLL RALFEGQTLV LLKGNERKKF EDDQKTGVYR IDVKLSINFR
VMVLHLVTWP MKPVVRCHLK IPLALGSSNS TGGHKKMLLI GQLVKDTSAN LREASETDHR
RDVAQSKKIA DAKLAKDFEA ALKEFQKAQH ITVERETSYI PFDPKGSFSS SEVDIGYDRS
QEQRVLMESR RQEIVLLDNE ISLNEARIEA REQGIQEVKH QISEVMEMFK DLAVMVDHQG
TIDDIDEKID NLRSAAAQGK SHLVKASNTQ GSNSSLLFSC SLLLFFFLSG DLCRCVCVGS
ENPRLNPTRR KAWCEEEDEE QRKKQQKKKT MSEKRRREEK KVNKPNGFVF CVLGHK*

Below the homology is shown between NHL10 (Upper line) and a syntaxin protein. (bottom line). The identical amino acids are shown in the middle line.

IVRPRAIKFHVTDASLTRFDHTSPDNILRYNLALTVPVRNPNKRIGLYYDRIEAHAYYEG
 VR KF V DA LT FD S N L Y L L RN IG YDR EA YY
MVRSNDVKFQVYDAELTHFDLESNNN-LQYSLSLNLSIRNSKSSIGIHYDRFEATVYYMN

KRFSTITLTPFYQGHKNTTVLTPTFQGQNLVIFNAGQSRTLNAERISGVYNIEIKFRLRV
 R FY G KNT L F GQ LV GVY I K
QRLGAVPMPLFYLGSKNTMLLRALFEGQTLVLLKGNERKKFEDDQKTGVYRIDVKLSINF

RFKLGDLKFRRIKPKVDCDDLRLPLSTSNGTTT
R L KP V C L PL T
RVMVLHLVTWPMKPVVRCH-LKIPLALGSSNST

That syntaxins and NDR/NHL genes share large homology becomes even more clear when performing a database search using the following site:

http://mips.gsf.de/proj/thal/db/search/search_frame.html

searching for homologous sequences with the sequence At1g32270

gene code:	predicted function:
At1g32270 syntaxin, putative	Syntaxin
At5g46860 syntaxin related protein AtVam3p (gb|AAC49823.1)	Syntaxin
At4g17730 syntaxin	Syntaxin
At5g16830 syntaxin homologue	Syntaxin
At3g11650 unknown protein	NDR HNL
At2g35460 similar to harpin-induced protein	NDR HNL
At5g06320 harpin-induced protein-like	NDR HNL
At2g35980 similar to harpin-induced protein	NDR HNL
At1g65690 hypothetical protein	NDR HNL
At4g05220 putative protein	NDR HNL
At3g05710 putative syntaxin protein AtSNAP33	Syntaxin
At2g27080 unknown protein	NDR HNL
At3g52470 putative protein	NDR HNL
At1g61760 hypothetical protein	
At5g21130 putative protein	NDR HNL
At3g52400 syntaxin-like protein synt4	Syntaxin
At2g35960 putative harpin-induced protein	NDR HNL
At5g06330 harpin-induced protein-like	NDR HNL
At5g26980 tSNARE	Syntaxin
At5g36970 putative protein	
At3g44220 putative protein	
At3g03800 s-syntaxin-like protein	Syntaxin
At2g35970 putative harpin-induced protein	NDR HNL
At4g09590 putative protein	
At4g23930 putative protein	

At1g61290	similar to syntaxin-related protein	Syntaxin
At3g11660	unknown protein	
At1g54540	hypothetical protein	
At3g24350	syntaxin-like protein	Syntaxin
At5g22200	NDR1/HIN1-like	NDR HNL
At1g11250	syntaxin-related protein At-SYR1	Syntaxin
At5g53880		
At3g11820	putative syntaxin	Syntaxin
At3g54200		
At5g05760	t-SNARE SED5	Syntaxin
At5g53730		
At4g03330	SYR1-like syntaxin 1	Syntaxin
At3g47910		
At5g08080	syntaxin-like protein	Syntaxin

This observation provides the explanation for understanding the mechanism by which the RKS / NDR-NHL complex functions. Cell wall immobilized RKS gene products (containing the extensin-like extracellular domain) respond to a local ligand signal, in combination with the heterodimerizing ELS protein (s) either as homodimers, as RKS heterodimers or in combination with the heterodimerizing ELS protein(s).

Predicted ligands for the RKS / ELS receptor binding consist of peptide ligands (based on the LRR ligand binding domain of this class of receptors). These ligands are normally produced as a pre pro protein. The N-terminal signal sequence is removed by the transport through the golgi system and allows modification of the ligand at this stage (e.c. glycosylation). The ligands can then be secreted after which further processing is possible (e.c. proteolytic cleavage, removal of sugar groups etc.) The resulting peptide, possible as a monomer or a (hetero)dimerizing molecule binds the transmembrane receptor complex with high affinity, resulting in transmission of the signal from the ligand through the transmembrane receptor component towards the other site of the membrane.

One class of ligands interacting with the RKS and / or ELS receptors consists of the family of pre(pro)proteins shown hereunder in table 3.

Table 3 Ligands within the RKS signaling complex (herein also called RKS/ELS ligand proteins)

At2g13780 MYSKAGMLLL LLHVLGFMLL AILRIKLLVC MFLSLCLLFC SLCWFCLNEW FNNPFGNLLFDVCLVTLGMQ NYLESWFQNL VSF*

At2g18420 MAVFRVLLAS LLISLLVLDF VHADMVRCSL SSRPNLCHRA CGTCCARCNC VAPGTSGNYDKCPCYGSLTT HGGRRKEVKE FSFFTHGS*

At4g09610 MAVFRSTLVL LLIIVCLTTY ELHVHAADGA KVGEGVVKID CGGRCKDRCS KSSRTKLCLRACNSCCSRCN CVPPGTSGNT HLCPCYASIT THGGRLKCP**
= GASA2

At4g09600 MAIFRSTLVL LLILFCLTTF ELHVHAAEDS QVGEGVVKID CGGRCKGRCS KSSRPNLCLRACNSCCYRCN CVPPGTAGNH HLCPCYASIT TRGGRLKCP**
= GASA3

At1g22690 MKKMNVVAFV TLIISFLLLS QVLAELSSSS NNETSSVSQT NDENQTAAFK RTYHHRPRINCGHACARRCS KTSRKKVCHR ACGSCCAKCQ CVPPGTSGNT ASCPCYASIR THGNKLKCP*

At2g39540 MKLVVVQFFI ISLLLTSSFS VLSSADSSCG GKCNVRCSKA GQHEECLKYC NICCQKCNCVPSGTFGHKDE CPCYRDMKNS KGGSKCP*

At3g02885 MANCIRRNAL FFLTLLFLLS VSNLVQAARG GGKLKPQQCN SKCSFRCSAT SHKKPCMFFCLKCCKKCLCV PPGTFGNKQT CPCYNNWKTK EGRPKCP**

At1g74670 MSKEAEYHPE SYGPGSLKSY QCGGQCTRRC SNTKYHKPCM FFCQKCCAKC LCVPPGTYGNKQVCPCYNNW KTQQGGPKCP *

At2g30810 MIYEFREIKF FFLCVYVQGD ELESQAQAPA IHKNGGEGSL KPEECPKACE YRCSATSHRKPCLFFCNKCC NKCLCVPSGT YGHKEECPCY NNWTTKEGGP KCP*

At2g14900 MKIIVSILVL ASLLLISSSL ASATISDAFG SGAVAPAPQS KDGPALEKWC GQKCEGRCKEAGMKDRCLKY CGICCKDCQC VPSGTYGNKH ECACYRDKLS SKGTPKCP*

At5g15230 MAKSYGAIFL LTLIVLFMLQ TMVMASSGSN VKWSQKRYGP GSLKRTQCPS ECDRRCKKTQYHKACITFCN KCCRKCLCVP PGYYGNKQVC SCYNNWKTQE GGPKCP**
= GASA4

At1g75750 MAISKALIAS LLISLLVLQL VQADVENSQK KNGYAKKIDC GSACVARCRL SRRPRLCHRACGTCCYRCNC VPPGTYGNYD KCQCYASLTT HGGRRKCP*

At1g51915 MATERFSTML ISVLVLALVL SPILPCQATR AHLDAETRML RRVCPSCVCC APAPRGACCPCRCPKNP**

At3g15353 MSSNCGSCDC ADKTQCVKKG TSYTFDIVET QESYKEAMIM DVGAEENNAN CKCKCGSSCSCVNCTCCPN**

At1g51920 MASFHSGKSI FLKLFVLFLL LVLPLSQSNA TRIPRAPISS RRPICPACVC CEPAPLGSCCRCCASPIVTQ THHHSQSP*

They consist of a N-terminal signal peptide, followed by a variable hydrophilic domain, probably resulting in a membrane attached (pro)peptide and a conserved cysteine-rich domain. The conserved cysteine domain probably represents the functional peptide ligand. Proteolytic cleavage of this domain from the hydrophilic (transmembrane) domain releases the active ligand and allows functional interaction with the transmembrane receptor complex. The conserved cysteines have conserved positions and can be characterized in the following order:

(.) = any amino acid; (//) number of amino acids; *stopcodon

.....///.....c..c...c.....c...c..cc..c.c.....c.c..

....../..kcp*

Some members of this gene family have been described previously, and represent the GASA family in Arabidopsis thaliana (plant molec. biol. 36 (1998). Similar family members containing the same structural motifs are present in rice (like GASR1) and tomato (Plant Journal 2 (1992) 153-159; Mol. Gen. Genet. 243 (1994) Taylor and Scheuring.

Intracellularly, this signal is transmitted onto membrane (but not necessarily plasma membrane) associated NDR-NHL proteins. At least some of the functions of the syntaxin-like NDR-NHL proteins would thereby result in the regulation of vesicle transport and /or the positioning of new cell wall formation. Neighboring cells are known to influence and determine the developmental state and the differentiation of cells. In transgenic plants with RKS and / or NDR-NHL expression cassettes the positioning of new cell walls is modified, resulting in abnormal neighboring cells, resulting in abnormal development of groups of cells like flower meristem primordia as observed and shown with RKS0, RKS13 and NHL10.

Table 2 overview of accessions numbers of RKS signal complex genes in arabidopsis and in rice:

Gene	code	contig	gene prediction in At database	Oryzo sativa japonica contig	approximate position in bp around:
RKS0	At1g71830	f14o23	ok	OSJNBa0036B21	52.000
RKS1	At1g60800	f8a5	ok	P0038C05	60.000
RKS2	At5g65240	mqn23	ok	OJ1212_C08	8000
RKS3	At5g63710	mbk5	ok	see rks2	
RKS4	At2g23950	t29e15	wrong, exon missing	P0708B04	35.000
RKS5	At5g45780	mra19	wrong, exon missing	OJ1077_A12	102.000
RKS6	At5g10290	wt e 23	ok	see rks2	
RKS7	At5g16000	ku e 24	ok	P0038C05	60.000
RKS8	At1g34210	f23m19	ok	OJ1134_B10	90.000 & 1000 2
different genes!					
RKS10	At4g33430	en d 25	wrong, exon missing	see rks0	
RKS11	At4g30520	wu d 20	wrong, exon missing	see rks4	
RKS12	At2g13800	f13j11	wrong, exon missing	see rks10	
RKS13	At2g13790	f13j11	ok	P0633E08	36.000
RKS14	At3g25560	mwl2	wrong, exon missing	OSJNBb0015G09	36.000
ELS1	At5g21090	ch e 52	ok	P0003H10	53.000
ELS2	possibly allelic variant of ELS1 no genomic sequence identified yet				see els1
ELS3	At3g43740	by c 21	ok	P0468B07	52.000

Homology between aa sequences from arabidopsis proteins are compared with the rice databases using:
http://mips.gsf.de/proj/thal/db/search/search_frame.html
protein sequences based on Oriza sativa japonica contig sequences.

Arabidopsis thaliana ELS1 cDNA

The start codon encoding the first predicted methionine residue of the gene product has been indicated by bold capitals. The first stopcodon has been underlined. Nucleotides predicted to encode protein sequences are in capitals. Leader and tailer sequences are in lowercase letters.

ttactctcaaattccttttcgatttccctctcttaaacctccgaaagctcac
ATGGCGTCTCGAAACTATCGGTGGGAGCTCTTCGCAGCTTCGTTAACCCTAA
CCTTAGCTTTGATTCACCTGGTCGAAGCAAACTCCGAAGGAGATGCTCTCTA
CGCTCTTCGCCGGAGTTTGACAGATCCAGACCATGTCCTCCAGAGCTGGGAT
CCAACTCTTGTTAATCCTTGTACCTGGTTCCATGTCACCTGTAACCAAGACA
ACCGCGTCACTCGTGTGGATTTGGGAAATTCAAACCTCTCTGGACATCTTGC
GCCTGAGCTTGGGAAGCTTGAACATTTACAGTATCTAGAGCTCTACAAAAAC
AACATCCAAGGAACTATACCTTCCGAACTTGGAAATCTGAAGAATCTCATCA
GCTTGGATCTGTACAACAACAATCTTACAGGGATAGTTCCCACTTCTTTGGG
AAAATTGAAGTCTCTGGTCTTTTTACGGCTTAATGACAACCGATTGACGGTC
CAATCCCTAGAGCACTCACGGCAATCCCAAGCCTTTAAAGTTGTGACGTCTC
AAGCAATGATTTGTGTGGACAATCCCACAAACGGACCCTTTGCTCACATTCC
TTTACAGAACTTTGAGAACAACCCGAGATTGGAGGGACCGGAATTACTCGGT
CTTGCAAGCTACGACACTAACTGCACCTGAacaactggcaaaacctgaaaat
gaagaattgggggtgaccttgtaagaacacttcaccactttatcaaatatc
acatctactatgtaataagtatatatatgtagtccaaaaaaaaaaaaaaaaa

Predicted amino acid sequence of the *Arabidopsis thaliana* ELS1 protein.

Different domains are spaced and shown from the N-terminus towards the C-terminus. Overall domain structure is similar as described in Schmidt et al (1997).

At the predicted extracellular domain the first domain represents a signal sequence. The second domain contains a leucine zipper motif, containing 4 leucine residues, each separated by seven other amino acids. The third domain contains conserved cysteine residues, involved in disulphate bridge formation. The fourth domain contains a leucine rich

repeat domain, consisting of 5 complete repeats of each approximately 24 amino acid residues. The last domain might be involved in attachment to other proteins or structures within the cell wall.

MASRNYRWELFAASL
TLTLALIHLVEANSEG

DALYALRRSLTDP
DHVLQSWDPTLVN

PCTWFHVTCNQDNRVTRV

DLGNSNLSGHLA
P ELGKLEHLQYLELYKNNIQGTI
PSELGNLKNLISLDLYNNNLTGIV
PTSLGKLKSLVFLRLNDNRLTGPI
PRALTAIPSLKVVDVSSNDLCGTI
PTNGPFAHIPLQNFENNPRLEGPE

LLGLASYDTNCT

Arabidopsis thaliana ELS2 cDNA
The start codon encoding the first predicted methionine residue
of the gene product has been indicated by bold capitals.
The first stopcodon has been underlined.
Nucleotides predicted to encode protein sequences are in capitals. Leader and tailer sequences are in lowercase letters.

aaaattactcaaattcctattagattactctcttcgacctccgatagctcac
ATGGCGTCTCGAAACTATCGGTGGGAGCTCTTCGCAGCTTCGTTAATCCTAA
CCTTAGCTTTGATTCACCTGGTCGAAGCAAACTCCGAAGGAGATGCTCTTTA
CGCTCTTCGCCGGAGTTTAACAGATCCGGACCATGTCCTCCAGAGCTGGGAT
CCAACTCTTGTTAATCCTTGTACCTGGTTCCATGTCACCTGTAACCAAGACA
ACCGCGTCACTCGTGTGGATTTGGGGAATTCAAACCTCTCTGGACATCTTGC
GCCTGAGCTTGGGAAGCTTGAACATTTACAGTATCTAGAGCTCTACAAAAAC
AACATCCAAGGAACTATACCTTCCGAACTTGGAAATCTGAAGAATCTCATCA
GCTTGGATCTGTACAACAACAATCTTACAGGGATAGTTCCCACTTCTTTGGG
AAAATTGAAGTCTCTGGTCTTTTTACGGCTTAATGACAACCGATTGACGGGG
CAATCCCTAGAGCACTCACTGCCAATCCCAAGCCTTAAAAGTTGTGGATGTC
TAAGCAATGATTTGTGTGGAACAATCCCAACAAACGGACCTTTTGCTCACAT
TCCTTTACAGAACTTTGAGAACAACCCGAGGTTGGAGGGACCGGAATTACTC
GGTCTTGCAAGCTACGACACTAACTGCACCTGAagaaattggcaaaacctga
aaatgaagaattgggggggaccttgtaagaacacttcaccactttatcaaat
atcacatctactatgtaataagtatatatatgtagtccaaaaaaaaaatgaa
gaatcgaatagtaatatcatctggtctcaattgagaactttgaggtctgtgt
atgaaaattaaagattgtactgtaatgttcggttgtgggattctgagaagta
acatttgtattggtatggtatcaagttgttctgccttgtctgcaaaaaaaaa

Predicted amino acid sequence of the *Arabidopsis thaliana* ELS2 protein.
Different domains are spaced and shown from the N-terminus towards the C-terminus. Overall domain structure is similar as described in Schmidt et al (1997).
At the predicted extracellular domain the first domain represents a signal sequence. The second domain contains a leucine zipper motif, containing 4 leucine residues, each separated by seven other amino acids. The third domain

contains conserved cysteine residues, involved in disulphate bridge formation. The fourth domain contains a leucine rich repeat domain, consisting of 5 complete repeats of each approximately 24 amino acid residues. The last domain might be involved in attachment to other proteins or structures within the cell wall.

MASRNYRWELFAASL
ILTLALIHLVEANSEG

DALYALRRSLTDP
DHVLQSWDPTLVN

PCTWFHVTCNQDNRVTRV

DLGNSNLSGHLA
P ELGKLEHLQYLQLYKNNIQGTI
PSELGNLKNLISLDLYNNNLTGIV
PTSLGKLKSLVFLRLNDNRLTGPI
PRALTAIPSLKVVDVSSNDLCGTI
PTNGPFAHIPLQNFENNPRLEGPE

LLGLASYDTNCT

Arabidopsis thaliana ELS3 cDNA
The start codon encoding the first predicted methionine residue
of the gene product has been indicated by bold capitals.
The first stopcodon has been underlined.
Nucleotides predicted to encode protein sequences are in capitals. Leader and tailer sequences are in lowercase letters.

ttctctctccggcgaaaaccATGGTGGCGCAAAACAGTCGGCGGGAGCTTCTAGCAGCTT
CCCTGATCCTAACTTTAGCTCTAATTCGTCTAACGGAAGCAAACTCCGAAGGGGACGCTC
TTCACGCGCTTCGCCGGAGCTTATCAGATCCAGACAATGTTGTTCAGAGTTGGGATCCAA
CTCTTGTTAATCCTTGTACTTGGTTTCATGTCACTTGTAATCAACACCATCAAGTCACTC
GTCTGGATTTGGGGAATTCAAACTTATCTGGACATCTAGTACCTGAACTTGGGAAGCTTG
AACATTTACAATATCTTGAACTCTACAAAAACGAGATTCAAGGAACTATACCTTCTGAGC
TTGGAAATCTGAAGAGTCTAATCAGTTTGGATCTGTACAACAACAATCTCACCGGGAAAA
TCCCATCTTCTTTGGGAAAATTGAAGCGGCTTAACGAAAACCGATTGACCGGTCCTATTC
CTAGAGAACTCACAGTTATTTCAAGCCTTAAAGTTGTTGATGTCTCAGGGAATGATTTGT
GTGGAACAATTCCAGTAGAAGGACCTTTTGAACACATTCCTATGCAAAACTTTGAGAACA
ACCTGAGATTGGAGGGACCAGAACTACTAGGTCTTGCGAGCTATGACACCAATTGCACTT
AAaaagaagttgaagaa

Predicted amino acid sequence of the *Arabidopsis thaliana* ELS3 protein.
Different domains are spaced and shown from the N-terminus towards the C-terminus. Overall domain structure is similar as described in Schmidt et al (1997).
At the predicted extracellular domain the first domain represents a signal sequence. The second domain contains a leucine zipper motif, containing 2 leucine residues, each separated by seven other amino acids. The third domain contains conserved cysteine residues, involved in disulphate bridge formation. The fourth domain contains a leucine rich repeat domain, consisting of 5 complete repeats of each approximately 24 amino acid residues. The last domain might be involved in attachment to other proteins or structures within the cell wall.

MVAQNSRRELLAASL
ILTLALIRLTEANSEG

DALHALRRSLSDP
DNVVQSWDPTLVN

PCTWFHVTCNQHHQVTRL

DLGNSNLSGHLV
P ELGKLEHLQYLELYKNEIQGTI
PSELGNLKSLISLDLYNNNLTGKI
P SSLGKLKRLNENRLTGPI
PRELTVISSLKVVDVSGNDLCGTI
PVEGPFEHIPMQNFENNLRLEGPE

LLGLASYDTNCT

Arabidopsis thaliana RKS0 cDNA

The start codon encoding the first predicted methionine residue

of the gene product has been indicated by bold capitals.

The first stopcodon has been underlined.

Nucleotides predicted to encode protein sequences are in capitals. Leader and tailer sequences are in lowercase letters.

attttattttatttttactctttgtttgttttaatgctaatgggtttttaaaagggtt
atcgaaaaaatgagtgagtttgtgttgaggttgtctctgtaaagtgttaatggtggtgat
tttcggaagttagggttttctcggatctgaagagatcaaatcaagattcgaaatttacca
ttgttgtttgaa**ATG**GAGTCGAGTTATGTGGTGTTTATCTTACTTTCACTGATCTTACTT
CCGAATCATTCACTGTGGCTTGCTTCTGCTAATTTGGAAGGTGATGCTTTGCATACTTTG
AGGGTTACTCTAGTTGATCCAAACAATGTCTTGCAGAGCTGGGATCCTACGCTAGTGAAT
CCTTGCACATGGTTCCATGTCACTTGCAACAACGAGAACAGTGTCATAAGAGTTGATTTG
GGGAATGCAGAGTTATCTGGCCATTTAGTTCCAGAGCTTGGTGTGCTCAAGAATTTGCAG
TATTTGGAGCTTTACAGTAACAACATAACTGGCCCGATTCCTAGTAATCTTGGAAATCTG
ACAAACTTAGTGAGTTTGGATCTTTACTTAAACAGCTTCTCCGGTCCTATTCCGGAATCA
TTGGGAAAGCTTTCAAAGCTGAGATTTCTCCGGCTTAACAACAACAGTCTCACTGGGTCA
ATTCCTATGTCACTGACCAATATTACTACCCTTCAAGTGTTAGATCTATCAAATAACAGA
CTCTCTGGTTCAGTTCCTGACAATGGCTCCTTCTCACTCTTCACACCCATCAGTTTTGCT
AATAACTTAGACCTATGTGGACCTGTTACAAGTCACCCATGTCCTGGATCTCCCCCGTTT
TCTCCTCCACCACCTTTTATTCAACCTCCCCCAGTTTCCACCCCGAGTGGGTATGGTATA
ACTGGAGCAATAGCTGGTGGAGTTGCTGCAGGTGCTGCTTTGCCCTTTGCTGCTCCTGCA
ATAGCCTTTGCTTGGTGGCGACGAAGAAGCCCACTAGATATTTTCTTCGATGTCCCTGCC
GAAGAAGATCCAGAAGTTCATCTGGGACAGCTCAAGAGGTTTTCTTTGCGGGAGCTACAA
GTGGCGAGTGATGGGTTTAGTAACAAGAACATTTTGGGCAGAGGTGGGTTTGGGAAAGTC
TACAAGGGACGCTTGGCAGACGGAACTCTTGTTGCTGTCAAGAGACTGAAGGAAGAGCGA
ACTCCAGGTGGAGAGCTCCAGTTTCAAACAGAAGTAGAGATGATAAGTATGGCAGTTCAT
CGAAACCTGTTGAGATTACGAGGTTTCTGTATGACACCGACCGAGAGATTGCTTGTGTAT
CCTTACATGGCCAATGGAAGTGTTGCTTCGTGTCTCAGAGAGAGGCCACCGTCACAACCT
CCGCTTGATTGGCCAACGCGGAAGAGAATCGCGCTAGGCTCAGCTCGAGGTTTGTCTTAC
CTACATGATCACTGCGATCCGAAGATCATTCACCGTGACGTAAAAGCAGCAAACATCCTC
TTAGACGAAGAATTCGAAGCGGTTGTTGGAGATTTCGGGTTGGCAAAGCTTATGGACTAT
AAAGACACTCACGTGACAACAGCAGTCCGTGGCACCATCGGTCACATCGCTCCAGAATAT
CTCTCAACCGGAAAATCTTCAGAGAAAACCGACGTTTTCGGATACGGAATCATGCTTCTA
GAACTAATCACAGGACAAAGAGCTTTCGATCTCGCTCGGCTAGCTAACGACGACGACGTC
ATGTTACTTGACTGGGTGAAAGGATTGTTGAAGGAGAAGAAGCTAGAGATGTTAGTGGAT
CCAGATCTTCAAACAAACTACGAGGAGAGAGAACTGGAACAAGTGATACAAGTGGCGTTG

CTATGCACGCAAGGATCACCAATGGAAAGACCAAAGATGTCTGAAGTTGTAAGGATGCTG
GAAGGAGATGGGCTTGCGGAGAAATGGGACGAATGGCAAAAAGTTGAGATTTTGAGGGAA
GAGATTGATTTGAGTCCTAATCCTAACTCTGATTGGATTCTTGATTCTACTTACAATTTG
CACGCCGTTGAGTTATCTGGTCCAAGGTAAaaaaaaaaaaaaaaaa

Predicted amino acid sequence of the *Arabidopsis thaliana* RKS0 protein.

Different domains are spaced and shown from the N-terminus towards the C-terminus. Overall domain structure is similar as described in Schmidt et al (1997).

At the predicted extracellular domain the first domain represents a signal sequence. The second domain contains a leucine zipper motif, containing 4 leucine residues, each separated by seven other amino acids. The third domain contains conserved cysteine residues, involved in disulphate bridge formation. The fourth domain contains a leucine rich repeat domain, consisting of 5 complete repeats of each approximately 24 amino acid residues. The fifth domain contains many serine and proline residues, and is likely to contain hydroxy-proline residues, and to be a site for O-glycosylation. The sixth domain contains a single transmembrane domain after which the prodicted intracellular domains are positioned. The seventh domain has an unknown function. The eight domain represents a serine / threonine protein kinase domain (Schmidt et al. 1997) and is probably also containing sequences for protein / protein interactions. The ninth domain has an unknown function. The last and tenth domain at the C-terminal end represents part of a single leucine rich repeat, probably
involved in protein / protein interactions.

MESSYVVFILLSLILLPNHSL
WLASANLEG

DALHTLRVTLVDP
NNVLQSWDPTLVN

PCTWFHVTCNNENSVIRV

DLGNAELSGHLV
P ELGVLKNLQYLELYSNNITGPI
PSNLGNLTNLVSLDLYLNSFSGPI
PESLGKLSKLRFLRLNNNSLTGSI
PMSLTNITTLQVLDLSNNRLSGSV
PDNGSFSLFTPISFANNLDLCGPV

TSHPCPGSPPFSPPPP
FIQPPPVSTPSGYGITG

AIAGGVAAGAAL
PFAAPAIAFAWW

RRRKPLDIFFDVPAEEDPE
VHLGQLKRFSLRELQVAS

DGFSNKNILGRGGFGKVYKGRLAD
GTLVAVKRLKEERTPGGELQFQ
TEVEMISMAVHRNLLRLRGFCM
TPTERLLVYPYMANGSVASCLR
ERPPSQPPLDWPTRKRIALGSA
RGLSYLHDHCDPKIIHRDVKAA
NILLDEEFEAVVGDFGLAKLMD
YKDTHVTTAVRGTIGHIAPEYL
STGKSSEKTDVFGYGIMLLELI
TGQRAFDLARLANDDDVMLLDW
VKGLLKEKKLEMLVDPDLQTNY
EERELEQVIQVALLCTQGSPME
RPKMSEVVRMLE

GDGLAEKWDEWQKVEILREEIDLS

PNPNSDWILDSTYNLHAVELSGPR

Arabidopsis thaliana RKS1 cDNA

The start codon encoding the first predicted methionine residue

of the gene product has been indicated by bold capitals.

The first stopcodon has been underlined.

Nucleotides predicted to encode protein sequences are in capitals. Leader and tailer sequences are in lowercase letters.

ccaaagttgattgctttaagaagggat**ATG**GAAGGTGTGAGATTTGTGGTGTGGAGATTA
GGATTTCTGGTTTTTGTATGGTTCTTTGATATCTCTTCTGCTACACTTTCTCCTACTGGT
GTAAACTATGAAGTGACAGCTTTGGTTGCTGTGAAGAATGAATTGAATGATCCGTACAAA
GTTCTTGAGAATTGGGATGTGAATTCAGTTGATCCTTGTAGCTGGAGAATGGTTTCTTGC
ACTGATGGCTATGTCTCTTCACTGGATCTTCCTAGCCAAAGCTTGTCTGGTACATTGTCT
CCTAGAATCGGAAACCTCACCTATTTACAATCAGTGGTGTTGCAAAACAATGCAATCACT
GGTCCAATTCCGGAAACGATTGGGAGGTTGGAGAAGCTTCAGTCACTTGATCTTTCGAAC
AATTCATTCACCGGGGAGATACCGGCCTCACTTGGAGAACTCAAGAACTTGAATTACTTG
CGGTTAAACAATAACAGTCTTATAGGAACTTGCCCTGAGTCTCTATCCAAGATTGAGGGA
CTCACTCTAGTCGACATTTCGTATAACAATCTTAGTGGTTCGCTGCCAAAAGTTTCTGCC
AGAACTTTCAAGGTAATTGGTAATGCGTTAATCTGTGGCCCAAAAGCTGTTTCAAACTGT
TCTGCTGTTCCCGAGCCTCTCACGCTTCCACAAGATGGTCCAGATGAATCAGGAACTCGT
ACCAATGGCCATCACGTTGCTCTTGCATTTGCCGCAAGCTTCAGTGCAGCATTTTTTGTT
TTCTTTACAAGCGGAATGTTTCTTTGGTGGAGATATCGCCGTAACAAGCAAATATTTTTT
GACGTTAATGAACAATATGATCCAGAAGTGAGTTTAGGGCACTTGAAGAGGTATACATTC
AAAGAGCTTAGATCTGCCACCAATCATTTCAACTCGAAGAACATTCTCGGAAGAGGCGGA
TACGGGATTGTGTACAAAGGACACTTAAACGATGGAACTTTGGTGGCTGTCAAACGTCTC
AAGGACTGTAACATTGCGGGTGGAGAAGTCCAGTTTCAGACAGAAGTAGAGACTATAAGT
TTGGCTCTTCATCGCAATCTCCTCCGGCTCCGCGGTTTCTGTAGTAGCAACCAGGAGAGA
ATTTTAGTCTACCCTTACATGCCAAATGGGAGTGTCGCATCACGCTTAAAAGATAATATC
CGTGGAGAGCCAGCATTAGACTGGTCGAGAAGGAAGAAGATAGCGGTTGGGACAGCGAGA
GGACTAGTTTACCTACACGAGCAATGTGACCCGAAGATTATACACCGCGATGTGAAAGCA
GCTAACATTCTGTTAGATGAGGACTTCGAAGCAGTTGTTGGTGATTTTGGGTTAGCTAAG
CTTCTAGACCATAGAGACTCTCATGTCACAACTGCAGTCCGTGGAACTGTTGGCCACATT
GCACCTGAGTACTTATCCACGGGTCAGTCCTCAGAGAAGACTGATGTCTTTGGCTTTGGC
ATACTTCTCCTTGAGCTCATTACTGGTCAGAAAGCTCTTGATTTTGGCAGATCCGCACAC
CAGAAAGGTGTAATGCTTGACTGGGTGAAGAAGCTGCACCAAGAAGGGAAACTAAAGCAG
TTAATAGACAAAGATCTAAATGACAAGTTCGATAGAGTAGAACTCGAAGAAATCGTTCAA
GTTGCGCTACTCTGCACTCAATTCAATCCATCTCATCGACCGAAAATGTCAGAAGTTATG
AAGATGCTTGAAGGTGACGGTTTGGCTGAGAGATGGGAAGCGACGCAGAACGGTACTGGT
GAGCATCAGCCACCGCCATTGCCACCGGGGATGGTGAGTTCTTCGCCGCGTGTGAGGTAT

TACTCGGATTATATTCAGGAATCGTCTCTTGTAGTAGAAGCCATTGAGCTCTCGGGTCCT
CGATGAttatgactcactgtttttaaaaaa

Predicted amino acid sequence of the *Arabidopsis thaliana* RKS1 protein.

Different domains are spaced and shown from the N-terminus towards the C-terminus. Overall domain structure is similar as described in Schmidt et al (1997).

At the predicted extracellular domain the first domain represents a signal sequence. The second domain contains a leucine zipper motif, containing 3 leucine residues, each separated by seven other amino acids. The third domain contains conserved cysteine residues, involved in disulphate bridge formation. The fourth domain contains a leucine rich repeat domain, consisting of 5 complete repeats of each approximately 24 amino acid residues. The fifth domain contains many serine and proline residues, and is likely to contain hydroxy-proline residues, and to be a site for O-glycosylation. The sixth domain contains a single transmembrane domain after which the prodicted intracellular domains are positioned. The seventh domain has an unknown function. The eight domain represents a serine / threonine protein kinase domain (Schmidt et al. 1997) and is probably also containing sequences for protein / protein interactions. The ninth domain has an unknown function. The last and tenth domain at the C-terminal end represents part of a single leucine rich repeat, probably
involved in protein / protein interactions.

MEGVRFVVWRLGFL
VFVWFFDISSATLSPTGVNYEV

TALVAVKNELNDP
YKVLENWDVNSVD

PCSWRMVSCTDGYVSSL

DLPSQSLSGT

LSPRIGNLTYLQSVLQNNAITGPI

PETIGRLEKLQSLDLSNNSFTGEI

PASLGELKNLNYLRLNNNSLIGTC

PESLSKIEGLTLVDISYNNLSGSL

PKVSARTFK VIGNALICGPK

AVSNCSAVPEPLTL

PQDGPDESGTRTNG

HHVALAFAASFS

AAFFVFFTSGMFLWW

RYRRNKQIFFDVNEQYDPE

VSLGHLKRYTFKELRSAT

NHFNSKNILGRGGYGIVYKGHLND

GTLVAVKRLKDCNIAGGEVQFQ

TEVETISLALHRNLLRLRGFCS

SNQERILVYPYMPNGSVASRLK

DNIRGEPALDWSRRKKIAVGTA

RGLVYLHEQCDPKIIHRDVKAA

NILLDEDFEAVVGDFGLAKLLD

HRDSHVTTAVRGTVGHIAPEYL

STGQSSEKTDVFGFGILLLELI

TGQKALDFGRSAHQKGVMLDW

VKKLHQEGKLKQLIDKDLNDKF

DRVELEEIVQVALLCTQFNPSH

RPKMSEVMKMLE

GDGLAERWEATQNGTGEHQPPPLPPGMVSSS

PRVRYYSDYIQESSLVVEAIELSGPR

Arabidopsis thaliana RKS2 cDNA

The start codon encoding the first predicted methionine residue

of the gene product has been indicated by bold capitals.

The first stopcodon has been underlined.

Nucleotides predicted to encode protein sequences are in capitals. Leader and tailer sequences are in lowercase letters.

Italics indicate the presence of an alternatively spliced gene product.

tcaattttggtagctcttagaaaa**ATG**GCTCTGCTTATTATCACTGCCTTAGTTTTTAGT
AGTTTATGGTCATCTGTGTCACCAGATGCTCAAGGGGATGCATTATTTGCGTTGAGGAGC
TCGTTACGTGCATCTCCTGAACAGCTTAGTGATTGGAACCAGAATCAAGTCGATCCTTGT
ACTTGGTCTCAAGTTATTTGTGATGACAAGAAACATGTTACTTCTGTAACCTTGTCTTAC
ATGAACTTCTCCTCGGGAACACTGTCTTCAGGAATAGGAATCTTGACAACTCTCAAGACT
CTTACATTGAAGGGAAATGGAATAATGGGTGGAATACCAGAATCCATTGGAAATCTGTCT
AGCTTGACCAGCTTAGATTTGGAGGATAATCACTTAACTGATCGCATTCCATCCACTCTC
GGTAATCTCAAGAATCTACAGTTCTTCAGGACCTTGAGTAGGAATAACCTTAATGGTTCT
ATCCCGGATTCACTTACAGGTCTATCAAAACTGATAAATATTCTGCTCGACTCAAATAAT
CTCAGT*GGTGAGATTCCTCAGAGTTTATTCAAAATCCCAAAATACAAT*TTCACAGCAAAC
AACTTGAGCTGTGGTGGCACTTTCCCGCAACCTTGTGTAACCGAGTCCAGTCCTTCAGGT
GATTCAAGCAGTAGAAAAACTGGAATCATCGCTGGAGTTGTTAGCGGAATAGCGGTTATT
CTACTAGGATTCTTCTTCTTTTTCTTCTGCAAGGATAAACATAAAGGATATAAACGAGAC
GTATTTGTGGATGTTGCAGGAACGAACTTTAAAAAAGGTTTGATTTCAGGTGAAGTGGAC
AGAAGGATTGCTTTTGGACAGTTGAGAAGATTTGCATGGAGAGAGCTTCAGTTGGCTACA
GATGAGTTCAGTGAAAAGAATGTTCTCGGACAAGGAGGCTTTGGGAAAGTTTACAAAGGA
TTGCTTTCGGATGGCACCAAAGTCGCTGTAAAAAGATTGACTGATTTTGAACGTCCAGGA
GGAGATGAAGCTTTCCAGAGAGAAGTTGAGATGATAAGTGTAGCTGTTCATAGGAATCTG
CTTCGCCTTATCGGCTTTTGTACAACACAAACTGAACGACTTTTGGTGTATCCTTTCATG
CAGAATCTAAGTGTTGCATATTGCTTAAGAGAGATTAAACCCGGGGATCCAGTTCTGGAT
TGGTTCAGGAGGAAACAGATTGCGTTAGGTGCAGCACGAGGACTCGAATATCTTCATGAA
CATTGCAACCCGAAGATCATACACAGAGATGTGAAAGCTGCAAATGTGTTACTAGATGAA
GACTTTGAAGCAGTGGTTGGTGATTTTGGTTTAGCCAAGTTGGTAGATGTTAGAAGGACT
AATGTAACCACTCAGGTCCGAGGAACAATGGGTCATATTGCACCAGAATGTATATCCACA
GGGAAATCGTCAGAGAAAACCGATGTTTTCGGGTACGGAATTATGCTTCTGGAGCTTGTA
ACTGGACAAAGAGCAATTGATTTCTCGCGGTTAGAGGAAGAAGATGATGTCTTATTGCTA
GACCATGTGAAGAAACTGGAAAGAGAGAAGAGATTAGAAGACATAGTAGATAAGAAGCTT
GATGAGGATTATATAAAGGAAGAAGTTGAAATGATGATACAAGTAGCTCTGCTATGCACA
CAAGCAGCACCGGAAGAACGACCAGCGATGTCGGAAGTAGTAAGAATGCTAGAAGGAGAA

GGGCTTGCAGAGAGATGGGAAGAGTGGCAGAATCTTGAAGTGACGAGACAAGAAGAGTTT
CAGAGGTTGCAGAGGAGATTTGATTGGGGTGAAGATTCCATTAATAATCAAGATGCTATT
GAATTATCTGGTGGAAGATAGaaacaaaaaa

Predicted amino acid sequence of the *Arabidopsis thaliana* RKS2 protein.

Different domains are spaced and shown from the N-terminus towards the C-terminus. Overall domain structure is similar as described in Schmidt et al (1997).

At the predicted extracellular domain the first domain represents a signal sequence. The second domain contains a leucine zipper motif, containing 3 leucine residues, each separated by seven other amino acids. The third domain contains conserved cysteine residues, involved in disulphate bridge formation. The fourth domain contains a leucine rich repeat domain, consisting of 3 complete and 2 incomplete repeats of each approximately 24 amino acid residues. The fifth domain contains many serine and proline residues, and is likely to contain hydroxy-proline residues, and to be a site for O-glycosylation. The sixth domain contains a single transmembrane domain after which the prodicted intracellular domains are positioned. The seventh domain has an unknown function. The eight domain represents a serine / threonine protein kinase domain (Schmidt et al. 1997) and is probably also containing sequences for protein / protein interactions. The ninth domain has an unknown function. The last and tenth domain at the C-terminal end represents part of a single leucine rich repeat, probably involved in protein / protein interactions. Italics indicate an alternatively spliced gene product.

MALLIITALVFSSL
WSSVSPDAQG

DALFALRSSLR
ASPEQLSDWNQNQVD

PCTWSQVICDDKKHVTSV

TLSYMNFSS GTLSSGI
G ILTTLKTLTLKGNGIMGGI
PESIGNLSSLTSLDLEDNHLTDRI
PSTLGNLKNLQFLTLSRNNLNGSI
PDSLTGLSKLINILLDSNNLSGEI
PQSLFKIPKYN FTANNLSCGG

TFPQPCVTESSPSGDSSSRKTG

IIAGVVSGIAVIL
LGFFFFFFC

KDKHKGYKRDVFVDVAGTNFKKGLISGE
VDRRIAFGQLRRFAWRELQLAT

DEFSEKNVLGQGGFGKVYKGLLSD
GTKVAVKRLTDFERPGGDEAFQ
REVEMISVAVHRNLLRLIGFCT
TQTERLLVYPFMQNLSVAYCLR
EIKPGDPVLDWFRRKQIALGAA
RGLEYLHEHCNPKIIHRDVKAA
NVLLDEDFEAVVGDFGLAKLVD
VRRTNVTTQVRGTMGHIAPECI
STGKSSEKTDVFGYGIMLLELV
TGQRAIDFSRLEEEDDVLLLDH
VKKLEREKRLEDIVDKKLDEDY
IKEEVEMMIQVALLCTQAAPEE
RPAMSEVVRMLE

GEGLAERWEEWQNLEVTRQEEFQ

RLQRRFDWGEDSINNQDAIELSGGR

Arabidopsis thaliana RKS3 cDNA

The start codon encoding the first predicted methionine residue

of the gene product has been indicated by bold capitals.

The first stopcodon has been underlined.

Nucleotides predicted to encode protein sequences are in capitals. Leader and tailer sequences are in lowercase letters.

aacggtgaaagtttccatgatcctcttcgaggattcattcaaagaaattgctttagatgg

aacaatcagaaattgatcttacaatgtttc**ATG**GCCTTAGCTTTTGTGGGAATCACTTCG

TCAACAACTCAACCAGATATCGAAGGAGGAGCTCTGTTGCAGCTCAGAGATTCGCTTAAT..

GATTCGAGCAATCGTCTAAAATGGACACGCGATTTTGTGAGCCCTTGCTATAGTTGGTCT

TATGTTACCTGCAGAGGCCAGAGTGTTGTGGCTCTAAATCTTGCCTCGAGTGGATTCACA

GGAACACTCTCTCCAGCTATTACAAAACTGAAGTTCTTGGTTACCTTAGAGTTACAGAAC

AATAGTTTATCTGGTGCCTTACCAGATTCTCTTGGGAACATGGTTAATCTACAGACTTTA

AACCTATCAGTGAATAGTTTCAGCGGATCGATACCAGCGAGCTGGAGTCAGCTCTCGAAT

CTAAAGCACTTGGATCTCTCATCCAATAATTTAACAGGAAGCATCCCAACACAATTCTTC

TCAATCCCAACATTCGATTTTTCAGGAACTCAGCTTATATGCGGTAAAAGTTTGAATCAG

CCTTGTTCTTCAAGTTCTCGTCTTCCAGTCACATCCTCCAAGAAAAAGCTGAGAGACATT

ACTTTGACTGCAAGTTGTGTTGCTTCTATAATCTTATTCCTTGGAGCAATGGTTATGTAT

CATCACCATCGCGTCCGCAGAACCAAATACGACATCTTTTTTGATGTAGCTGGGGAAGAT

GACAGGAAGATTTCCTTTGGACAACTAAAACGATTCTCTTTACGTGAAATCCAGCTCGCA

ACAGATAGTTTCAACGAGAGCAATTTGATAGGACAAGGAGGATTTGGTAAAGTATACAGA

GGTTTGCTTCCAGACAAAACAAAAGTTGCAGTGAAACGCCTTGCGGATTACTTCAGTCCT

GGAGGAGAAGCTGCTTTCCAAAGAGAGATTCAGCTCATAAGCGTTGCGGTTCATAAAAAT

CTCTTACGCCTTATTGGCTTCTGCACAACTTCCTCTGAGAGAATCCTTGTTTATCCATAC

ATGGAAAATCTTAGTGTTGCATATCGACTAAGAGATTTGAAAGCGGGAGAGGAAGGATTA

GACTGGCCAACAAGGAAGCGTGTAGCTTTTGGTTCAGCTCACGGTTTAGAGTATCTACAC

GAACATTGTAACCCGAAGATCATACACCGCGATCTCAAGGCTGCAAACATACTTTTAGAC

AACAATTTTGAGCCAGTTCTTGGAGATTTCGGTTTAGCTAAGCTTGTGGACACATCTCTG

ACTCATGTCACAACTCAAGTCCGAGGCACAATGGGTCACATTGCGCCAGAGTATCTCTGC

ACAGGAAAATCATCTGAAAAAACCGATGTTTTTGGTTACGGTATAACGCTTCTTGAGCTT

GTTACTGGTCAGCGCGCAATCGATTTTTCACGCTTGGAAGAAGAGGAAAATATTCTCTTG

CTTGATCATATAAAGAAGTTGCTTAGAGAACAGAGACTTAGAGACATTGTTGATAGCAAT

TTGACTACATATGACTCCAAAGAAGTTGAAACAATCGTTCAAGTGGCTCTTCTCTGCACA

CAAGGCTCACCAGAAGATAGACCAGCGATGTCTGAAGTGGTCAAAATGCTTCAAGGGACT

GGTGGTTTGGCTGAGAAATGGACTGAATGGGAACAACTTGAAGAAGTTAGGAACAAAGAA

GCATTGTTGCTTCCGACTTTACCGGCTACTTGGGATGAAGAAGAAACCACCGTTGATCAA

GAATCTATCCGATTATCGACAGCAAGA<u>TGA</u>agaagaaacagagagagaaagatatctatg

aaaa

Predicted amino acid sequence of the *Arabidopsis thaliana* RKS3 protein.

Different domains are spaced and shown from the N-terminus towards the C-terminus. Overall domain structure is similar as described in Schmidt et al (1997).

At the predicted extracellular domain the first domain represents a signal sequence. The second domain contains a leucine zipper motif, containing 3 leucine residues, each separated by seven other amino acids. The third domain contains conserved cysteine residues, involved in disulphate bridge formation. The fourth domain contains a leucine rich repeat domain, consisting of 4 complete repeats of each approximately 24 amino acid residues. The fifth domain contains many serine and proline residues, and is likely to contain hydroxy-proline residues, and to be a site for O-glycosylation. The sixth domain contains a single transmembrane domain after which the prodicted intracellular domains are positioned. The seventh domain has an unknown function. The eight domain represents a serine / threonine protein kinase domain (Schmidt et al. 1997) and is probably also containing sequences for protein / protein interactions. The ninth domain has an unknown function. The last and tenth domain at the C-terminal end represents part of a single leucine rich repeat, probably
involved in protein / protein interactions.

MALAFVGITSSTTQPDIEG

GALLQLRDSLNDSSNRL
KWTRDFVS

PCYSWSYVTCRGQSVVAL

NLASSGFTGTLS
P AITKLKFLVTLELQNNSLSGAL

PDSLGNMVNLQTLNLSVNSFSGSI

PASWSQLSNLKHLDLSSNNLTGSI

PTQFFSIPTFEFSGTQLICGKS

LNQPCSSSRLPVTSSKKKLRD

ITLTASCVASIIL

FLGAMVMYHHH

RVRRTKYDIFFDVAGEDDR

KISFGQLKRFSLREIQLAT

DSFNESNLIGQGGFGKVYRGLLPD

KTKVAVKRLADYFSPGGEAAFQ

REIQLISVAVHKNLLRLIGFCT

TSSERILVYPYMENLSVAYRLR

DLKAGEEGLDWPTRKRVAFGSA

HGLEYLHEHCNPKIIHRDLKAA

NILLDNNFEPVLGDFGLAKLVD

TSLTHVTTQVRGTMGHIAPEYL

CTGKSSEKTDVFGYGITLLELV

TGQRAIDFSRLEEEENILLLD

HIKKLLREQRLRDIVDSNLTTY

DSKEVETIVQVALLCTQGSPED

RPAMSEVVKMLQ

GTGGLAEKWTEWEQLEEVRNKEALLL

PTLPATWDEEETTVDQESIRLSTAR

Arabidopsis thaliana RKS4 cDNA

The start codon encoding the first predicted methionine residue

of the gene product has been indicated by bold capitals.

The first stopcodon has been underlined.

Nucleotides predicted to encode protein sequences are in capitals. Leader and tailer sequences are in lowercase letters.

tcttccttctccttctggtaatctaatctaaagcttttcATGGTGGTGATGAAGATATTC
TCTGTTCTGTTACTACTATGTTTCTTCGTTACTTGTTCTCTCTCTTCTGAACCCAGAAAC
CCTGAAGTGGAGGCGTTGATAAACATAAAGAACGAGTTACATGATCCACATGGTGTTTTC
AAAAACTGGGATGAGTTTTCTGTTGATCCTTGTAGCTGGACTATGATCTCTTGTTCTTCA
GACAACCTCGTAATTGGCTTAGGAGCTCCAAGTCAGTCTCTTTCAGGAACTTTATCTGGG
TCTATTGGAAATCTCACTAATCTTCGACAAGTGTCATTACAGAACAATAACATCTCCGGT
AAAATCCCACCGGAGATTTGTTCTCTTCCCAAATTACAGACTCTGGATTTATCCAATAAC
CGGTTCTCCGGTGAAATCCCCGGTTCTGTTAACCAGCTGAGTAATCTCCAATATCTGTTG
AACAACAACTCATTATCTGGGCCCTTTCCTGCTTCTCTGTCTCAAATCCCTCACCTCTCT
TTCTTAGACTTGTCTTATAACAATCTCAGAGGTCCTGTTCCTAAATTTCCTGCAAGGACA
TTCAATGTTGCTGGGAACCCTTTGATTTGTAAAAACAGCCTACCGGAGATTTGTTCAGGA
TCAATCAGTGCAAGCCCTCTTTCTGTCTCTTTACGTTCTTCATCAGGACGTAGAACCAAC
ATATTAGCAGTTGCACTTGGTGTAAGCCTTGGCTTTGCTGTTAGTGTAATCCTCTCTCTC
GGGTTCATTTGGTATCGAAAGAAACAAAGACGGTTAACGATGCTTCGCATTAACAAGCAA
GAGGAAGGGTTACTTGGGTTGGGAAATCTAAGAAGCTTCACATTCAGGGAACTTCATGTA
GCTACGGATGGTTTTAGTTCCAAGAGTATTCTTGGTGCTGGTGGGTTTGGTAATGTCTAC
AGAGGAAAATTCGGGGATGGGACAGTGGTTGCAGTGAAACGATTGAAAGATGTGAATGGA
ACCTCCGGGAACTCACAGTTTCGTACTGAGCTTGAGATGATCAGCTTAGCTGTTCATAGG
AATTTGCTTCGGTTAATCGGTTATTGTGCGAGTTCTAGCGAAAGACTTCTTGTTTACCCT
TACATGTCCAATGGCAGCGTCGCCTCTAGGCTCAAAGCTAAGCCAGCGTTGGACTGGAAC
ACAAGGAAGAAGATAGCGATTGGAGCTGCAAGAGGGTTGTTTTATCTACACGAGCAATGC
GATCCCAAGATTATTCACCGAGATGTCAAGGCAGCAAACATTCTCCTAGATGAGTATTTT
GAAGCAGTTGTTGGGGATTTTGGACTAGCAAAGCTACTCAACCACGAGGATTCACATGTC
ACAACCGCGGTTAGAGGAACTGTTGGTCACATTGCACCTGAGTATCTCTCCACCGGTCAG
TCATCTGAGAAAACCGATGTCTTTGGGTTCGGTATACTTTTGCTAGAGCTCATCACAGGA
ATGAGAGCTCTCGAGTTTGGCAAGTCTGTTAGCCAGAAAGGAGCTATGCTAGAATGGGTG
AGGAAGCTACACAAGGAAATGAAAGTAGAGGAGCTAGTAGACCGAGAACTGGGGACAACC
TACGATAGAATAGAAGTTGGAGAGATGCTACAAGTGGCACTGCTCTGCACTCAGTTTCTT
CCAGCTCACAGACCCAAAATGTCTGAAGTAGTTCAGATGCTTGAAGGAGATGGATTAGCT
GAGAGATGGGCTGCTTCACATGACCATTCACATTTCTACCATGCCAACATGTCTTACAGG
ACTATTACCTCTACTGATGGCAACAACCAAACCAAACATCTGTTTGGCTCCTCAGGATTT

GAAGATGAAGATGATAATCAAGCGTTAGATTCATTCGCCATGGAACTATCTGGTCCAAGG
TAGtaaatcttggacacagaaagaaacagatataatatccccatgacttcaatttttgtt

Predicted amino acid sequence of the *Arabidopsis thaliana* RKS4 protein.

Different domains are spaced and shown from the N-terminus towards the C-terminus. Overall domain structure is similar as described in Schmidt et al (1997).

At the predicted extracellular domain the first domain represents a signal sequence. The second domain contains a leucine zipper motif, containing 2 leucine residues, each separated by seven other amino acids. The third domain contains conserved cysteine residues, involved in disulphate bridge formation. The fourth domain contains a leucine rich repeat domain, consisting of 5 complete repeats of each approximately 24 amino acid residues. The fifth domain contains many serine and proline residues, and is likely to contain hydroxy-proline residues, and to be a site for O-glycosylation. The sixth domain contains a single transmembrane domain after which the prodicted intracellular domains are positioned. The seventh domain has an unknown function. The eight domain represents a serine / threonine protein kinase domain (Schmidt et al. 1997) and is probably also containing sequences for protein / protein interactions. The ninth domain has an unknown function. The last and tenth domain at the C-terminal end represents part of a single leucine rich repeat, probably
involved in protein / protein interactions.

MVVMKLITMKIFSVLLLL
CFFVTCSLSSEPRNPEV

EALINIKNELHDP
HGVFKNWDEFSVD

PCSWTMISCSSDNLVIGL

GAPSQSLSGTLS

G SIGNLTNLRQVSLQNNNISGKI

PPEICSLPKLQTLDLSNNRFSGEI

PGSVNQLSNLQYLRLNNNSLSGPF

PASLSQIPHLSFLDLSYNNLRGPV

PKFPARTFNVAGNPLICKNS

LPEICSGSISASPL

SVSLRSSSGRRTN

ILAVALGVSLGFAVSVIL

SLGFIWY

RKKQRRLTMLRINKQEE

GLLGLGNLRSFTFRELHVAT

DGFSSKSILGAGGFGNVYRGKFGD

GTVVAVKRLKDVNGTSGNSQFR

TELEMISLAVHRNLLRLIGYCA

SSSERLLVYPYMSNGSVASRLK

AKPALDWNTRKKIAIGAA

RGLFYLHEQCDPKIIHRDVKAA

NILLDEYFEAVVGDFGLAKLLN

HEDSHVTTAVRGTVGHIAPEYL

STGQSSEKTDVFGFGILLLELI

TGMRALEFGKSVSQKGAMLEW

VRKLHKEMKVEELVDRELGTTY

DRIEVGEMLQVALLCTQFLPAH

RPKMSEVVQMLE

GDGLAERWAASHDHSHFYHANM

SYRTITSTDGNNQTKHLFG

SSGFEDEDDNQALDSFAMELSGPR

Arabidopsis thaliana RKS5 cDNA

The start codon encoding the first predicted methionine residue

of the gene product has been indicated by bold capitals.

The first stopcodon has been underlined.

Nucleotides predicted to encode protein sequences are in capitals. Leader and tailer sequences are in lowercase letters.

ctagagaattcttatacttttctacgATGGAGATTTCTTTGATGAAGTTTCTGTTTTTA
GGAATCTGGGTTTATTATTACTCTGTTCTTGACTCTGTTTCTGCCATGGATAGTCTTTTA
TCTCCCAAGGTGGCTGCGTTAATGTCAGTGAAGAACAAGATGAAAGATGAGAAAGAGGTT
TTGTCTGGTTGGGATATTAACTCTGTTGATCCTTGTACTTGGAACATGGTTGGTTGTTCT
TCTGAAGGTTTTGTGGTTTCTCTAGAGATGGCTAGTAAAGGATTATCAGGGATACTATCT
ACTAGTATTGGGGAATTAACTCATCTTCATACTTTGTTACTTCAGAATAATCAGTTAACT
GGTCCGATTCCTTCTGAGTTAGGCCAACTCTCTGAGCTTGAAACGCTTGATTTATCGGGG
AATCGGTTTAGTGGTGAAATCCCAGCTTCTTTAGGGTTCTTAACTCACTTAAACTACTTG
CGGCTTAGCAGGAATCTTTTATCTGGGCAAGTCCCTCACCTCGTCGCTGGCCTCTCAGGT
CTTTCTTTCTTGGATCTATCTTTCAACAATCTAAGCGGACCAACTCCGAATATATCAGCA
AAAGATTACAGGAAATGCATTTCTTTGTGGTCCAGCTTCCCAAGAGCTTTGCTCAGATGC
TACACCTGTGAGAAATGCTGCAATCGATCTGCAGCGACGGGTTTGTCTGAAAAGGACAAT
AGCAAACATCACAGCTTAGTGCTCTCTTTTGCATTTGGCATTGTTGTTGCCTTTATCATC
TCCCTAATGTTTCTCTTCTTCTGGGTGCTTTGGCATCGATCACGTCTCTCAAGATCACAC
GTGCAGCAAGACTACGAATTTGAAATCGGCCATCTGAAAAGGTTCAGTTTTCGCGAAATA
CAAACCGCAACAAGCAATTTTAGTCCAAAGAACATTTTGGGACAAGGAGGGTTTGGGATG
GTTTATAAAGGGTATCTCCCAAATGGAACTGTGGTGGCAGTTAAAAGATTGAAAGATCCG
ATTTATACAGGAGAAGTTCAGTTTCAAACCGAAGTAGAGATGATTGGCTTAGCTGTTCAC
CGTAACCTTTTACGCCTCTTTGGATTCTGTATGACCCCGGAAGAGAGAATGCTTGTGTAT
CCGTACATGCCAAATGGAAGCGTAGCTGATCGTCTGAGAGATTGGAATCGGAGGATAAGC
ATTGCACTCGGCGCAGCTCGAGGACTTGTTTACTTGCACGAGCAATGCAATCCAAAGATT
ATTCACAGAGACGTCAAAGCTGCAAATATTCTACTTGATGAGAGCTTTGAAGCAATAGTT
GGCGATTTTGGTCTAGCAAAGCTTTTAGACCAGAGAGATTCACATGTCACTACCGCAGTC
CGAGGAACCATTGGACACATCGCTCCCGAGTACCTTTCCACTGGACAGTCCTCAGAGAAA
ACCGATGTTTTCGGATTCGGAGTACTAATCCTTGAACTCATAACAGGTCATAAGATGATT
GATCAAGGCAATGGTCAAGTTCGAAAAGGAATGATATTGAGCTGGGTAAGGACATTGAAA
GCAGAGAAGAGATTTGCAGAGATGGTGGACAGAGATTTGAAGGGAGAGTTTGATGATTTG
GTGTTGGAGGAAGTAGTGGAATTGGCTTTGCTTTGTACACAGCCACATCCGAATCTAAGA
CCGAGGATGTCTCAAGTGTTGAAGGTACTAGAAGGTTTAGTGGAACAGTGTGAAGGAGGG
TATGAAGCTAGAGCTCCAAGTGTCTCTAGGAACTACAGTAATGGTCATGAAGAGCAGTCC
TTTATTATTGAAGCCATTGAGCTCTCTGGACCACGATGAtagacttcatagtgtcttaac

tagtcttcttgattttgttgtcattgtcatggc

Predicted amino acid sequence of the *Arabidopsis thaliana* RKS5 protein.

Different domains are spaced and shown from the N-terminus towards the C-terminus. Overall domain structure is similar as described in Schmidt et al (1997).

At the predicted extracellular domain the first domain represents a signal sequence. The second domain contains no leucine zipper motif, in contrast to the other RKS proteins. The third domain contains conserved cysteine residues, involved in disulphate bridge formation. The fourth domain contains a leucine rich repeat domain, consisting of 5 complete repeats of each approximately 24 amino acid residues. The fifth domain contains many serine residues, and is likely to be a site for O-glycosylation. The sixth domain contains a single transmembrane domain after which the prodicted intracellular domains are positioned. The seventh domain has an unknown function. The eight domain represents a serine / threonine protein kinase domain (Schmidt et al. 1997) and is probably also containing sequences for protein / protein interactions. The ninth domain has an unknown function. The last and tenth domain at the C-terminal end represents part of a single leucine rich repeat, probably involved in protein / protein interactions.

MEISLMKFLFLGIWVYYYS
VLDSVSAMDSLLSPKV

AALMSVKNKMKDE
KEVLSGWDINSVD

PCTWNMVGCSSEGFVVS

LEMASKGLSGILS
T SIGELTHLHTLLLQNNQLTGPI
PSELGQLSELETLDLSGNRFSGEI
PASLGFLTHLNYLRLSRNLLSGQV

PHLVAGLSGLSFLDLSFNNLSGPT

PNISAK DYRKCISLWSSFPR

ALLRCYTCEKCCNR

SAATGLSEKDNSK

HHSLVLSFAFGIVV

AFIISLMFLFFWVLWH

RSRLSRSHVQQDYEF

EIGHLKRFSFREIQTAT

SNFSPKNILGQGGFGMVYKGYLPN

GTVVAVKRLKDPIYTGEVQFQ

TEVEMIGLAVHRNLLRLFGFCM

TPEERMLVYPYMPNGSVADRLR

DWNRRISIALGAA

RGLVYLHEQCNPKIIHRDVKAA

NILLDESFEAIVGDFGLAKLLD

QRDSHVTTAVRGTIGHIAPEYL

STGQSSEKTDVFGFGVLILELI

TGHKMIDQGNGQVRKGMILSW

VRTLKAEKRFAEMVDRDLKGEF

DDLVLEEVVELALLCTQPHPNL

RPRMSQVLKV

LEGLVEQCEGGYEARA

PASVSRNYSNGHEEQSFIIEAIELSGPR

Arabidopsis thaliana RKS6 cDNA
The start codon encoding the first predicted methionine residue
of the gene product has been indicated by bold capitals.
The first stopcodon has been underlined.
Nucleotides predicted to encode protein sequences are in capitals. Leader and tailer sequences are in lowercase letters.

attgtttccttcttttgggattttctccttggatggaaccagctcaattaatgagatgag
ATGAGAATGTTCAGCTTGCAGAAGATGGCTATGGCTTTTACTCTCTTGTTTTTTGCCTGT
TTATGCTCATTTGTGTCTCCAGATGCTCAAGGGGATGCACTGTTTGCGTTGAGGATCTCC
TTACGTGCATTACCGAATCAGCTAAGTGACTGGAATCAGAACCAAGTTAATCCTTGCACT
TGGTCCCAAGTTATTTGTGATGACAAAAACTTTGTCACTTCTCTTACATTGTCAGATATG
AACTTCTCGGGAACCTTGTCTTCAAGAGTAGGAATCCTAGAAAATCTCAAGACTCTTACT
TTAAAGGGAAATGGAATTACGGGTGAAATACCAGAAGACTTTGGAAATCTGACTAGCTTG
ACTAGTTTGGATTTGGAGGACAATCAGCTAACTGGTCGTATACCATCCACTATCGGTAAT
CTCAAGAAACTTCAGTTCTTGACCTTGAGTAGGAACAAACTTAATGGGACTATTCCGGAG
TCACTCACTGGTCTTCCAAACCTGTTAAACCTGCTGCTTGATTCCAATAGTCTCAGTGGT
CAGATTCCTCAAAGTCTGTTTGAGATCCCAAAATATAATTTCACGTCAAACAACTTGAAT
TGTGGCGGTCGTCAACCTCACCCTTGTGTATCCGCGGTTGCCCATTCAGGTGATTCAAGC
AAGCCTAAAACTGGCATTATTGCTGGAGTTGTTGCTGGAGTTACAGTTGTTCTCTTTGGA
ATCTTGTTGTTTCTGTTCTGCAAGGATAGGCATAAAGGATATAGACGTGATGTGTTTGTG
GATGTTGCAGGTGAAGTGGACAGGAGAATTGCATTTGGACAGTTGAAAAGGTTTGCATGG
AGAGAGCTCCAGTTAGCGACAGATAACTTCAGCGAAAAGAATGTACTTGGTCAAGGAGGC
TTTGGGAAAGTTTACAAAGGAGTGCTTCCGGATACACCCAAAGTTGCTGTGAAGAGATTG
ACGGATTTCGAAAGTCCTGGTGGAGATGCTGCTTTCCAAAGGGAAGTAGAGATGATAAGT
GTAGCTGTTCATAGGAATCTACTCCGTCTTATCGGGTTCTGCACCACACAAACAGAACGC
CTTTTGGTTTATCCCTTCATGCAGAATCTAAGTCTTGCACATCGTCTGAGAGAGATCAAA
GCAGGCGACCCGGTTCTAGATTGGGAGACGAGGAAACGGATTGCCTTAGGAGCAGCGCGT
GGTTTTGAGTATCTTCATGAACATTGCAATCCGAAGATCATACATCGTGATGTGAAAGCA
GCTAATGTGTTACTAGATGAAGATTTTGAAGCAGTGGTTGGTGATTTTGGTTTAGCCAAG
CTAGTAGATGTTAGAAGGACTAATGTGACTACTCAAGTTCGAGGAACAATGGGTCACATT
GCACCAGAATATTTATCAACAGGGAAATCATCAGAGAGAACCGATGTTTTCGGGTATGGA
ATTATGCTTCTTGAGCTTGTTACAGGACAACGCGCAATAGACTTTTCACGTTTGGAGGAA
GAAGATGATGTCTTGTTACTTGACCACGTGAAGAAACTGGAAAGAGAGAAGAGATTAGGA
GCAATCGTAGATAAGAATTTGGATGGAGAGTATATAAAAGAAGAAGTAGAGATGATGATA
CAAGTGGCTTTGCTTTGTACACAAGGTTCACCAGAAGACCGACCAGTGATGTCTGAAGTT
GTGAGGATGTTAGAAGGAGAAGGGCTTGCGGAGAGATGGGAAGAGTGGCAAAACGTGGAA
GTCACGAGACGTCATGAGTTTGAACGGTTGCAGAGGAGATTTGATTGGGGTGAAGATTCT

ATGCATAACCAAGATGCCATTGAATTATCTGGTGGAAGATGAccaaaaaacatcaaaccttt

Predicted amino acid sequence of the *Arabidopsis thaliana* RKS6 protein.

Different domains are spaced and shown from the N-terminus towards the C-terminus. Overall domain structure is similar as described in Schmidt et al (1997).

At the predicted extracellular domain the first domain represents a signal sequence. The second domain contains a leucine zipper motif, containing 3 leucine residues, each separated by seven other amino acids. The third domain contains conserved cysteine residues, involved in disulphate bridge formation. The fourth domain contains a leucine rich repeat domain, consisting of 5 complete repeats of each approximately 24 amino acid residues. The fifth domain contains many serine and proline residues, and is likely to contain hydroxy-proline residues, and to be a site for O-glycosylation. The sixth domain contains a single transmembrane domain after which the prodicted intracellular domains are positioned. The seventh domain has an unknown function. The eight domain represents a serine / threonine protein kinase domain (Schmidt et al. 1997) and is probably also containing sequences for protein / protein interactions. The ninth domain has an unknown function. The last and tenth domain at the C-terminal end represents part of a single leucine rich repeat, probably
involved in protein / protein interactions.

MRMFSL

QKMAMAFTLLFFACLCSFVSPDAQG

DALFALRISLRALP

NQLSDWNQNQVN

PCTWSQVICDDKNFVTSL

TLSDMNFSGTLSSRV

GILENLKTLTLKGNGITGEI

PEDFGNLTSLTSLDLEDNQLTGRI
PSTIGNLKKLQFLTLSRNKLNGTI
PESLTGLPNLLNLLLDSNSLSGQI
PQSLFEIPKYNFTSNNLNCGG

RQPHPCVSAVAHSGDSSKPKTG

IIAGVVAGVTVVL
FGILLFLFC

KDRHKGYRRDVFVDVAGE
VDRRIAFGQLKRFAWRELQLAT

DNFSEKNVLGQGGFGKVYKGVLPD
TPKVAVKRLTDFESPGGDAAFQ
REVEMISVAVHRNLLRLIGFCT
TQTERLLVYPFMQNLSLAHRLR
EIKAGDPVLDWETRKRIALGAA
RGFEYLHEHCNPKIIHRDVKAA
NVLLDEDFEAVVGDFGLAKLVD
VRRTNVTTQVRGTMGHIAPEYL
STGKSSERTDVFGYGIMLLELV
TGQRAIDFSRLEEEDDVLLLDH
VKKLEREKRLGAIVDKNLDGEY
IKEEVEMMIQVALLCTQGSPED
RPVMSEVVRMLE

GEGLAERWEEWQNVEVTRRHEFE

RLQRRFDWGEDSMHNQDAIELSGGR

Arabidopsis thaliana RKS7 cDNA

The start codon encoding the first predicted methionine residue of the gene product has been indicated by bold capitals.

The first stopcodon has been underlined.

Nucleotides predicted to encode protein sequences are in capitals. Leader and tailer sequences are in lowercase letters.

acatcttgttttctgctcattcctctgtttcaacaATGGAGAGTACTATTGTTATGATGA
TGATGATAACAAGATCTTTCTTTTGCTTCTTGGGATTTTTATGCCTTCTCTGCTCTTCTG
TTCACGGATTGCTTTCTCCTAAAGGTGTTAACTTTGAAGTGCAAGCTTTGATGGACATAA
AAGCTTCATTACATGATCCTCATGGTGTTCTTGATAACTGGGATAGAGATGCTGTTGATC
CTTGTAGTTGGACAATGGTCACTTGTTCTTCTGAAAACTTTGTCATTGGCTTAGGCACAC
CAAGTCAGAATTTATCTGGTACACTATCTCCAAGCATTACCAACTTAACAAATCTTCGGA
TTGTGCTGTTGCAGAACAACAACATAAAAGGAAAAATTCCTGCTGAGATTGGTCGGCTTA
CGAGGCTTGAGACTCTTGATCTTTCTGATAATTTCTTCCACGGTGAAATTCCTTTTTCAG
TAGGCTATCTACAAAGCCTGCAATATCTGAGGCTTAACAACAATTCTCTCTCTGGAGTGT
TTCCTCTGTCACTATCTAATATGACTCAACTTGCCTTTCTTGATTTATCATACAACAATC
TTAGTGGTCCTGTTCCAAGATTTGCTGCAAAGACGTTTAGCATCGTTGGGAACCCGCTGA
TATGTCCAACGGGTACCGAACCAGACTGCAATGGAACAACATTGATACCTATGTCTATGA
ACTTGAATCAAACTGGAGTTCCTTTATACGCCGGTGGATCGAGGAATCACAAAATGGCAA
TCGCTGTTGGATCCAGCGTTGGGACTGTATCATTAATCTTCATTGCTGTTGGTTTGTTTC
TCTGGTGGAGACAAAGACATAACCAAAACACATTCTTTGATGTTAAAGATGGGAATCATC
ATGAGGAAGTTTCACTTGGAAACCTGAGGAGATTTGGTTTCAGGGAGCTTCAGATTGCGA
CCAATAACTTCAGCAGTAAGAACTTATTGGGGAAAGGTGGCTATGGAAATGTATACAAAG
GAATACTTGGAGATAGTACAGTGGTTGCAGTGAAAAGGCTTAAAGATGGAGGAGCATTGG
GAGGAGAGATTCAGTTTCAGACAGAAGTTGAAATGATCAGTTTAGCTGTTCATCGAAATC
TCTTAAGACTCTACGGTTTCTGCATCACACAAACTGAGAAGCTTCTAGTTTATCCTTATA
TGTCTAATGGAAGCGTTGCATCTCGAATGAAAGCAAAACCTGTTCTTGACTGGAGCATAA
GGAAGAGGATAGCCATAGGAGCTGCAAGAGGGCTTGTGTATCTCCATGAGCAATGTGATC
CGAAGATTATCCACCGCGATGTCAAAGCAGCGAATATACTTCTTGATGACTACTGTGAAG
CTGTGGTTGGCGATTTTGGTTTAGCTAAACTCTTGGATCATCAAGATTCTCATGTGACAA
CCGCGGTTAGAGGCACGGTGGGTCACATTGCTCCAGAGTATCTCTCAACTGGTCAATCCT
CTGAGAAAACAGATGTTTTTGGCTTCGGGATTCTTCTTCTTGAGCTTGTAACCGGACAAA
GAGCTTTTGAGTTTGGTAAAGCGGCTAACCAGAAAGGTGTGATGCTTGATTGGGTTAAAA
AGATTCATCAAGAGAAGAAACTTGAGCTACTTGTGGATAAAGAGTTGTTGAAGAAGAAGA
GCTACGATGAGATTGAGTTAGACGAAATGGTAAGAGTAGCTTTGTTGTGCACACAGTACC
TGCCAGGACATAGACCAAAAATGTCTGAAGTTGTTCGAATGCTGGAAGGAGATGGACTTG
CAGAGAAATGGGAAGCTTCTCAAAGATCAGACAGTGTTTCAAAATGTAGCAACAGGATAA

ATGAATTGATGTCATCTTCAGACAGATACTCTGATCTTACCGATGACTCTAGTTTACTTG
TGCAAGCAATGGAGCTCTCTGGTCCTAGATGAaatctatacatgaatctgaagaagaaga
agaacatgcatctgtttcttgaatcaagagggattcttgtttttttgtataatagagagg
ttttttggagggaaatgttgtgtctctgtaactgtataggcttgttgtgtaagaagttat
tactgcacttagggttaattcaaagttctttacataaaaaatgattagttgcgttgaata
gagggaacactttgggagatttcatgtatgaaatttggaaaaaaaaaaaaaaaaaaa

Predicted amino acid sequence of the *Arabidopsis thaliana* RKS7 protein.

Different domains are spaced and shown from the N-terminus towards the C-terminus. Overall domain structure is similar as described in Schmidt et al (1997).

At the predicted extracellular domain the first domain represents a signal sequence. The second domain contains a leucine zipper motif, containing 3 leucine residues, each separated by seven other amino acids. The third domain contains conserved cysteine residues, involved in disulphate bridge formation. The fourth domain contains a leucine rich repeat domain, consisting of 5 complete repeats of each approximately 24 amino acid residues. The fifth domain contains many serine and proline residues, and is likely to contain hydroxy-proline residues, and to be a site for O-glycosylation. The sixth domain contains a single transmembrane domain after which the prodicted intracellular domains are positioned. The seventh domain has an unknown function. The eight domain represents a serine / threonine protein kinase domain (Schmidt et al. 1997) and is probably also containing sequences for protein / protein interactions. The ninth domain has an unknown function. The last and tenth domain at the C-terminal end represents part of a single leucine rich repeat, probably involved in protein / protein interactions.

MESTIVMMMMITRSFF
CFLGFLCLLCSSVHGLLSPKGVNFEV

QALMDIKASLHDP

HGVLDNWDRDAVD

PCSWTMVTCSSENFVIG

LGTPSQNLSGTL

SPSITNLTNLRIVLLQNNNIKGKI

PAEIGRLTRLETLDLSDNFFHGEI

PFSVGYLQSLQYLRLNNNSLSGVF

PLSLSNMTQLAFLDLSYNNLSGPV

PRFAA KTFSIVGNPLICPT

GTEPDCNGTTLIPMSMNL

NQTGVPLYAGGSRNHKMA

IAVGSSVGTVSLIFIAVGLFLWW

RQRHNQNTFFDVKDGNHHE

EVSLGNLRRFGFRELQIAT

NNFSSKNLLGKGGYGNVYKGILGD

STVVAVKRLKDGGALGGEIQFQ

TEVEMISLAVHRNLLRLYGFCI

TQTEKLLVYPYMSNGSVA

SRMKAKPVLDWSIRKRIAIGAA

RGLVYLHEQCDPKIIHRDVKAA

NILLDDYCEAVVGDFGLAKLLD

HQDSHVTTAVRGTVGHIAPEYL

STGQSSEKTDVFGFGILLLELV

TGQRAFEFGKAANQKGVMLDW

VKKIHQEKKLELLVDKELLKKKSY

DEIELDEMVRVALLCTQYLPGH

RPKMSEVVRMLE

GDGLAEKWEASQRSDS

VSKCSNRINELMSSS

DRYSDLTDDSSLLVQAMELSGPR

Arabidopsis thaliana RKS8 cDNA

The start codon encoding the first predicted methionine residue

of the gene product has been indicated by bold capitals.

The first stopcodon has been underlined.

Nucleotides predicted to encode protein sequences are in capitals. Leader and tailer sequences are in lowercase letters.

gttttttttttttaccctcttggaggatctgggaggagaaatttgctttttttggtaa
ATGGGGAGAAAAAAGTTTGAAGCTTTTGGTTTTGTCTGCTTAATCTCACTGCTTCTTCTG
TTTAATTCGTTATGGCTTGCCTCTTCTAACATGGAAGGTGATGCACTGCACAGTTTGAGA
GCTAATCTAGTTGATCCAAATAATGTCTTGCAAAGCTGGGATCCTACGCTTGTTAATCCG
TGTACTTGGTTTCACGTAACGTGTAACAACGAGAACAGTGTTATAAGAGTCGATCTTGGG
AATGCAGACTTGTCTGGTCAGTTGGTTCCTCAGCTAGGTCAGCTCAAGAACTTGCAGTAC
TTGGAGCTTTATAGTAATAACATAACCGGGCCGGTTCCAAGCGATCTTGGGAATCTGACA
AACTTAGTGAGCTTGGATCTTTACTTGAACAGCTTCACTGGTCCAATTCCAGATTCTCTA
GGAAAGCTATTCAAGCTTCGCTTTCTTCGGCTCAACAATAACAGTCTCACCGGACCAATT
CCCATGTCATTGACTAATATCATGACCCTTCAAGTTTTGGATCTGTCGAACAACCGATTA
TCCGGATCTGTTCCTGATAATGGTTCCTTCTCGCTCTTCACTCCCATCAGTTTTGCTAAC
AACTTGGATCTATGCGGCCCAGTTACTAGCCGTCCTTGTCCTGGATCTCCCCCGTTTTCT
CCTCCACCACCTTTTATACCACCTCCCATAGTTCCTACACCAGGTGGGTATAGTGCTACT
GGAGCCATTGCGGGAGGAGTTGCTGCTGGTGCTGCTTTACTATTTGCTGCCCCTGCTTTA
GCTTTTGCTTGGTGGCGTAGAAGAAAACCTCAAGAATTCTTCTTTGATGTTCCTGCCGAA
GAGGACCCTGAGGTTCACTTGGGGCAGCTTAAGCGGTTCTCTCTACGGGAACTTCAAGTA
GCAACTGATAGCTTCAGCAACAAGAACATTTTGGGCCGAGGTGGGTTCGGAAAAGTCTAC
AAAGGCCGTCTTGCTGATGGAACACTTGTTGCAGTCAAACGGCTTAAAGAAGAGCGAACC
CCAGGTGGCGAGCTCCAGTTTCAGACAGAAGTGGAGATGATAAGCATGGCCGTTCACAGA
AATCTCCTCAGGCTACGCGGTTTCTGTATGACCCCTACCGAGAGATTGCTTGTTTATCCT
TACATGGCTAATGGAAGTGTCGCTTCCTGTTTGAGAGAACGTCCACCATCACAGTTGCCT
CTAGCCTGGTCAATAAGACAGCAAATCGCGCTAGGATCAGCGAGGGGTTTGTCTTATCTT
CATGATCATTGCGACCCCAAAATTATTCACCGTGATGTGAAAGCTGCTAATATTCTGTTG
GACGAGGAATTTGAGGCGGTGGTAGGTGATTTCGGGTTAGCTAGACTTATGGACTATAAA
GATACTCATGTCACAACGGCTGTGCGTGGGACTATTGGACACATTGCTCCTGAGTATCTC
TCAACTGGAAAATCTTCAGAGAAAACTGATGTTTTTGGCTACGGGATCATGCTTTTGGAA
CTGATTACAGGTCAGAGAGCTTTTGATCTTGCAAGACTGGCGAATGACGATGACGTTATG
CTCCTAGATTGGGTGAAAGGGCTTTTGAAGGAGAAGAAGCTGGAGATGCTTGTGGATCCT
GACCTGCAAAGCAATTACACAGAAGCAGAAGTAGAACAGCTCATACAAGTGGCTCTTCTC
TGCACACAGAGCTCACCTATGGAACGACCTAAGATGTCTGAGGTTGTTCGAATGCTTGAA
GGTGACGGTTTAGCGGAGAAATGGGACGAGTGGCAGAAAGTGGAAGTTCTCAGGCAAGAA

GTGGAGCTCTCTTCTCACCCCACCTCTGACTGGATCCTTGATTCGACTGATAATCTTCAT
GCTATGGAGTTGTCTGGTCCAAGATAAacgacattgtaatttgcctaacagaaaagagaa
agaacagagaaatattaagagaatcacttctctgtattctt

Predicted amino acid sequence of the *Arabidopsis thaliana* RKS8 protein.

Different domains are spaced and shown from the N-terminus towards the C-terminus. Overall domain structure is similar as described in Schmidt et al (1997).

At the predicted extracellular domain the first domain represents a signal sequence. The second domain contains a leucine zipper motif, containing 4 leucine residues, each separated by seven other amino acids. The third domain contains conserved cysteine residues, involved in disulphate bridge formation. The fourth domain contains a leucine rich repeat domain, consisting of 5 complete repeats of each approximately 24 amino acid residues. The fifth domain contains many serine and proline residues, and is likely to contain hydroxy-proline residues, and to be a site for O-glycosylation. The sixth domain contains a single transmembrane domain after which the prodicted intracellular domains are positioned. The seventh domain has an unknown function. The eight domain represents a serine / threonine protein kinase domain (Schmidt et al. 1997) and is probably also containing sequences for protein / protein interactions. The ninth domain has an unknown function. The last and tenth domain at the C-terminal end represents part of a single leucine rich repeat, probably
involved in protein / protein interactions.

MGRKKFEAFGFVCLISLLLLFNSL
WLASSNMEG

DALHSLRANLVDP
NNVLQSWDPTLVN

PCTWFHVTCNNENSVIRV

DLGNADLSGQLV
P QLGQLKNLQYLELYSNNITGPV
PSDLGNLTNLVSLDLYLNSFTGPI
PDSLGKLFKLRFLRLNNNSLTGPI
PMSLTNIMTLQVLDLSNNRLSGSV
PDNGSFSLFTPISFANNLDLCGPV

TSRPCPGSPPFSPPPP
FIPPPIVPTPGGYSATG

AIAGGVAAGAAL
LFAAPALAFAWW

RRRKPQEFFFDVPAEEDPE
VHLGQLKRFSLRELQVAT

DSFSNKNILGRGGFGKVYKGRLAD
GTLVAVKRLKEERTPGGELQFQ
TEVEMISMAVHRNLLRLRGFCM
TPTERLLVYPYMANGSVASCLR
ERPPSQLPLAWSIRQQIALGSA
RGLSYLHDHCDPKIIHRDVKAA
NILLDEEFEAVVGDFGLARLMD
YKDTHVTTAVRGTIGHIAPEYL
STGKSSEKTDVFGYGIMLLELI
TGQRAFDLARLANDDDVMLLDW
VKGLLKEKKLEMLVDPDLQSNY
TEAEVEQLIQVALLCTQSSPME
RPKMSEVVRMLE

GDGLAEKWDEWQKVEVLRQEVELS

SHPTSDWILDSTDNLHAMELSGPR

Arabidopsis thaliana rks10 cDNA

The start codon encoding the first predicted methionine residue

of the gene product has been indicated by bold capitals.

The first stopcodon has been underlined.

Nucleotides predicted to encode protein sequences are in capitals. Leader and tailer sequences are in lowercase letters.

atcaggggttttaacaatgatggattttctctgatgagggatagttctagggtttgtttt
taatctcttgaggataaa**ATG**GAACGAAGATTAATGATCCCTTGCTTCTTTTGGTTGATT
CTCGTTTTGGATTTGGTTCTCAGAGTCTCGGGCAACGCCGAAGGTGATGCTCTAAGTGCA
CTGAAAAACAGTTTAGCCGACCCTAATAAGGTGCTTCAAAGTTGGGATGCTACTCTTGTT
ACTCCATGTACATGGTTTCATGTTACTTGCAATAGCGACAATAGTGTTACACGTGTTGAC
CTTGGGAATGCAAATCTATCTGGACAGCTCGTAATGCAACTTGGTCAGCTTCCAAACTTG
CAGTACTTGGAGCTTTATAGCAATAACATTACTGGGACAATCCCAGAACAGCTTGGAAAT
CTGACGGAATTGGTGAGCTTGGATCTTTACTTGAACAATTTAAGCGGGCCTATTCCATCA
ACTCTCGGCCGACTTAAGAAACTCCGTTTCTTGCGTCTTAATAACAATAGCTTATCTGGA
GAAATTCCAAGGTCTTTGACTGCTGTCCTGACGCTACAAGTTCTGGATCTCTCAAACAAT
CCTCTCACCGGAGATATTCCTGTTAATGGTTCCTTTTCACTTTTCACTCCAATCAGTTTT
GCCAACACCAAGTTGACTCCCCTTCCTGCATCTCCACCGCCTCCTATCTCTCCTACACCG
CCATCACCTGCAGGGAGTAATAGAATTACTGGAGCGATTGCGGGAGGAGTTGCTGCAGGT
GCTGCACTTCTATTTGCTGTTCCGGCCATTGCACTAGCTTGGTGGCGAAGGAAAAAGCCG
CAGGACCACTTCTTTGATGTACCAGCTGAAGAGGACCCAGAAGTTCATTTAGGACAACTG
AAGAGGTTTTCATTGCGTGAACTACAAGTTGCTTCGGATAATTTTAGCAACAAGAACATA
TTGGGTAGAGGTGGTTTTGGTAAAGTTTATAAAGGACGGTTAGCTGATGGTACTTTAGTG
GCCGTTAAAAGGCTAAAAGAGGAGCGCACCCAAGGTGGCGAACTGCAGTTCCAGACAGAG
GTTGAGATGATTAGTATGGCGGTTCACAGAAACTTGCTTCGGCTTCGTGGATTTTGCATG
ACTCCAACCGAAAGATTGCTTGTTTATCCCTACATGGCTAATGGAAGTGTTGCCTCCTGT
TTAAGAGAACGTCCCGAGTCCCAGCCACCACTTGATTGGCCAAAGAGACAGCGTATTGCG
TTGGGATCTGCAAGAGGGCTTGCGTATTTACATGATCATTGCGACCCAAAGATTATTCAT
CGAGATGTGAAAGCTGCAAATATTTTGTTGGATGAAGAGTTTGAAGCCGTGGTTGGGGAT
TTTGGACTTGCAAAACTCATGGACTACAAAGACACACATGTGACAACCGCAGTGCGTGGG
ACAATTGGTCATATAGCCCCTGAGTACCTTTCCACTGGAAAATCATCAGAGAAAACCGAT
GTCTTTGGGTATGGAGTCATGCTTCTTGAGCTTATCACTGGACAAAGGGCTTTTGATCTT
GCTCGCCTCGCGAATGATGATGATGTCATGTTACTAGACTGGGTGAAAGGGTTGTTAAAA
GAGAAGAAATTGGAAGCACTAGTAGATGTTGATCTTCAGGGTAATTACAAAGACGAAGAA
GTGGAGCAGCTAATCCAAGTGGCTTTACTCTGCACTCAGAGTTCACCAATGGAAAGACCC
AAAATGTCTGAAGTTGTAAGAATGCTTGAAGGAGATGGTTTAGCTGAGAGATGGGAAGAG
TGGCAAAAGGAGGAAATGTTCAGACAAGATTTCAACTACCCAACCCACCATCCAGCCGTG

TCTGGCTGGATCATTGGCGATTCCACTTCCCAGATCGAAAACGAATACCCCTCGGGTCCA
AGATAAgattcgaaacacgaatgttttttctgtattttgtttttctctgtatttattgag
ggttttagcttc

Predicted amino acid sequence of the *Arabidopsis thaliana* RKS10 protein.

Different domains are spaced and shown from the N-terminus towards the C-terminus. Overall domain structure is similar as described in Schmidt et al (1997).

At the predicted extracellular domain the first domain represents a signal sequence. The second domain contains a leucine zipper motif, containing 4 leucine residues, each separated by seven other amino acids. The third domain contains conserved cysteine residues, involved in disulphate bridge formation. The fourth domain contains a leucine rich repeat domain, consisting of 5 complete repeats of each approximately 24 amino acid residues. The fifth domain contains many serine and proline residues, and is likely to contain hydroxy-proline residues, and to be a site for O-glycosylation. The sixth domain contains a single transmembrane domain after which the prodicted intracellular domains are positioned. The seventh domain has an unknown function. The eight domain represents a serine / threonine protein kinase domain (Schmidt et al. 1997) and is probably also containing sequences for protein / protein interactions. The ninth domain has an unknown function. The last and tenth domain at the C-terminal end represents part of a single leucine rich repeat, probably
involved in protein / protein interactions.

MERRLMIPCFFWLILVL
DLVLRVSGNAEG

DALSALKNSLADP
NKVLQSWDATLVT

PCTWFHVTCNSDNSVTRV

DLGNANLSGQLV
M QLGQLPNLQYLELYSNNITGTI
PEQLGNLTELVSLDLYLNNLSGPI
PSTLGRLKKLRFLRLNNNSLSGEI
PRSLTAVLTLQVLDLSNNPLTGDI
PVNGSFSLTPISFANTK LT PL

PASPPPPISPTPPSPAGSNRITG

AIAGGVAAGAAL
LFAVPAIALAWW

RRKKPQDHFFDVPAEEDPE
VHLGQLKRFSLRELQVAS

DNFSNKNILGRGGFGKVYKGRLAD
GTLVAVKRLKEERTQGGELQFQ
TEVEMISMAVHRNLLRLRGFCM
TPTERLLVYPYMANGSVASCLR
ERPESQPPLDWPKRQRIALGSA
RGLAYLHDHCDPKIIHRDVKAA
NILLDEEFEAVVGDFGLAKLMD
YKDTHVTTAVRGTIGHIAPEYL
STGKSSEKTDVFGYGVMLLELI
TGQRAFDLARLANDDDVMLLDW
VKGLLKEKKLEALVDVDLQGNY
KDEEVEQLIQVALLCTQSSPME
RPKMSEVVRMLE

GDGLAERWEEWQKEEMFRQDFNYPTHH

PAVSGWIIGDSTSQIENEYPSGPR

Arabidopsis thaliana RKS 11 cDNA

The start codon encoding the first predicted methionine residue

of the gene product has been indicated by bold capitals.

The first stopcodon has been underlined.

Nucleotides predicted to encode protein sequences are in capitals. Leader and tailer sequences are in lowercase letters.

ttgttaacctctcgtaactaaaatcttcc**ATG**GTAGTAGTAACAAAGAAGACCATGAAGA
TTCAAATTCATCTCCTTTACTCGTTCTTGTTCCTCTGTTTCTCTACTCTCACTCTATCTT
CTGAGCCCAGAAACCCTGAAGTTGAGGCGTTGATAAGTATAAGGAACAATTTGCATGATC
CTCATGGAGCTTTGAACAATTGGGACGAGTTTTCAGTTGATCCTTGTAGCTGGGCTATGA
TCACTTGCTCTCCCGACAACCTCGTCATTGGACTAGGAGCGCCGAGCCAGTCTCTCTCGG
GAGGTTTATCTGAGTCTATCGGAAATCTCACAAATCTCCGACAAGTGTCATTGCAAAATA
ACAACATCTCCGGCAAAATTCCACCGGAGCTCGGTTTTCTACCCAAATTACAAACCTTGG
ATCTTTCCAACAACCGATTCTCCGGTGACATCCCTGTTTCCATCGACCAGCTAAGCAGCC
TTCAATATCTGAGACTCAACAACAACTCTTTGTCTGGGCCCTTCCCTGCTTCTTTGTCCC
AAATTCCTCACCTCTCCTTCTTGGACTTGTCTTACAACAATCTCAGTGGCCCTGTTCCTA
AATTCCCAGCAAGGACTTTAAACGTTGCTGGTAATCCTTTGATTTGTAGAAGCAACCCAC
CTGAGATTTGTTCTGGATCAATCAATGCAAGTCCACTTTCTGTTTCTTTGAGCTCTTCAT
CAGGACGCAGGTCTAATAGATTGGCAATAGCTCTTAGTGTAAGCCTTGGCTCTGTTGTTA
TACTAGTCCTTGCTCTCGGGTCCTTTTGTTGGTACCGAAAGAAACAAAGAAGGCTACTGA
TCCTTAACTTAAACGCAGATAAACAAGAGGAAGGGCTTCAAGGACTTGGGAATCTAAGAA
GCTTCACATTCAGAGAACTCCATGTTTATACAGATGGTTTCAGTTCCAAGAACATTCTCG
GCGCTGGTGGATTCGGTAATGTGTACAGAGGCAAGCTTGGAGATGGGACAATGGTGGCAG
TGAAACGGTTGAAGGATATTAATGGAACCTCAGGGGATTCACAGTTTCGTATGGAGCTAG
AGATGATTAGCTTAGCTGTTCATAAGAATCTGCTTCGGTTAATTGGTTATTGCGCAACTT
CTGGTGAAAGGCTTCTTGTTTACCCTTACATGCCTAATGGAAGCGTCGCCTCTAAGCTTA
AATCTAAACCGGCATTGGACTGGAACATGAGGAAGAGGATAGCAATTGGTGCAGCGAGAG
GTTTGTTGTATCTACATGAGCAATGTGATCCCAAGATCATTCATAGAGATGTAAAGGCAG
CTAATATTCTCTTAGACGAGTGCTTTGAAGCTGTTGTTGGTGACTTTGGACTCGCAAAGC
TCCTTAACCATGCGGATTCTCATGTCACAACTGCGGTCCGTGGTACGGTTGGCCACATTG
CACCTGAATATCTCTCCACTGGTCAGTCTTCTGAGAAAACCGATGTGTTTGGGTTCGGTA
TACTATTGCTCGAGCTCATAACCGGACTGAGAGCTCTTGAGTTTGGTAAAACCGTTAGCC
AGAAAGGAGCTATGCTTGAATGGGTGAGGAAATTACATGAAGAGATGAAAGTAGAGGAAC
TATTGGATCGAGAACTCGGAACTAACTACGATAAGATTGAAGTTGGAGAGATGTTGCAAG
TGGCTTTGCTATGCACACAATATCTGCCAGCTCATCGTCCTAAAATGTCTGAAGTTGTTT
TGATGCTTGAAGGCGATGGATTAGCCGAGAGATGGGCTGCTTCGCATAACCATTCACATT
TCTACCATGCCAATATCTCTTTCAAGACAATCTCTTCTCTGTCTACTACTTCTGTCTCAA

GGCTTGACGCACATTGCAATGATCCAACTTATCAAATGTTTGGATCTTCGGCTTTCGATG
ATGACGATGATCATCAGCCTTTAGATTCCTTTGCCATGGAACTATCCGGTCCAAGATAAc
acaatgaaagaaagatatcatttttacgatggatcaaacaatccaatgaaaaaa

Predicted amino acid sequence of the *Arabidopsis thaliana* RKS11 protein.

Different domains are spaced and shown from the N-terminus towards the C-terminus. Overall domain structure is similar as described in Schmidt et al (1997).

At the predicted extracellular domain the first domain represents a signal sequence. The second domain contains a leucine zipper motif, containing 3 leucine residues, each separated by seven other amino acids. The third domain contains conserved cysteine residues, involved in disulphate bridge formation. The fourth domain contains a leucine rich repeat domain, consisting of 5 complete repeats of each approximately 24 amino acid residues. The fifth domain contains many serine and proline residues, and is likely to contain hydroxy-proline residues, and to be a site for O-glycosylation. The sixth domain contains a single transmembrane domain after which the prodicted intracellular domains are positioned. The seventh domain has an unknown function. The eight domain represents a serine / threonine protein kinase domain (Schmidt et al. 1997) and is probably also containing sequences for protein / protein interactions. The ninth domain has an unknown function. The last and tenth domain at the C-terminal end represents part of a single leucine rich repeat, probably
involved in protein / protein interactions.

MVVVTKKTMKIQIHLLYSFLFL
CFSTLTLSSEPRNPEV

EALISIRNNLHDP
HGALNNWDEFSVD

PCSWAMITCSPDNLVIGL

GAPSQSLSGGLS
ESIGNLTNLRQVSLQNNNISGKI
PPELGFLPKLQTLDLSNNRFSGDI
PVSIDQLSSLQYLRLNNNSLSGPF
PASLSQIPHLSFLDLSYNNLSGPV
PKFPARTFNVAGNPLICRSN

PPEICSGSINASPL
SVSLSSSSGRRSNR

LAIALSVSLGSVVIL
VLALGSFCWY

RKKQRRLLILNLNGADKQEE
GLQGLGNLRSFTFRELHVYT

DGFSSKNILGAGGFGNVYRGKLGD
GTMVAVKRLKDINGTSGDSQFR
MELEMISLAVHKNLLRLIGYCA
TSGERLLVYPYMPNGSVASKLK
SKPALDWNMRKRIAIGAA
RGLLYLHEQCDPKIIHRDVKAA
NILLDECFEAVVGDFGLAKLLN
HADSHVTTAVRGTVGHIAPEYL
STGQSSEKTDVFGFGILLLELI
TGLRALEFGKTVSQKGAMLEW
VRKLHEEMKVEELLDRELGTNY
DKIEVGEMLQVALLCTQYLPAH
RPKMSEVVLMLE

GDGLAERWAASHNHSHFYHANI
SFKTISSLSTTSVSRLDAHCNDPTYQMFG

SSAFDDDDDHQPLDSFAMELSGPR

Arabidopsis thaliana RKS12 cDNA

The start codon encoding the first predicted methionine residue
of the gene product has been indicated by bold capitals.
The first stopcodon has been underlined.
Nucleotides predicted to encode protein sequences are in capitals. Leader and tailer sequences are in lowercase letters.

tttaaaaaccttgctagttctcaattctcatgactttgcttttagtcttagaagtggaaa
ATGGAACATGGATCATCCCGTGGCTTTATTTGGCTGATTCTATTTCTCGATTTTGTTTCC
AGAGTCACCGGAAAAACACAAGTTGATGCTCTCATTGCTCTAAGAAGCAGTTTATCATCA
GGTGACCATACAAACAATATACTCCAAAGCTGGAATGCCACTCACGTTACTCCATGTTCA
TGGTTTCATGTTACTTGCAATACTGAAAACAGTGTTACTCGTCTTGACCTGGGGAGTGCT
AATCTATCTGGAGAACTGGTGCCACAGCTTGCTCAGCTTCCAAATTTGCAGTACTTGGAA
CTTTTTAACAATAATATTACTGGGGAGATACCTGAGGAGCTTGGCGACTTGATGGAACTA
GTAAGCTTGGACCTTTTTGCAAACAACATAAGCGGTCCCATCCCTTCCTCTCTTGGCAAA
CTAGGAAAACTCCGCTTCTTGCGTCTTTATAACAACAGCTTATCTGGAGAAATTCCAAGG
TCTTTGACTGCTCTGCCGCTGGATGTTCTTGATATCTCAAACAATCGGCTCAGTGGAGAT
ATTCCTGTTAATGGTTCCTTTTCGCAGTTCACTTCTATGAGTTTTGCCAATAATAAATTA
AGGCCGCGACCTGCATCTCCTTCACCATCACCTTCAGGAACGTCTGCAGCAATAGTAGTG
GGAGTTGCTGCGGGTGCAGCACTTCTATTTGCGCTTGCTTGGTGGCTGAGAAGAAAACTG
CAGGGTCACTTTCTTGATGTACCTGCTGAAGAAGACCCAGAGGTTTATTTAGGACAATTT
AAAAGGTTCTCCTTGCGTGAACTGCTAGTTGCTACAGAGAAATTTAGCAAAAGAAATGTA
TTGGGCAAAGGACGTTTTGGTATATTGTATAAAGGACGTTTAGCTGATGACACTCTAGTG
GCTGTGAAACGGCTAAATGAAGAACGTACCAAGGGTGGGGAACTGCAGTTTCAAACCGAA
GTTGAGATGATCAGTATGGCCGTTCATAGGAACTTGCTTCGGCTTCGTGGCTTTTGCATG
ACTCCAACTGAAAGATTACTTGTTTATCCCTACATGGCTAATGGAAGTGTTGCTTCTTGT
TTAAGAGAGCGTCCTGAAGGCAATCCAGCCCTTGACTGGCCAAAAAGAAAGCATATTGCT
CTGGGATCAGCAAGGGGGCTCGCATATTTACACGATCATTGCGACCAAAAGATCATTCAC
CTGGATGTGAAAGCTGCAAATATACTGTTAGATGAAGAGTTTGAAGCTGTTGTTGGAGAT
TTTGGGCTAGCAAAATTAATGAATTATAACGACTCCCATGTGACAACTGCTGTACGGGGT
ACGATTGGCCATATAGCGCCCGAGTACCTCTCGACAGGAAAATCTTCTGAGAAGACTGAT
GTTTTTGGGTACGGGGTCATGCTTCTCGAGCTCATCACTGGACAAAAGGCTTTCGATCTT
GCTCGGCTTGCAAATGATGATGATATCATGTTACTCGACTGGGTGAAAGAGGTTTTGAAA
GAGAAGAAGTTGGAAAGCCTTGTGGATGCAGAACTCGAAGGAAAGTACGTGGAAACAGAA
GTGGAGCAGCTGATACAAATGGCTCTGCTCTGCACTCAAAGTTCTGCAATGGAACGTCCA
AAGATGTCAGAAGTAGTGAGAATGCTGGAAGGAGATGGTTTAGCTGAGAGATGGGAAGAA
TGGCAAAAGGAGGAGATGCCAATACATGATTTTAACTATCAAGCCTATCCTCATGCTGGC
ACTGACTGGCTCATCCCCTATTCCAATTCCCTTATCGAAAACGATTACCCCTCGGGGCCA

AGATAAccttttagaaagggtcatttcttgtgggttcttcaacaagtatatatataggta
gtgaagttgtaagaagcaaaaccccacattcacctttgaatatcactactctataa

Predicted amino acid sequence of the *Arabidopsis thaliana* RKS12 protein.

Different domains are spaced and shown from the N-terminus towards the C-terminus. Overall domain structure is similar as described in Schmidt et al (1997).

At the predicted extracellular domain the first domain represents a signal sequence. The second domain contains a leucine zipper motif, containing 2 leucine residues, each separated by seven other amino acids. The third domain contains conserved cysteine residues, involved in disulphate bridge formation. The fourth domain contains a leucine rich repeat domain, consisting of 5 complete repeats of each approximately 24 amino acid residues. The fifth domain contains many serine and proline residues, and is likely to contain hydroxy-proline residues, and to be a site for O-glycosylation. The sixth domain contains a single transmembrane domain after which the prodicted intracellular domains are positioned. The seventh domain has an unknown function. The eight domain represents a serine / threonine protein kinase domain (Schmidt et al. 1997) and is probably also containing sequences for protein / protein interactions. The ninth domain has an unknown function. The last and tenth domain at the C-terminal end represents part of a single leucine rich repeat, probably
involved in protein / protein interactions.

MEHGSSRGFI
WLILFLDFVSRVTGKTQV

DALIALRSSLSSGDHTNNILQ
SWNATHVT

PCSWFHVTCNTENSVTRL

DLGSANLSGELV

P QLAQLPNLQYLELFNNNITGEI

PEELGDLMELVSLDLFANNISGPI

PSSLGKLGKLRFLRLYNNSLSGEI

PRSLTALP LDVLDISNNRLSGDI

PVNGSFSQFTSMRFA NNKLRPR

PASPSPSPSGGTS

AAIVVGVAAGAALLFALAWWL

RRKLQGHFLDVPAAEEDPE

VYLGQFKRFSLRELLVAT

EKFSKRNVLGKGRFGILYKGRLAD

DTLVAVKRLNEERTKGGELQFQ

TEVEMISMAVHRNLLRLRGFCM

TPTERLLVYPYMANGSVASCLR

ERPEGNPALDWPKRKHIALGSA

RGLAYLHDHCDQKIIHLDVKAA

NILLDEEFEAVVGDFGLAKLMN

YNDSHVTTAVRGTIGHIAPEYL

STGKSSEKTDVFGYGVMLLELI

TGQKAFDLARLANDDDIMLLDW

VKEVLKEKKLESLVDAELEGKY

VETEVEQLIQMALLCTQSSAME

RPKMSEVVRMLE

GDGLAERWEEWQKEEMPIHDFNYQAY

PHAGTDWLIPYSNSLIENDYPSGPR

Arabidopsis thaliana RKS13 cDNA

The start codons encoding predicted the methionine residue of the gene product has been indicated by bold capitals. The first stopcodon has been underlined.

Nucleotides predicted to encode protein sequences are in capitals. Leader and tailer sequences are in lowercase letters.

taataaacctctaataataatggctttgcttttactctgatgacaagttcaaaaATGGAA
CAAAGATCACTCCTTTGCTTCCTTTATCTGCTCCTACTATTCAATTTCACTCTCAGAGTC
GCTGGAAACGCTGAAGGTGATGCTTTGACTCAGCTGAAAAACAGTTTGTCATCAGGTGAC
CCTGCAAACAATGTACTCCAAAGCTGGGATGCTACTCTTGTTACTCCATGTACTTGGTTT
CATGTTACTTGCAATCCTGAGAATAAAGTTACTCGTGTTGACCTTGGGAATGCAAAACTA
TCTGGAAAGTTGGTTCCAGAACTTGGTCAGCTTTTAAACTTGCAGTACTTGGAGCTTTAT
AGCAATAACATTACAGGGGAGATACCTGAGGAGCTTGGCGACTTGGTGGAACTAGTAAGC
TTGGATCTTTACGCAAACAGCATAAGCGGTCCCATCCCTTCGTCTCTTGGCAAACTAGGA
AAACTCCGGTTCTTGCGTCTTAACAACAATAGCTTATCAGGGGAAATTCCAATGACTTTG
ACTTCTGTGCAGCTGCAAGTTCTGGATATCTCAAACAATCGGCTCAGTGGAGATATTCCT
GTTAATGGTTCTTTTTCGCTCTTCACTCCTATCAGTTTTGCGAATAATAGCTTAACGGAT
CTTCCCGAACCTCCGCCTACTTCTACCTCTCCTACGCCACCACCACCTTCAGGGGGGCAA
ATGACTGCAGCAATAGCAGGGGGAGTTGCTGCAGGTGCAGCACTTCTATTTGCTGTTCCA
GCCATTGCGTTTGCTTGGTGGCTCAGAAGAAAACCACAGGACCACTTTTTTGATGTACCT
GCTGAAGAAGACCCAGAGGTTCATTTAGGACAACTCAAAAGGTTTACCTTGCGTGAACTG
TTAGTTGCTACTGATAACTTTAGCAATAAAAATGTATTGGGTAGAGGTGGTTTTGGTAAA
GTGTATAAAGGACGTTTAGCCGATGGCAATCTAGTGGCTGTCAAAAGGCTAAAAGAAGAA
CGTACCAAGGGTGGGGAACTGCAGTTTCAAACCGAAGTTGAGATGATCAGTATGGCCGTT
CATAGGAACTTGCTTCGGCTTCGTGGCTTTTGCATGACTCCAACTGAAAGATTACTTGTT
TATCCCTACATGGCTAATGGAAGTGTTGCTTCTTGTTTAAGAGAGCGTCCTGAAGGCAAT
CCAGCACTTGATTGGCCAAAAAGAAAGCATATTGCTCTGGGATCAGCAAGGGGGCTTGCG
TATTTACATGATCATTGCGACCAAAAAATCATTCACCGGGATGTTAAAGCTGCTAATATA
TTGTTAGATGAAGAGTTTGAAGCTGTTGTTGGAGATTTTGGGCTCGCAAAATTAATGAAT
TATAATGACTCCCATGTGACAACTGCTGTACGCGGTACAATTGGCCATATAGCGCCCGAG
TACCTCTCGACAGGAAAATCTTCTGAGAAGACTGATGTTTTTGGGTACGGGGTCATGCTT
CTCGAGCTCATCACTGGACAAAAGGCTTTCGATCTTGCTCGGCTTGCAAATGATGATGAT
ATCATGTTACTCGACTGGGTGAAAGAGGTTTTGAAAGAGAAGAAGTTGGAAAGCCTTGTG
GATGCAGAACTCGAAGGAAAGTACGTGGAAACAGAAGTGGAGCAGCTGATACAAATGGCT
CTGCTCTGCACTCAAAGTTCTGCAATGGAACGTCCAAAGATGTCAGAAGTAGTGAGAATG
CTGGAAGGAGATGGTTTAGCTGAGAGATGGGAAGAATGGCAAAAGGAGGAGATGCCAATA
CATGATTTTAACTATCAAGCCTATCCTCATGCTGGCACTGACTGGCTCATCCCCTATTCC
AATTCCCTTATCGAAAACGATTACCCCTCGGGTCCAAGATAAcctttagaaagggtctt

ttcttgtgggttcttcaacaagtatatatatagattggtgaagttttaagatgcaaaaaa
aa

Predicted amino acid sequence of the *Arabidopsis thaliana* RKS13 protein.
Different domains are spaced and shown from the N-terminus towards the C-terminus. Overall domain structure is similar as described in Schmidt et al (1997).
At the predicted extracellular domain the first domain represents a signal sequence. The second domain contains leucine zipper motifs, containing 2 times 2 leucine residues, each separated by seven other amino acids. The third domain contains conserved cysteine residues, involved in disulphate bridge formation. The fourth domain contains a leucine rich repeat domain, consisting of 5 complete repeats of each approximately 24 amino acid residues. The fifth domain contains many serine and proline residues, and is likely to contain hydroxy-proline residues, and to be a site for O-glycosylation. The sixth domain contains a single transmembrane domain after which the prodicted intracellular domains are positioned. The seventh domain has an unknown function. The eight domain represents a serine / threonine protein kinase domain (Schmidt et al. 1997) and is probably also containing sequences for protein / protein interactions. The ninth domain has an unknown function. The last and tenth domain at the C-terminal end represents part of a single leucine rich repeat, probably involved in protein / protein interactions.

MEQRSLLCFLYLL
LLFNFTLRVAGNAEG

DALTQLKNSLSSGDP
ANNVLQSWDATLVT

PCTWFHVTCNPENKVTRV

DLGNAKLSGKLV
P ELGQLLNLQYLELYSNNITGEI
PEELGDLVELVSLDLYANSISGPI
PSSLGKLGKLRFLRLNNNSLSGEI
PMTLTSVQLQV LDISNNRLSGDI
PVNGSFSLFTPISFANNSLTDLPE

PPPTSTSPTPPPPSG

GQMTAAIAGGVAAGAAL
LFAVPAIAFAWWL

RRKPQDHFFDVPGAEEDPE
VHLGQLKRFTLRELLVAT

DNFSNKNVLGRGGFGKVYKGRLAD
GNLVAVKRLKEERTKGGELQFQ
TEVEMISMAVHRNLLRLRGFCM
TPTERLLVYPYMANGSVASCLR
ERPEGNPALDWPKRKHIALGSA
RGLAYLHDHCDQKIIHRDVKAA
NILLDEEFEAVVGDFGLAKLMN
YNDSHVTTAVRGTIGHIAPEYL
STGKSSEKTDVFGYGVMLLELI
TGQKAFDLARLANDDDIMLLDW
VKEVLKEKKLESLVDAELEGKY
VETEVEQLIQMALLCTQSSAME
RPKMSEVVRMLE

GDGLAERWEEWQKEEMPIHDFNYQA

YPHAGTDWLIPYSNSLIENDYPSGPR

Arabidopsis thaliana RKS14 cDNA

The start codon encoding the first predicted methionine residue
of the gene product has been indicated by bold capitals.
The first stopcodon has been underlined.
Nucleotides predicted to encode protein sequences are in capitals. Leader and tailer sequences are in lowercase letters.

ctgcaccttagagattaatactctcaagaaaaacaagttttgattcggacaaag**ATG**TTG
CAAGGAAGAAGAGAAGCAAAAAAGAGTTATGCTTTGTTCTCTTCAACTTTCTTCTTCTTC
TTTATCTGTTTTCTTTCTTCTTCTTCTGCAGAACTCACAGACAAAGTTGTTGCCTTAATA
GGAATCAAAAGCTCACTGACTGATCCTCATGGAGTTCTAATGAATTGGGATGACACAGCA
GTTGATCCATGTAGCTGGAACATGATCACTTGTTCTGATGGTTTTGTCATAAGGCTAGAA
GCTCCAAGCCAAAACTTATCAGGAACTCTTTCATCAAGTATTGGAAATTTAACAAATCTT
CAAACTGTATACAGGTTATTGCAGAACAATTACATAACAGGAAACATCCCTCATGAGATT
GGGAAATTGATGAAACTCAAAACACTTGATCTCTCTACCAATAACTTCACTGGTCAAATC
CCATTCACTCTTTCTTACTCCAAAAATCTTCACAGGAGGGTTAATAATAACAGCCTGACA
GGAACAATTCCTAGCTCATTGGCAAACATGACCCAACTCACTTTTTTGGATTTGTCGTAT
AATAACTTGAGTGGACCAGTTCCAAGATCACTTGCCAAAACATTCAATGTTATGGGCAAT
TCTCAGATTTGTCCAACAGGAACTGAGAAAGACTGTAATGGGACTCAGCCTAAGCCAATG
TCAATCACCTTGAACAGTTCTCAAAGAACTAAAAACCGGAAAATCGCGGTAGTCTTCGGT
GTAAGCTTGACATGTGTTTGCTTGTTGATCATTGGCTTTGGTTTTCTTCTTTGGTGGAGA
AGAAGACATAACAAACAAGTATTATTCTTTGACATTAATGAGCAAAACAAGGAAGAAATG
TGTCTAGGGAATCTAAGGAGGTTTAATTTCAAAGAACTTCAATCCGCAACTAGTAACTTC
AGCAGCAAGAATCTGGTCGGAAAAGGAGGGTTTGGAAATGTGTATAAAGGTTGTCTTCAT
GATGGAAGTATCATCGCGGTGAAGAGATTAAAGGATATAAACAATGGTGGTGGAGAGGTT
CAGTTTCAGACAGAGCTTGAAATGATAAGCCTTGCCGTCCACCGGAATCTCCTCCGCTTA
TACGGTTTCTGTACTACTTCCTCTGAACGGCTTCTCGTTTATCCTTACATGTCCAATGGC
AGTGTCGCTTCTCGTCTCAAAGCTAAACCGGTATTGGATTGGGGCACAAGAAAGCGAATA
GCATTAGGAGCAGGAAGAGGGTTGCTGTATTTGCATGAGCAATGTGATCCAAAGATCATT
CACCGTGATGTCAAAGCTGCGAACATACTTCTTGACGATTACTTTGAAGCTGTTGTCGGA
GATTTCGGGTTGGCTAAGCTTTTGGATCATGAGGAGTCGCATGTGACAACCGCCGTGAGA
GGAACAGTGGGTCACATTGCACCTGAGTATCTCTCAACAGGACAATCTTCTGAGAAGACA
GATGTGTTCGGTTTCGGGATTCTTCTTCTCGAATTGATTACTGGATTGAGAGCTCTTGAA
TTCGGAAAAGCAGCAAACCAAAGAGGAGCGATACTTGATTGGGTAAAGAAACTACAACAA
GAGAAGAAGCTAGAACAGATAGTAGACAAGGATTTGAAGAGCAACTACGATAGAATAGAA
GTGGAAGAAATGGTTCAAGTGGCTTTGCTTTGTACACAGTATCTTCCCATTCACCGTCCT
AAGATGTCTGAAGTTGTGAGAATGCTTGAAGGCGATGGTCTTGTTGAGAAATGGGAAGCT
TCTTCTCAGAGAGCAGAAACCAATAGAAGTTACAGTAAACCTAACGAGTTTTCTTCCTCT

GAACGTTATTCGGATCTTACAGATGATTCCTCGGTGCTGGTTCAAGCCATGGAGTTATCA
GGTCCAAGATGAcaagagaaactatatgaatggctttgggtttgtaaaaaa

Predicted amino acid sequence of the *Arabidopsis thaliana* RKS14 protein.
Different domains are spaced and shown from the N-terminus towards the C-terminus. Overall domain structure is similar as described in Schmidt et al (1997).
At the predicted extracellular domain the first domain represents a signal sequence. The second domain contains a leucine zipper motif, containing 3 leucine residues, each separated by seven other amino acids. The third domain contains conserved cysteine residues, involved in disulphate bridge formation. The fourth domain contains a leucine rich repeat domain, consisting of 5 complete repeats of each approximately 24 amino acid residues. The fifth domain contains many serine and proline residues, and is likely to contain hydroxy-proline residues, and to be a site for O-glycosylation. The sixth domain contains a single transmembrane domain after which the prodicted intracellular domains are positioned. The seventh domain has an unknown function. The eight domain represents a serine / threonine protein kinase domain (Schmidt et al. 1997) and is probably also containing sequences for protein / protein interactions. The ninth domain has an unknown function. The last and tenth domain at the C-terminal end represents part of a single leucine rich repeat, probably involved in protein / protein interactions.

MLQGRREAKKSYALFSSTFF
FFFICFLSSSSAELTDKV

VALIGIKSSLTDP
HGVLMNWDDTAVD

PCSWNMITCSDGFVIR

LEAPSQNLSGTLSS

SIGNLTNLQTVYRLLQNNYITGNI

PHEIGKLMKLKTLDLSTNNFTGQI

PFTLSYSKNLHRRV NNNSLTGTI

PSSLANMTQLTFLDLSYNNLSGPV

PRSLAKTFNVMGNSQICPT

GTEKDCNGTQPKPMSITLNSSQR

TKNRK

IAVVFGVSLTCVCLLIIGFGFLLWW

RRRHNKQVLFFDINEQNKE

EMCLGNLRRFNFKELQSAT

SNFSSKNLVGKGGFGNVYKGCLHD

GSIIAVKRLKDINNGGGEVQFQ

TELEMISLAVHRNLLRLYGFCT

TSSERLLVYPYMSNGSVA

SRLKAKPVLDWGTRKRIALGAG

RGLLYLHEQCDPKIIHRDVKAA

NILLDDYFEAVVGDFGLAKLLD

HEESHVTTAVRGTVGHIAPEYL

STGQSSEKTDVFGFGILLLELI

TGLRALEFGKAANQRGAILDW

VKKLQQEKKLEQIVDKDLKSNY

DRIEVEEMVQVALLCTQYLPIH

RPKMSEVVRMLE

GDGLVEKWEASSQRAET

NRSYSKPNEFSSS

ERYSDLTDDSSVLVQAMELSGPR

Legends

Figure 1

The different domains of the predicted RKS gene product have the following functions:
The first domain of the predicted protein structure at the N-terminal end consists of a signal sequence, involved in targeting the protein towards the plasma membrane. Protein cleavage removes this sequence from the final mature protein product (Jain et al. 1994, J. Biol. Chemistry 269: 16306-16310). The second domain consists of different numbers of leucine zipper motifs, and is likely to be involved in protein protein dimerization. The next domain contains a conserved pair of cystein residues, involved in disulphate bridge formation. The next domain consists of 5 (or in the case of RKS3 only 4) leucine rich repeats (LRRs) shown in a gray colour, likely to be involved in ligand binding (Kobe and Deisenhofer 1994, TIBS 19: 415-420). This domain is again bordered by a domain containing a conserved pair of cystein residues involved in disulphate bridge formation often followed by a serine /. proline rich region. The next domain displays all the characteristics of a single transmembrane domain (http://genome.cbs.dtu.dk/services/TMHMM/). At the predicted cytoplasmic site of protein a domain is situated with unknown function, followed by a domain with serine /threonine kinase activity (Schmidt et al. 1997, Development 124: 2049-2062). The kinase domain is followed by a domain with unknown function whereas at the C-terminal end of the protein part of a leucine rich repeat is positioned, probably involved in protein-protein interactions.

Figure 2

Alagnment of the predicted protein sequences of the different RKS gene products from Arabidopsis thaliana with alignX, Vector NTI Suite 5.5 resulted in a phylogenetic tree in which

the relative homology between the different RKS members is shown.

Figure 3

Intron-Exon bounderies of the genomic regions on the chromosomes of *Arabidopsis thaliana* encoding the different RKS gene products. Exons are shown as boxes, whereas intron sequences are shown as lines. Sequences encoding LRR domains are displayed in gray colour, transmembrane regions in black.

Figure 4.

Cromosomal location of RKS genes in Arabidopsis thaliana

Figure 5. A signaling complex comprising molecules of RKS proteins, ELS proteins, NDR/NHL proteins and SBP/SPL proteins.

Figure 6.

Second generation (T2) tobacco seedlings germinated on MS medium. Transformations were performed with DNA clone 2212-15, representing the overexpression construct GT-RKS4-s. T2 seedlings derived from
T1 plant 15.7 shows co-suppression effects while T1 plant 15.6 shows no obvious changes in level of RKS4. T1 plants 15.9 and 15.3 show overexpression effects. Plant 15.7 has the lowest remaining level of RKS4 gene product, whereas plant 15.3 has the highest level of RKS4 gene product.

Figure 7

Second generation (T2) tobacco plants. In the upper row the offspring from a co-suppressing T1 plant 15.7 is shown. The middle row shows plants derived from a transgenic T1 plant 15.6 with no clear changes in level of RKS4 is shown while the bottom row shows plants derived from a T1 plant 15.3 in which the
levels of RKS4 are increased by the introduction of the overexpression construct GT-RKS4-s.

Figure 8

Second generation (T2) tobacco plants. Plants derived from a co-suppressing T1 plant 15.7 show a reduction in plant size and a delay in the initiation and outgrowth of primordia. The control empty vector transgenic plants show no visible differences in growth compared with the offspring from the transgenic 15.6 plant, in which the endogenous level of RKS4 gene product was not changed. In the overexpressing plants 15.9 and 15.3 organ size was increased, similar as the number of initiated leaf primordia.

Figure 9

Arabidopsis thaliana WS plants in which the endogenous level of RKS4 gene product is decreased (right picture) due to the presence of a transgenic RKS4 antisense construct (GT-RKS4-16a). The left picture shows a wildtype plant of the same age as the transgenic antisense plant, grown under similar growth conditions. Plant size, organ size and number of organ primordia is decreased in the transgenic antisense plant compared with the wildtype control.

Figure 10

Arabidopsis thaliana WS plants in which the endogenous level of RKS4 gene product is decreased (right picture) due to the presence of a transgenic RKS4 antisense construct (GT-RKS4-16a). The left picture shows a wildtype flower of the same age as the transgenic antisense flower, grown under similar growth conditions. Total flower size is only slightly decreased in the transgenic antisense flower compared with the control flower, whereas organ size of petals is strongly decreased.

Figure 11

Arabidopsis thaliana WS plants in which the endogenous level of RKS4 gene product is increased (right picture) due to the presence of a transgenic RKS4 overexpressing construct (GT-RKS4-6s). The left picture shows a wildtype flower of the same age as the transgenic antisense flower, grown under similar

growth conditions. Compared with the wildtype control flower, total flower size of the transgenic flower is clearly increased. Both sepal and petal organ size is clearly increased compared with the control.

Figure 12
Arabidopsis thaliana WS plants in which the endogenous level of RKS4 gene product is modulated due to the presence of a transgenic RKS4 construct. The left picture shows a wildtype flower of the same age as the transgenic flowers, grown under similar growth conditions. Compared with the wildtype control flower, total flower size of the transgenic RKS4 overexpressing flower (middle) is clearly increased. Both sepal and petal organ size is clearly increased compared with the control. In *Arabidopsis thaliana* WS plants in which the endogenous level of RKS4 gene product is decreased (right picture) due to the presence of a transgenic RKS4 antisense construct, total flower size is decreased compared with the control.

Figure 13
Organ size can be influenced by either modulating cell division or cell elongation or a combination of both. In order to identify the total number of cells and the cell size within an organ the apical site of petals of mature *Arabidopsis* flowers was investigated. Petal organ size is clearly influenced by modulation of RKS4 gene product levels (bottom row for the flowers from which the apical petal epidermal cells were identified). Epidermal cell size is not changed in transgenic plants compared with the control.

Figure 14
Arabidopsis thaliana WS plants in which the endogenous level of RKS10 gene product is increased (right picture) due to the presence of a transgenic RKS10 overexpressing construct. The left picture shows the apical epidermus of a full grown cotyl from an empty vector transgenic seedling of the same age as

the transgenic overexpressing cotyl, grown under similar growth conditions..

Figure 15

Arabidopsis thaliana WS plants in which the endogenous level of RKS10 gene product is decreased (right picture) due to the presence of a RKS10 antisense construct The left picture shows a wildtype plant of the same age as the transgenic antisense plant, grown under similar growth conditions. Plant size, organ size and number of organ primordia remains similar in both the transgenic antisense plants and the wildtype control.

Figure 16

In order to determine organ size varietions in transgenic RKS10 transgenic plants compared with empty vector control transgenic plants (pGreen4K), flower organ size was determined of the four
open flower stages of *Arabidopsis* influorescenses. The four successive flower stages are photographed under similar magnifications. No obvious changes in organ length could be observed in size of sepals, petals, stamen and carpel between empty vector control flowers (pGreen4K), flowers with an antisense
RKS10 construct (a) or plants overexpressing the RKS10 cDNA under the control of a 35S promoter (S

Figure 17

Tissue cultured auxin treated *transgenic Arabidopsis* T2 seedlings were grown on MS agar plates without hormones for a period of 3 weeks. Regeneration potential was scored and the formation and outgrowth of multiple shoot apical meristems from single seedling origin was displayed as (+). The formation and outgrowth of only one shoot apical meristem, leading to the formation of a normal rosette of leaves from individual plants was displayed as (-). Postive regeneration controls consisted of seedlings overexpressing either KNAT1,

CUC2, IPT or cycD3. All of these showed an increase of regeneration capacity (+) compared with a negative control GUS overexpressing plant pGreen5K (-).
Representative examples of RKS and ELS cDNA overexpressing (s) or antisense (a) cosuppressing constructs in transgenic plants are shown in the bottem panels.

Figure 18.
Tobacco leaf discs were stably transformed with the RKS0 overexpressing construct GT-RKS0-23S and from a single transformation event, large numbers of regeneration plantlets were isolated and subcultured. All of the regenerated plants were potted and flowered. The original transformation event could be kept continuously in tissue culture indefinitely.

Figure 19
Seedlings from transgenic Arabidopsis thaliana containing either constructs overexpressing (s) or co-suppressing by antisense (a) the RKS gene products were screened for the appearance of fasciation. Several examples in which fasciation could be routinely observed are shown together with a negative control plant (pGreen5K, overexpressing the GUS gene) in which fasciation could never be observed.

Figure 20 - 23
Primary root tips of transgenic Arabidopsis plants (top rows) photographed under similar magnification. The bottom rows show the corresponding seedlings (also between each other under the same magnification). Figure 23 shows the specific Arabidopsis transgenes with a strong increase in root outgrowth.

Figure 24
Avarage root length of 10-30 transgenic Arabidopsis T2 seedlings from one T1 transgenic plant is shown.

Figure 25

T3 seedlings are shown from a strong co-suppressing RKS10 antisense construct line (T1-4; T2-6; T3 generation) and a strong overexpressing line (T1-4; T2-6; T3 generation). The overexpressing line is different and stronger from the one shown in Figure 4.1-4.5. Pictures are taken under similar magnifications.

Figure 26

T2 seed was germinated on horizontal MS agar plates and pictures were taken under similar magnification of representative examples of the lateral root development from transgenic RKS and ELS transgenic roots.

Figure 27

Pictures taken from transgenic RKS8 or RKS10 overexpressing roots taken directly behind the tip zone. Pictures are taken under same magnification.

Figure 28

Arabidopsis thaliana WS plants in which the endogenous level of RKS or ELS gene product is modified result in the formation of new meristem formation and / or outgrowth, resulting in a complex, bushy influorescense in transgenic Arabidopsis plants conmpared with control empty vector control plants (pGreen4K). Overexpression of RKS10 and ELS1 (S) and cosuppression with antisense constructs of RKS8 and also RKS10, result in increased numbers of developing generative meristems. The generative shoots are photographed with similar maginification.

Figure 29

Arabidopsis thaliana WS plants in which the endogenous level of RKS gene product is modified result in the formation of new meristem formation and / or outgrowth, resulting in a complex, bushy influorescense in transgenic Arabidopsis plants conmpared with control empty vector control plants (pGreen4K). The top panel shows adult plants under similar maginification.

Compared with the control, RKS10 overexpression results in an extreme bushy phenotypic plant. The results of co-suppressing the RKS8 gene product are less dramatic with respect to the bushiness. However, also in these transgenic plants the number of generative meristems is strongly increased compared with the control. The bottem panel shows the generative shoot in detail under similar magnification.

Figure 30

Scematic drawing of the different flower organs in an empty vector control pGreen4K flower (left) compared with a complex transgenic flower structure seen in transgenic Arabidopsis plants containing an antisense (a) RKS10 construct. The terminal flower meristem produces 2 sepals, 1 petal, 2 stamen, a carpel which is not a closed structure but open with groups of ovules on the inside and outside of this structure, and stigmatic cells protruding from the top part. Two new flowers are protruding from this structure, containing all flower organs in normal numbers.

Figure 31

Scematic drawing of the different flower organs in a complex transgenic flower structure seen in transgenic Arabidopsis plants T1-11 containing an antisense (a) RKS10 construct. The terminal flower meristem produces 1 sepal, 2 petals, 2 stamen, a carpel which is not a closed structure but open with groups of ovules on the inside and outside of this structure, and stigmatic cells protruding from the top part. An indetermined flower meristem is protruding from the open carpel structure and forms a number of new flowers, including normal flowers (right) and another abnormal flower (left) which consists of a flower with half of the sepal, petal and stamen organs formed and a new terminal flower mersitem protruding from this structure, developing in structures as seen in Figure 7.5. The stamen contain only small numbers of (viable) pollen compared with wildtype stamen (see also chapter 5).

Figure 32

Scematic drawing of the different flower organs in an empty vector control pGreen4K flower (left) compared with a complex transgenic flower structure seen in a transgenic *Arabidopsis* plant T1-11 containing an antisense (a) RKS10 construct (overview shown in Figure 7.4). The terminal flower meristem produces half the normal number of sepals, petals and stamen. The remaining part of the flower structure has converted into a new structure containing a new stem containing a single organ structure resembling a fusion between a petal and a sepal. On this structure seveal (viable) pollen grains can be observed.

Figure 33

Schematic drawing of the different flower organs in a complex transgenic flower structure seen in a transgenic *Arabidopsis* plant T1-12 containing an antisense (a) RKS10 construct. The terminal flower meristem originating from an indetermined generative meristem is here producing an axillary secondary indetermined meristem (left picture), a single organ resembling a stamen (bottem left), a normal flower and a terminal flower. This terminal flower structure contains 2 normal sepals, 2 normal petals, 2 normal stamen (with only a few viable pollen) and two organs resembling a fusion of sepals /petals/stamen (see also figure 7.7). From this terminal flower structure two new flowers emerge (in a similar fashion as observed in Figure 7.3) containing normal numbers of flower organs (right photos). At the top of this figure a control influorescense is shown schematically with terminal flower meristems as normally origninate from the generative Arabidopsis thaliana generative meristem.

Figure 34

Scematic drawing and detailed pictures of several of the structures as shown in figure 7.6. At the right the organs resembling a fusion between sepals/petals/stamen are shown with viable pollen sticking out from these structures. At the

top left the single stamen-like organ directly protruding from the main stem is shown.

Figure 35

Transgenic Arabidopsis plants overexpressing the RKS13 gene product show a modification of the normal flower influorescense architechture, somewhat resembling the structures observed in RKS10 antisense plants. A terminal flower containing a normal seed developing silique and a small number of sepals, petals and stamen, develops at least 4 additional terminal flower meristems that develop abnormally themselves, resulting in open carpel structures and modifications of organ structures.

Figure 36

Transgenic plants in which the RKS and / or ELS genes are introduced behind a constitutive 35S promoter in an overexpressing (S) or antisense (a) configuration are analysed for sterility and characterised further for defects in proper pollen development. As a negative control the normal pollen development of a transgene containing the empty expression vector (pG4K) was included. First generation transgenic flowers of RKS10 expressing constructs and second generation control vector and ELS2 are shown under similar magnification. In detail the stigmatic surface and surrounding stamen, are shown under similar magnification, showing the presence or absense of pollen on the stamen or the stigmatic surface.

Detailed description

1.Modifying organ size

Plant size is determined by both cell elongation and cell division rate. Modifying either one or both processes results in a change in final organ size. Increasing the level of specific members of the family of RKS genes results in an increase in organ size, growth rate and yield. Modulating plant growth, organ size and yield of plant organs is the most important process to be optimized in plant performance. Here we show that modulating the level of members of the family of the RKS signaling complex is sufficient to modulate these processes. The invention provides herewith a method for modulating a developmental pathway of a plant or plant cell comprising modifying a gene or modifying expression of said gene, wherein said gene is encoding a protein belonging to a signaling complex comprising RKS protein, ELS protein, NDR/NHL protein, SBP/SPL protein and RKS/ELS ligand protein allowing modulating cellular division during plant growth or organ formation, in particular wherein said gene comprises an RKS4 or RKJS 10 gene or functional equivalent thereof. Inactivation of endogenous RKS gene product results in a decrease in plant growth, proving that the normal function of these endogenous RKS gene products is the regulation of growth and organ size. Elevation of the levels of the regulating of the RKS signaling complex in plant cells is provided in order to increase:
the size of plant organs
the growth rate
the yield of harvested crop
the yield of total plant material
the total plant size

Decreasing the levels of endogenous RKS gene product is provided in order to decrease:
the size of plant organs

the growth rate
the total plant size

Results obtained (see also figures 6 to 13)

Overexpression and antisense constructs of full length RKS cDNA clones have been made under the control of 35S promoters. Transgenic plants have been produced in *Arabidopsis thaliana* and in *Nicotiana tabacum*. Subsequent generations of stably transformed plants were investigated for phenotypes and analysed in detail. The phenotype observed in transgenic plants with antisense constructs of RKS4 (GT-RKS4-a) could be described as dwarf plants in which all plant organs showed a decrease in organs size and growth rate. Overexpression of RKS4 (GT-RKS4-s) resulted in plants with increased size of organs and an increase in growth rate Since cell size alone was not responsible for the modifications in organ size of petals it can be concluded that RKS4 is involved in the regulation of the cellular divisions during plant growth and organ formation. Overexpression of RKS 4 results in an increase of cellular divisions whereas a decrease in endogenous RKS 4 gene product levels within the plant results in a decrease of cellular division rates.

Literature

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-Plant organ size control: Aintegumenta regulates growth and cell numbers during organogenesis. Y. Mizukami and R.L. Fischer PNAS 97: 942-947

-Measuring dimensions: the regulation of size and shape. S.J. Day and P.A.Lawrence 2000; Development 127: 2977-2987

-A matter of size: developmental control of organ size in plants. Y. Mizukami 2001; Current opinions in plant biology 4: 533-539

2. Cell division

The mitotic cell cycle in eukaryotes determines the total number of cells within the organism and the number of cells within individual organs. The links between cell proliferation, cell differentiation and cell-cycle machinery are of primary importance for eucaryotes, and regulation of these processes allows modifications during every single stage of development. Here we show that modulating the level of members of the family of the RKS signaling complex is sufficient to modulate these processes. The invention provides herewith a method for modulating a developmental pathway of a plant or plant cell comprising modifying a gene or modifying expression of said gene, wherein said gene is encoding a protein belonging to a signaling complex comprising RKS protein, ELS protein, NDR/NHL protein, SBP/SPL protein and RKS/ELS ligand protein allowing modulating cellular division during plant growth or organ formation, in particular wherein said gene comprises an RKS4 or RKJS 10 gene or functional equivalent Herewith the invention provides a method for modulating the number of cells to be formed within an eukaryotic organism as a whole or for modulating the cell number within individual organs is, which of primary importance in modulating plant developmental processes, especially of arable plants. Here we show that members of the RKS signaling complex are able to regulate the number of cellular divisions, thereby regulating the total number of cells within the organism or different organs.

Possible Applications

Elevation of the levels of the regulating RKS signaling complex members in plant cells in order to increase:

the size of plant organs

the growth rate

the yield of harvested crop

the yield of total plant material

the total plant size

Decreasing the levels of endogenous RKS signaling complex members in order to decrease:
the size of plant organs
the growth rate
the total plant size

Results obtained

Overexpression and antisense constructs of full length RKS cDNA clones have been made under the control of 35S promoters. Transgenic plants have been produced in *Arabidopsis thaliana* and in *Nicotiana tabacum*. Subsequent generations of stably transformed plants were investigated for phenotypes and analysed in detail.

Overexpression of RKS 4 results in an increase of cellular divisions whereas a decrease in endogenous RKS 4 gene product levels within the plant results in a decrease of cellular division . Another example of RKS genes involved in cellular proliferation is provided by RKS10. Overexpression of RKS10 (S) results in a decrease in apical epidermal cells (Figure 14) compared with control plants containing an empty expression casette (pGreen4K). Co-suppressing the endogenous RKS 10 gene in plants containing an antisense construct (a) showed clearly larger epidermal cells as the corresponding cells in wildtype control plants (Figure 15). In contrast to the plant phenotypes shown in RKS4 transgenic plants , no differences in plant or organ size could be observed in the RKS10 transgenic plants or organs. This shows that although the organ size remains constant, the number of cells within these organs is variable due to the differences in size of individual cells. These results indicate that normal RKS4 function within the plant can be described as an activator of cellular division.

Normal RKS10 function also involves an activation process on cellular division rate. This effect is also detectable in the root in the region directly behind the tip zone, where in the RKS10 overexpressing transgenes cellular divisions were

detectable in a region where normally cell proliferation has ceased. The plane of divisions of root cells in these transgenes is also clearly different from the normal plane of root cell division, resulting in clumps of cells with all types of division planes possible.

In contrast to RKS4, the final organ size in RKS10 transgenic plants is under the control of other organ size restriction processes, in such a way that the final organ volume remains constant (Figure 16). RKS4 and RKS10 are essentially involved in the same cell cycle activation process, but either addition organ size controlling functions of these RKS genes or the hierarchical order in which they regulate the cell cycle is different.

Literature

-Not being the wrong size. R.H. Gomer 2001; Nature reviews 2: 48-54

-Cell cycling and cell enlargement in developing leaves of Arabidopsis. P.M Donnelly et al. 1999; Developmental biology 215: 407-419

-When plant cells decide to divide. H. Stals and D. Inze 2001. Trends in Plant Science 6: 359-363

-Cell cycling and cell enlargement in developing leaves of Arabidopsis. P.M. Donnelly et al. 1999. Developmental Biology 215: 407-419

-Triggering the cell cycle in plants. B.G.W. den Boer and J.A.H. Murray 2000. Trends in Cell Biology 10: 245-250

3.Regeneration

Modification the levels of different RKS and ELS genes within plants allows the initiation and / or outgrowth of apical meristems, resulting in the formation of large numbers of plantlets from a single source. A number of gene products that is able to increase the regeneration potential of plants is known already. Examples of these are KNAT1, cycD3, CUC2 and IPT. Here we show that modulation of the endogenous levels of RKS genes results in the formation of new shoots and plantlets in different plant species like *Nicotiana tabacum* and *Arabidopsis thaliana*. herewith the invention provides a method for modulating a developmental pathway of a plant or plant cell comprising modifying a gene or modifying expression of said gene, wherein said gene is encoding a protein belonging to a signaling complex comprising RKS protein, ELS protein, NDR/NHL protein, SBP/SPL protein and RKS/ELS ligand protein, allowing modulating apical meristem formation, in particular wherein said gene comprises an ELS1, RKS0, RKS3, RKS4, RKS8 or RKS10 gene or functional equivalent thereof. A direct application of a method according to the invention is the stable or transient expression of RKS and ELS genes or gene products in order to initiate vegetative reproduction. Regeneration can be induced after overexpression of for example RKS0 and ELS1; or by co-suppression of for example the endogenous RKS3, RKS4, RKS8 or RKS10 genes. Overexpression or co-suppression of these RKS and ELS gene products can be either transient, or stable by integration of the corresponding expression casettes in the plant genome.

Results obtained

Overexpression and antisense constructs of full length RKS and ELS cDNA clones have been made under the control of 35S promoters. Transgenic plants have been produced in *Arabidopsis thaliana* and in *Nicotiana tabacum*. Subsequent generations of

stably transformed plants were investigated for phenotypes and analysed in detail.

T2 transgenic seedlings of Arabidopsis were germinated in liquid MS medium supplemented with 1 mg/L 2,4-D for 1 week, followed by extensive washing and plating of the seedlings onto MS agar plates without hormones. Control transgenic seedstocks containing either a negative control vector (pGreen5K); or positive control overexpression constructs of gene products known to increase the regeneration potential (IPT, KNAT1, CUC2 and cycD3) were characterized for regeneration potential together with seedstocks from plants either overexpressing (s) or co-suppressing (a) all RKS and ELS gene products (Figure 17). Overexpression of the ELS1 and RKS0 cDNA clones resulted in an increase of shoot apical meristem formation and outgrowth, whereas antisense constructs (a) of these cDNA clones did not increase the regeneration potential (only increased regeneration results are shown). Antisense constructs of RKS3, RKS4, RKS8 and RKS10 also resulted in an increased formation and outgrowth of apical meristems (Figure 17).

T1 generation Nicotiana tabacum tissue cultures transformed with ELS and RKS gene products in either overexpression (s) casettes or antisense co-suppression (a) casettes allowed the regeneration of indefinite number of offspring plants from a single transformed cell origin (Figure 18). An example is shown for the overexpression of the GT-RKS0-23S construct. The resulting plants obtained from one transformation event in general showed no phenotypes. Only a subset of plants displayed RKS0 overexpression phenotypes (like loss of apical dominance and early flowering).

Literature

-Mechanisms that control knox gene expression in the Arabidopsis shoot. N. Ori et al. 2000,Development 127: 5523-5532

-Overexpression of KNAT1 in lettuce shifts leaf determinate growth to a shoot-like indeterminate growth associated with an accumulation of isopentenyltype cytokinins. G. Frugis et al. 2001. Plant Physiology 126: 1370-1380

-KNAT1 induces lobed leaves with ectopic meristems when overexpressed in Arabidopsis. Chuck et al. 1996. the Plant Cell 8: 1277-1289

-Cytokinin activation of Arabidopsis cell division through a D-type cyclin. C. Riou-Khamlichi et al. 1999. Science 283: 1541-1544

4. Fasciation

Fasciation is normally a result from an increased size of the apical meristem in apical plant organs.
Modulation of the number of cells within the proliferating zone of the shoot apical meristem results in an excess number of cellular divisions, giving rise to excess numbers of primordia formed or to stems in which the number of cells is increased. AThe inventiuon herewith provides a method for modulating a developmental pathway of a plant or plant cell comprising modifying a gene or modifying expression of said gene, wherein said gene is encoding a protein belonging to a signaling complex comprising RKS protein, ELS protein, NDR/NHL protein, SBP/SPL protein and RKS/ELS ligand protein allowing modulating fasciation, in particular wherein said gene comprises an RKS0, RKS3, RKS8 or RKS10 gene or functional equivalent thereof.Here we for example show that modulation of the levels of RKS gene products in plants like *Arabidopsis thaliana* can result in fasciated stems as shown in Figure 19. A direct application as provided herein is the regulated formation of fasciation in plant species in which such a trait is desired like ornamental plants. Regulationof the initiation and extent of fasciation, either by placing the responsible RKS encoding DNA sequences under the control of stage or tissue specific promoters, constitutive promoters or inducible promoters results in plants with localized or consitutive fasciation of stem tissue. Another application is modulating the number of primordias by regulation of the process of fasciation. An example is provided by for example sprouts, in which an increased number of primordia will result in an increased numbers of sprouts to be harvested. Fasciation can also result in a strong modification in the structural architecture of the influorescence, resulting in a terminal group of flowers resembling the *Umbelliferae* type (an expample is shown in Figure 19 where the fasciated meristem of a RKS0-

7S overexpressing *Arabidopsis* plant clearly terminates in an *Umbelliferae* type influorescense.

Results obtained

Overexpression and antisense constructs of full length RKS cDNA clones have been made under the control of 35S promoters. Transgenic plants have been produced in *Arabidopsis thaliana*. Subsequent generations of stably transformed plants were investigated for phenotypes and analysed in detail. T2 transgenic seedlings of *Arabidopsis* were germinated on MS agar plates without hormones. Control transgenic seedstocks containing a negative control vector (pGreen5K) were tested for their ability to induce fasciation (Overexpression constructs (s) of RKS0, RKS8 and RKS10 cDNA clones resulted in fasciated plants, whereas antisense constructs (a) of these cDNA clones did not increase the regeneration potential (only positive results are shown). Antisense constructs of RKS3 gave also rise to fasciation (Figure 19).

Literature

-Functional domains in plant shoot meristems. U. Brand et al. 2001. Bioassays 23: 134-141.

-Dependence of stem cell fate in Arabidopsis on a feedback loop regulated by CLV3 activity.
U. Brand et al. 2000. Science 289: 617-619

5. Root development

Fasciation is normally a result from an increased size of the apical meristem in apical plant organs. Modulation of the number of cells within the proliferating zone of the root apical meristem results in an excess number of cellular divisions, giving rise to excess numbers of primordia formed or to roots in which the number of cells is increased. Adaptation to soil conditions is possible by regulation of root development of plants. Here we describe several processes in root development that can be manipulated by modification of the levels of the RKS signalling complex within the root. The invention provides a method for modulating a developmental pathway of a plant or plant cell comprising modifying a gene or modifying expression of said gene, wherein said gene is encoding a protein belonging to a signaling complex comprising RKS protein, ELS protein, NDR/NHL protein, SBP/SPL protein and RKS/ELS ligand protein allowing modulating root development, in particular wherein said gene comprises an ELS1, ELS2, RKS1, RKS3, RKS4, RKS6 RKS8 or RKS10 gene or functional equivalent thereof. Root length, a result by either root cells proliferation or elongation, can for example be increased by overexpression of for example RKS3, RKS4, RKS6 and ELS2, or inactivation of the endogenous RKS10 gene product. Root length can also be decreased by decreasing of endogenous RKS1 levels or by strong overexpression of RKS10.The initiation of lateral roots is also regulated by RKS gene products. Overexpression of for example RKS10 can result in a strong increase in the initiation and outgrowth of lateral roots. Co-suppression of RKS1 also resulted in the initiation and outgrowth of large numbers of lateral roots. Root hair formation and elongation is important in determining the total contact surface between plant and soil. A strong increase of root hair length (elongation) can be obtained by overexpression of ELS1 and RKS3 gene products. As the roots of terrestrial plants are involved in the acquisition of water and nutrients, anchorage of the plant, synthesis of plant hormones, interaction with

the rhizosphere and storage functions, increasing or decreasing root length, for example for flexable adaptations to different water levels, can be manipulated by overexpressing or cosuppressing RKS and / or ELS gene products. Modulation of the total contact surface between plant cells and the outside environment can be manipulated by regulation lateral root formation (increased by RKS10 overexpression and co-suppression of RKS1). Finally the contact surface between plant cells and the soil can be influenced by modulation of the number of root hairs formed or the elongation of the root hairs, as mediated by ELS1 and RKS3.

Results obtained

Overexpression and antisense constructs of full length RKS cDNA clones have been made under the control of 35S promoters. Transgenic plants have been produced in *Arabidopsis thaliana*. Subsequent generations of stably transformed plants were investigated for phenotypes and analysed in detail. T2 transgenic seedlings of *Arabidopsis* were germinated on MS agar plates without hormones. Control transgenic seedstocks containing a negative control vector pGreen4K (empty expression vector) and / or pGreen5K (a GUS overproducing vector) were included as references for normal root development. Seedlings from transgenic Arabidopsis thaliana containing either constructs overexpressing (s) or co-suppressing by antisense (a) the RKS gene products were screened for the appearance of fasciation. Several examples in which fasciation could be routinely observed are shown together with a negative
control plant (pGreen4K, containing an expressing casette without an insert cDNA). Seedlings are germinated and grown on vertically placed MS agar plates.

Literature

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-The development of plant roots: new approaches to underground problems. J.W. Schiefelbeim and P.N. Benfey 1991. the Plant Cell 3: 1147-1154

6. Apical meristems

All parts of the plant above the ground are generally the result on one apical shoot meristem that has been initiated early at embryogenesis and that gives rise to all apical organs. This development of a single meristem into complex tissue and repeated patterns is the result of tissue and stage-dependent differentiation processes within the meristems and its resulting offspring cells. The control of meristem formation, meristem identity and meristem differentiation is therefore an important tool in regulating plant architecture and development. Here we present evidence the function of RKS and ELS gene products in regulation of the meristem identity and the formation and outgrowth of new apical meristems. The invention provides a method for modulating a developmental pathway of a plant or plant cell comprising modifying a gene or modifying expression of said gene, wherein said gene is encoding a protein belonging to a signaling complex comprising RKS protein, ELS protein, NDR/NHL protein, SBP/SPL protein and RKS/ELS ligand protein allowing modulating meristem identity, in particular wherein said gene comprises an ELS1, RKS8, RKS10 or RKS13 gene or functional equivalent thereof. Introduction of for example the RKS10 gene product or an other member of the RKS signalling complex under the control of a tissue and / or stage specific promoter as provided herein allows localized and time regulated increases in the levels of gene product. For example the meristematic identity in a determined meristem might thereby be switched back into an indetermined meristem, thereby changing for example a terminal flower into an indetermined generative meristem.

Another application might be found in changing the meristematic identity at an early time point, during early vegetative growth, thereby switching the vegetative meristem into a generative meristem, allowing early flowering.

Modulation of meristem identity in terminal primordia, like for example as shown in Figure 30, where flower organ primordia are converted into terminal flower primordia, allows

the formation of completely new types of flowers and fused fruitstructures. Constitutive overexpression of RKS gene products results in plants with many apical meristems, as can clearly been seen in Figure 29, where RKS10 overexpression results in an extremely bushy phenotype.

Results obtained

Changing the normal levels of endogenous RKS10 within the plant, either by overexpressing or co-suppressing the RKS10 cDNA, results in an increase in generative meristem development (Figure 28).

Compared with the control empty vector transgenic pGreen4K plants, large number of meristems are initated at places were normally no meristems initiate and / or develop. A clear example is shown by co-suppressing the RKS8 gene (Figure 29), where many new influorescence meristems are initiated from the central generative meristem compared with control pGreen4K plants of the same age. This phenotype is even more extreme in RKS10 overexpressing plants where the resulting plants are extremely bushy with very large numbers of generative meristems formed. Inactivation of the endogenous RKS10 gene in Arabidopsis results in modification of meristematic identity as can be shown in Figure 30. A determined flower meristem develops into two new normal terminal flower meristems and a number of terminal flower organ primordia. Another example is shown in Figure 31 where meristem determination is switched from a terminal flower meristem, that normally result only in the normal numbers of terminal organ primordia, towards a number of organ primordia, a new indetermined generative meristem that develop into normal flowers or in a new terminal flower meristem with developmental abnormalities. Only half of the terminal flower primordia develop normally while an extra structure arises resembling a new flower stem with a petal/stamen like organ. The few pollen detectable on this structure (Figure 32) were able to pollinate a MS1 (male sterile) arabidopsis flower. Figure 33 shows the meristematic

developmental switch from a terminal flower meristem into a new indetermined generative meristem, that gives rise to a new formation of another indetermined meristem, and several normal and abnormal terminal flowers. The abnormal flowers again show the fusion of different structures, in this case from sepals, petals and stamen together (Figure 34). Surprisingly, directly on the generative stem another structure, resembling a single stamen was detectable. All these data indicate that a decrease in RKS1 expression levels results in switches in the meristematic identity. Meristems can switch forward and backward between developmental stages, indicating that RKS10 is normally involved in regulating the meristematic identity and the developmental order of meristematic development. RKS13 seems to be involved in similar processes, as can be concluded from the switches in flower meristematic outgrowths observed in figure 35. Modification of the expression levels of RKS1 also results in modified meristem identity. Suppression of endogenous RKS1 levels results in a developmental switching of generative meristems towards vegetative meristems, together with other phenotypes (results not shown).

Literature

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7. Male sterility

Male sterility is a highly desired trait in many plant species. For example, manipulation of pollen development is crucial for F1 hybrid seed production, to reduce labour costs and for the production of low-envirnmental impanct genetically engineerded crops. In order to produce hybrid seed from inbred plant lines, the male organs are removed from each flower, and pollen from another parent is applied manually to produce the hybrid seed. This labour-intensive method is used with a number of vegetables (e.g. hybrid tomatoes) and with many ornamental plants. Transgenic appoaches, in which one or more
introduced gene products interfere with normal pollen initiation and development is therefore highly desired. Expecially when the number of revertants (growing normal pollen) is extremely low.

Male sterility in plants is a desired trait that has been shown already in many plant species as a result of the inactivation of expression of a number of genes essential for proper stamen develpment, mitotic divisions in the pollen stem cells, or male gametogenesis. A method for modulating a developmental pathway of a plant or plant cell comprising modifying a gene or modifying expression of said gene, wherein said gene is encoding a protein belonging to a signaling complex comprising RKS protein, ELS protein, NDR/NHL protein, SBP/SPL protein and RKS/ELS ligand protein, allowing modulating pollen development, in particular wherein said gene comprises an ELS2 or RKS10 gene or functional equivalent thereof.

Here we present data that show that overexpression of gene products, like transmembrane receptor kinases (RKS) and extracellular proteins (ELS) can also result in the formation of male sterility. The ability to induce male sterility by overexpressing specific genes as provided herein allows the opportunity to produce transgenic overexpressing plants in which the pollen development is inhibited. Stable single copy

homozygous integration of such overexpressing traits into the plant genome will render such plants completely sterile, making them excellent material for the production of F1 hybrid seed. Furthermore, the combined integration of a male sterility inducing overexpressing gene coupled directly with another desired transgene result in transgenic plants which are unable to produce transgenic seed, making these transgenic plants excellent material for outside growth without problems affecting transgenic pollen spreading throughout the environment, thereby eliminating possible crosses with wild plant species or other non-transgenic crops. The combination of a desired transgene flanked on both sites by different male-sterility inducing overexpressing genes would decrease the frequency of pollen formation to an extremely low level. An example is an overexpressing construct of RKS10 at the 5'end of integrated DNA fragment, the desired transgene expression casette in the middle and at the 3'end of the integrated DNA the ELS2 overexpressing construct. This complete DNA fragment is integrated into the genome by convential techniques, like particle bombardment, *Agrobacterium* transformation etc. Another possible application concerns the modification of pollen in ornamental plant species like lilly, where the release of pollen from cut flowers can be avoided by making transgenic plants in which pollen development is initiated by release from the stamen is prevented (a desired trait that can be obtained by overexpressing for example ELS2, resulting in partial pollen development). Hereby the ornamental value of the stamen with pollen is not lost, but release of pollen is inhibited.

Results obtained

Overexpression and antisense constructs of full length RKS cDNA clones have been made under the control of 35S promoters. Transgenic plants have been produced in *Arabidopsis thaliana*. Subsequent generations of stably transformed plants were investigated for phenotypes and analysed in detail.

T2 transgenic seedlings of *Arabidopsis* were germinated on MS agar plates without hormones. Control transgenic plants containing a negative control vector pGreen4K (empty expression vector) were included as references for normal stamen and pollen development. RKS10 and ELS2 resulted in sterile plants when overexpressed in *Arabidopsis*. Antisense RKS10 plants resulted in a strong reduction in the number of pollen formed (Figure 36). In order to determine whether pollen development itself was the reason for sterility (and not a combination of pollen developmental mutants coupled to either embryo lethals or female gametogenesis defects), reciprocal crosses were performed between sterile transgenic plants and wildstype *Arabidopsis thaliana* WS plants. These results confirmed that the sterile plants with overexpressing RKS10 and ELS2 constructs were male sterile but completely femele fertile. No defects could be observed in embryo development from crosses between female transgenic overexpressors and male wildtype pollen (results not shown). Since both antisense and overexpressing constructs of the RKS10 gene showed defects in proper pollen development we conclude that normal levels of endogenous RKS10 gene product are essential for proper pollen formation, outgrowth and differentiation. In the ELS2 overexpressing plants the intiation of pollen grains was not inhibited. However the proper development of pollen grains in full grown viable pollen was clearly inhibited .

Literature

-The Arabidopsis male sterility1 (MS1) gene is a transcriptional regulator of male gametogenesis, with homology to the PHD-finger family of transcription factors. Wilson et al. 2001. the Plant Journal 28: 27-39

-Transposon tagging of a male sterility gene in Arabidopsis. Aarts et al. 1993. Nature 363: 715-717

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Science 1997: 278, 1963-1965

Plant Physiol. Biochem. 2000: 38, 789-796

Cell 1996: 84, 61-71

Annu. Rev. Plant Physiol. Plant Mol. Biol. 1999: 50, 505-537

Claims

1. A method for modulating a developmental pathway of a plant or plant cell comprising modifying a gene or modifying expression of said gene, wherein said gene is encoding a protein belonging to a signaling complex comprising RKS protein, ELS protein, NDR/NHL protein, SBP/SPL protein and RKS/ELS ligand protein or encoding a protein comprising a ligand for said complex.

2. A method according to claim 1 allowing modulating cellular division during plant growth or organ formation

3. A method according to claim 2 wherein said gene comprises an RKS4 or RKS10 gene or functional equivalent thereof.

4. A method according to claim 1 allowing modulating apical meristem formation.

5. A method according to claim 4 wherein said gene comprises an ELS1, RKS0, RKS3, RKS4, RKS8 or RKS10 gene or functional equivalent thereof.

6. A method according to claim 4 allowing modulating fasciation.

7. A method according to claim 6 wherein said gene comprises an RKS0, RKS3, RKS8 or RKS10 gene or functional equivalent thereof.

8. A method according to claim 4 allowing modulating root development.

9. A method according to claim 7 wherein said gene comprises an ELS1, ELS 2, RKS1, RKS3, RKS4, RKS6, RKS8 or RKS10 gene or functional equivalent thereof.

10. A method according to claim 4 allowing modulating meristem identity.

11. A method according to claim 9 wherein said gene comprises an ELS1, RKS8, RKS10 or RKS13 gene or functional equivalent thereof.

12. A method according to claim 1 allowing modulating pollen development.

13. A method according to claim 11 wherein said gene comprises an ELS2 or RKS10 gene or functional equivalent thereof.

14. A method for obtaining a plant or plant cell with a modulated development comprising subjecting a plant or plant cell to a method according to anyone of claims 1 to 12.

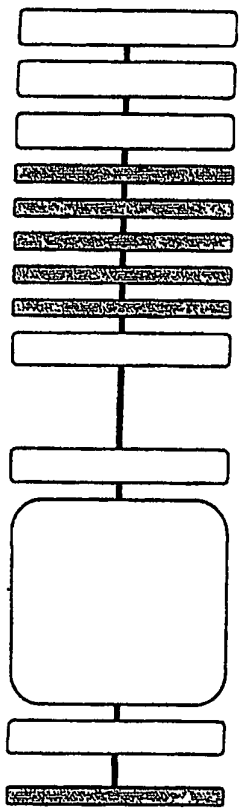
15. A plant or plant cell obtainable with a method according to claim 13.

Abstract

Title: Modulating developmental pathways in plants.

The invention relates to a method to modulate plant growth or development by modifying genes in plants. The invention among others relates to modifying RKS genes or gene products as found in Arabidopsis thaliana or other plants. The invention provides a method for modulating a developmental pathway of a plant or plant cell comprising modifying a gene or modifying expression of said gene, wherein said gene is encoding a protein belonging to a signaling complex comprising RKS protein, ELS protein, NDR/NHL protein, SBP/SPL protein and RKS/ELS ligand protein.

Figure 1
Different domains of RKS proteins



Signal Sequence

Leucine Zipper Domain

Cystein Motif

4-5 LRRs

Cystein Motif /
Serine and Proline rich

Transmembrane Domain

Unknown domain

Serine-Threonine
Kinase Domain

Unknown domain

1 LRR

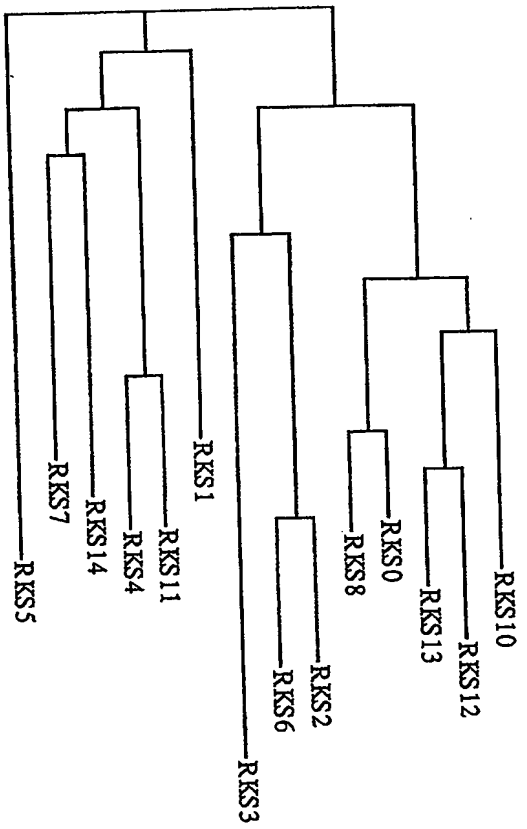


Figure 2
Developmental tree of the different Receptor Kinases like SERK (RKS) genes.

Figure 3
Intron-Exon structure of the RKS genes in *Arabidopsis thaliana* var. Columbia.
SS signal sequence; LRR leucine rich repeat domain; TM transmembrane domain.

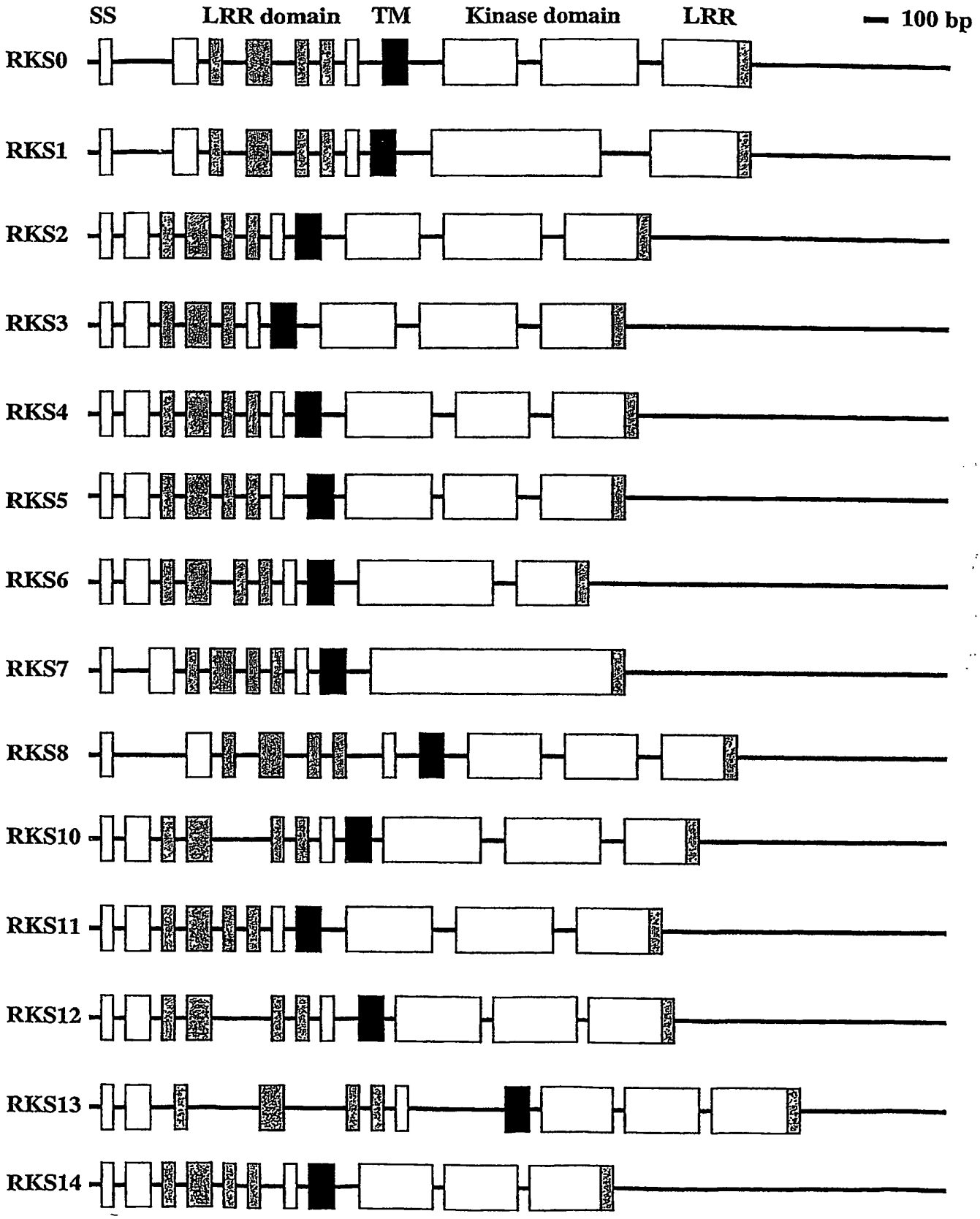


Fig. 4

Chromosomal location of RKS genes in *Arabidopsis thaliana*

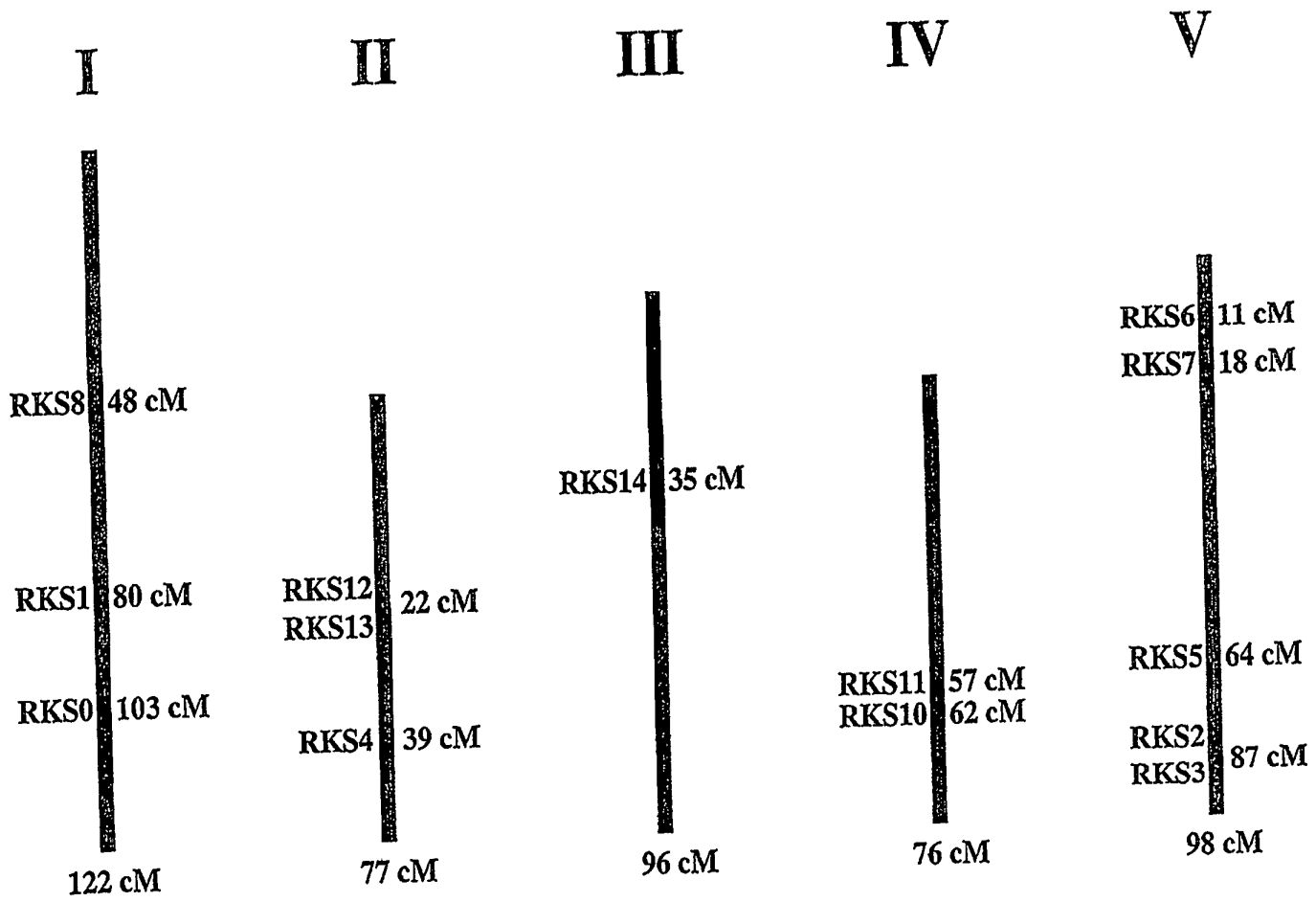
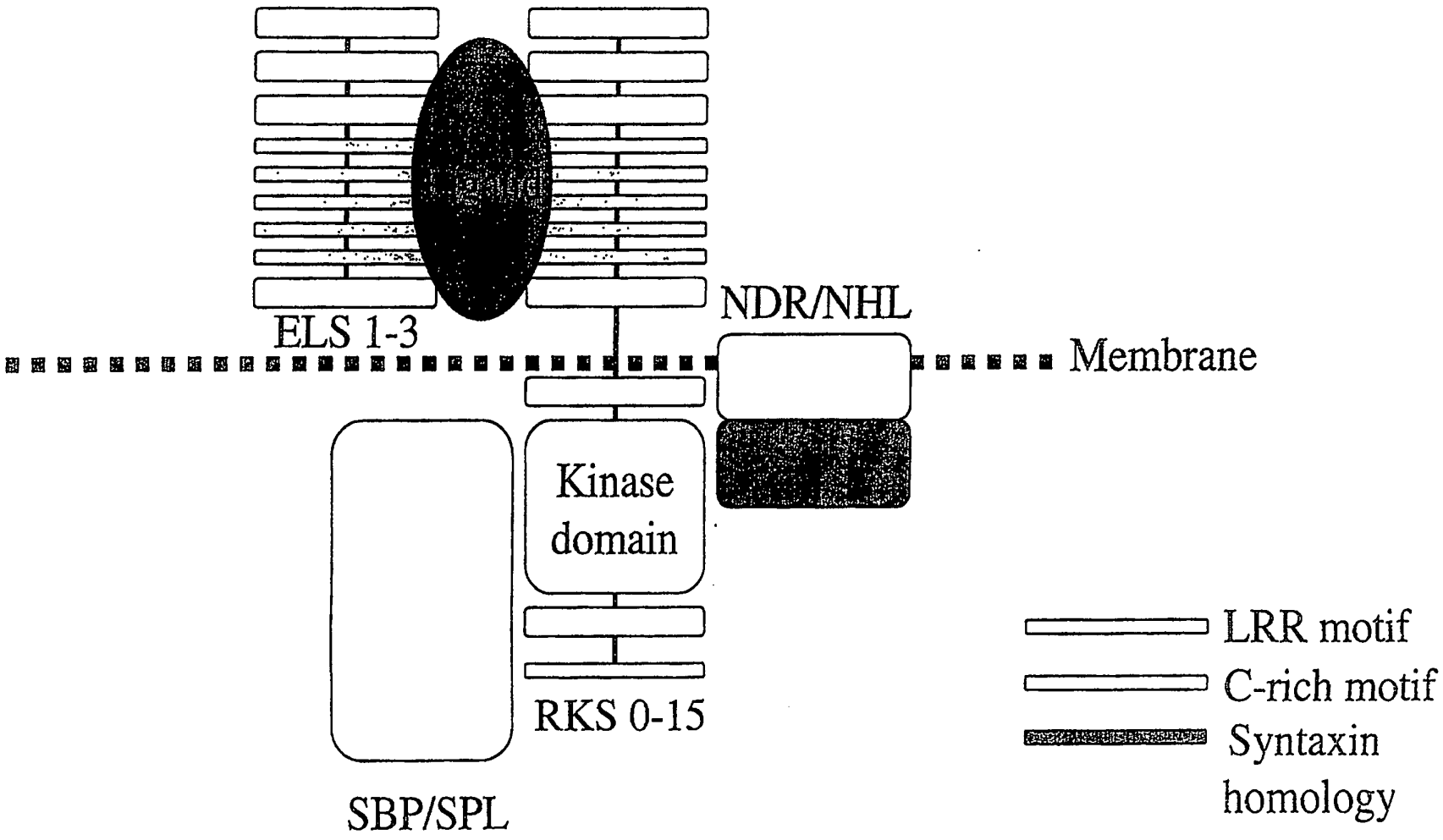


Fig. 5

RKS-mediated signal transduction pathway in plants

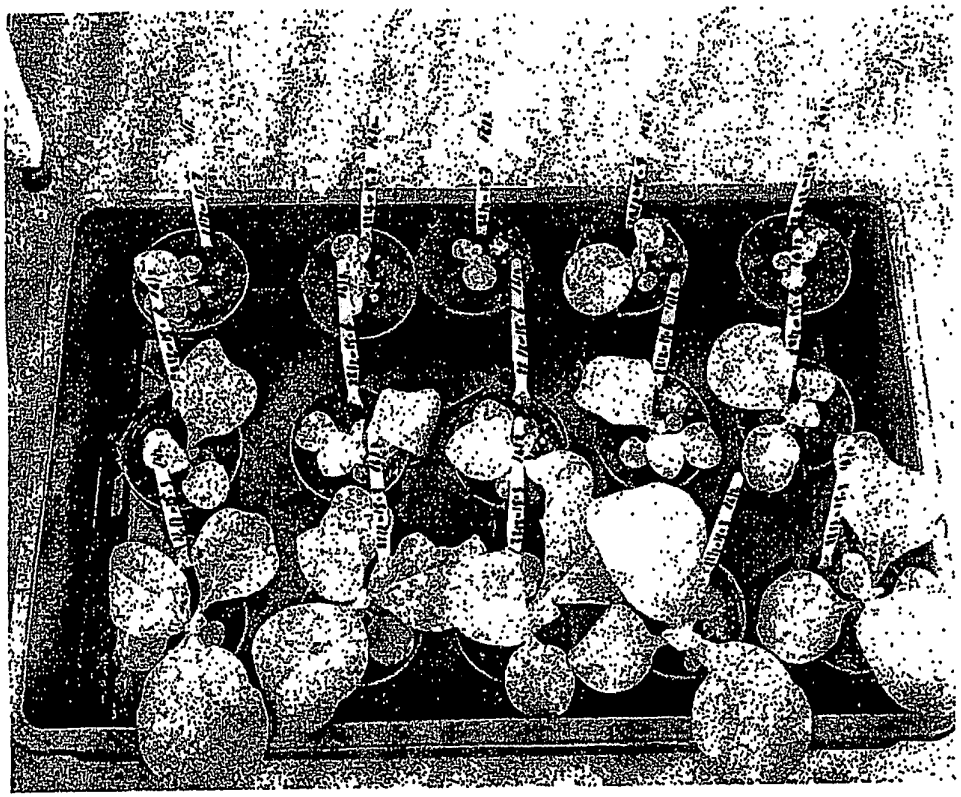


GT-RKS4 determines seeling size
in *Nicotiana tabacum*.

Modifications in the
expression proficle
of GT-RKS4 modulates
organ size within seedlings
of *Nicotiana tabacum*.

Fig. 7

GT-RKS4 determines organ size in *Nicotiana tabacum*.

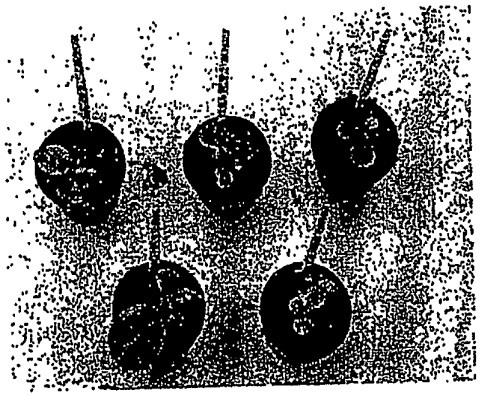


GT-RKS4-7S-T2

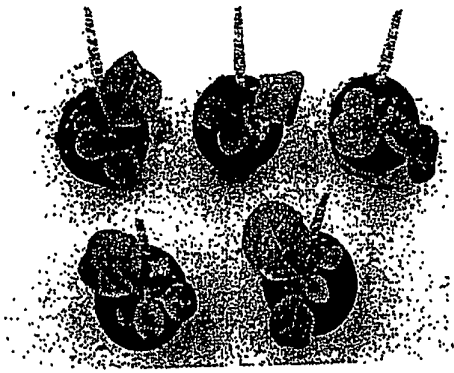
GT-RKS4-6S-T2

GT-RKS4-3S-T2

GT-RKS4 determines plant size in *Nicotiana tabacum*

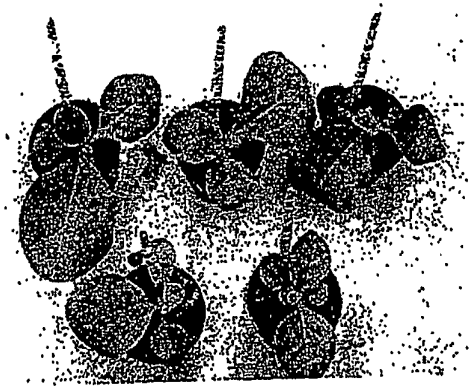


GT-RKS4-15S-7T2

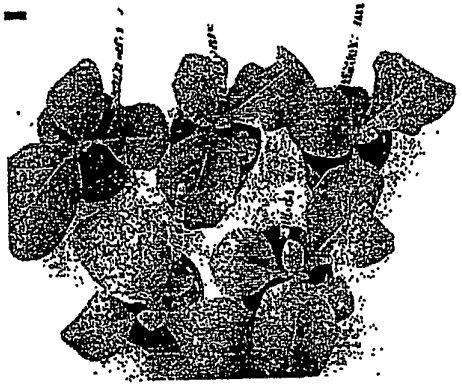


GT-RKS4-15S-6T2

Empty vector control



GT-RKS4-15S-9T2

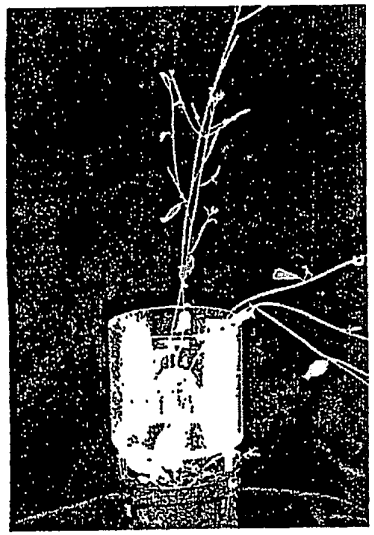


GT-RKS4-15S-3T2

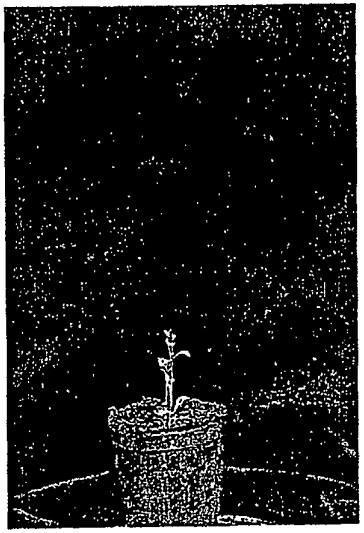
Fig. 9

Stable transformed GT-RKS4-antisense in *Arabidopsis thaliana*

Wildtype WS

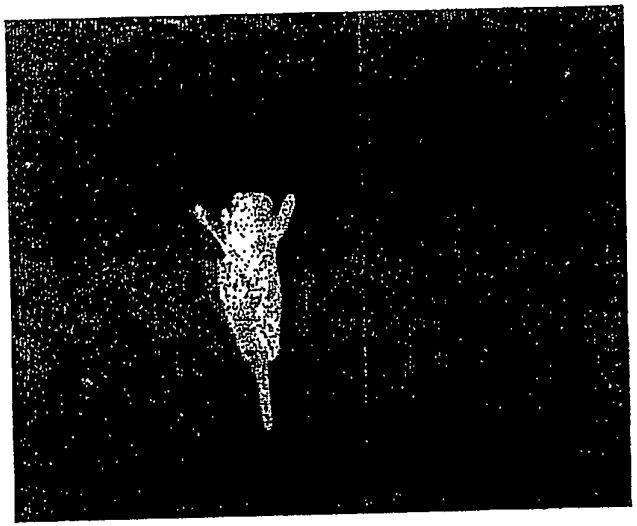


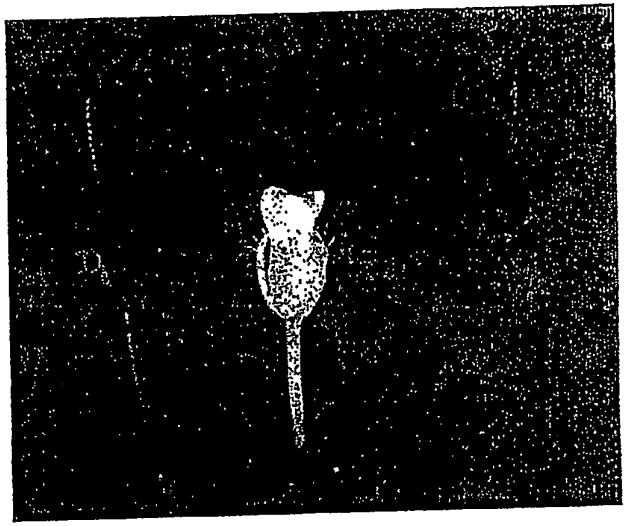
GT-RKS4-16a



Overexpression of antisense GT-RKS4-1a reduces plant and organ size.

GT-RKS4 regulates organ size in *Arabidopsis thaliana*

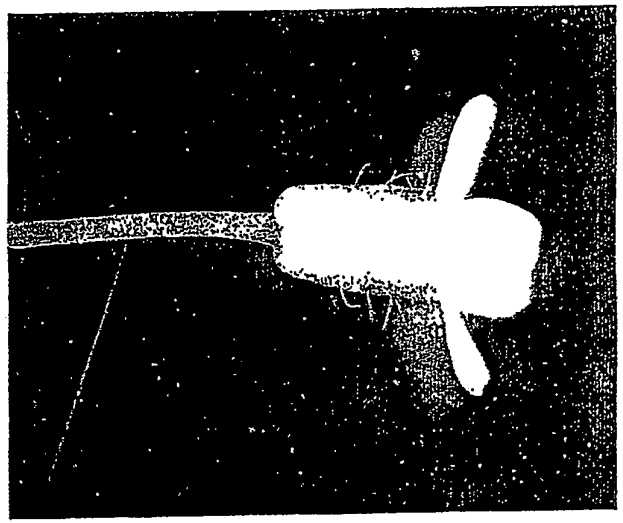




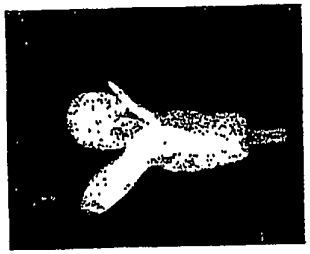
Stable transformed GT-RKS4-s in *Arabidopsis thaliana*

Wildtype WS

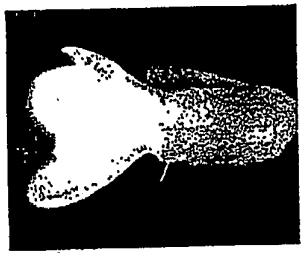
GT-RKS4-6s



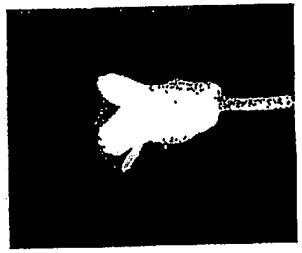
Flowers of Transgenic *Arabidopsis thaliana*



pG4K T1-1; T2



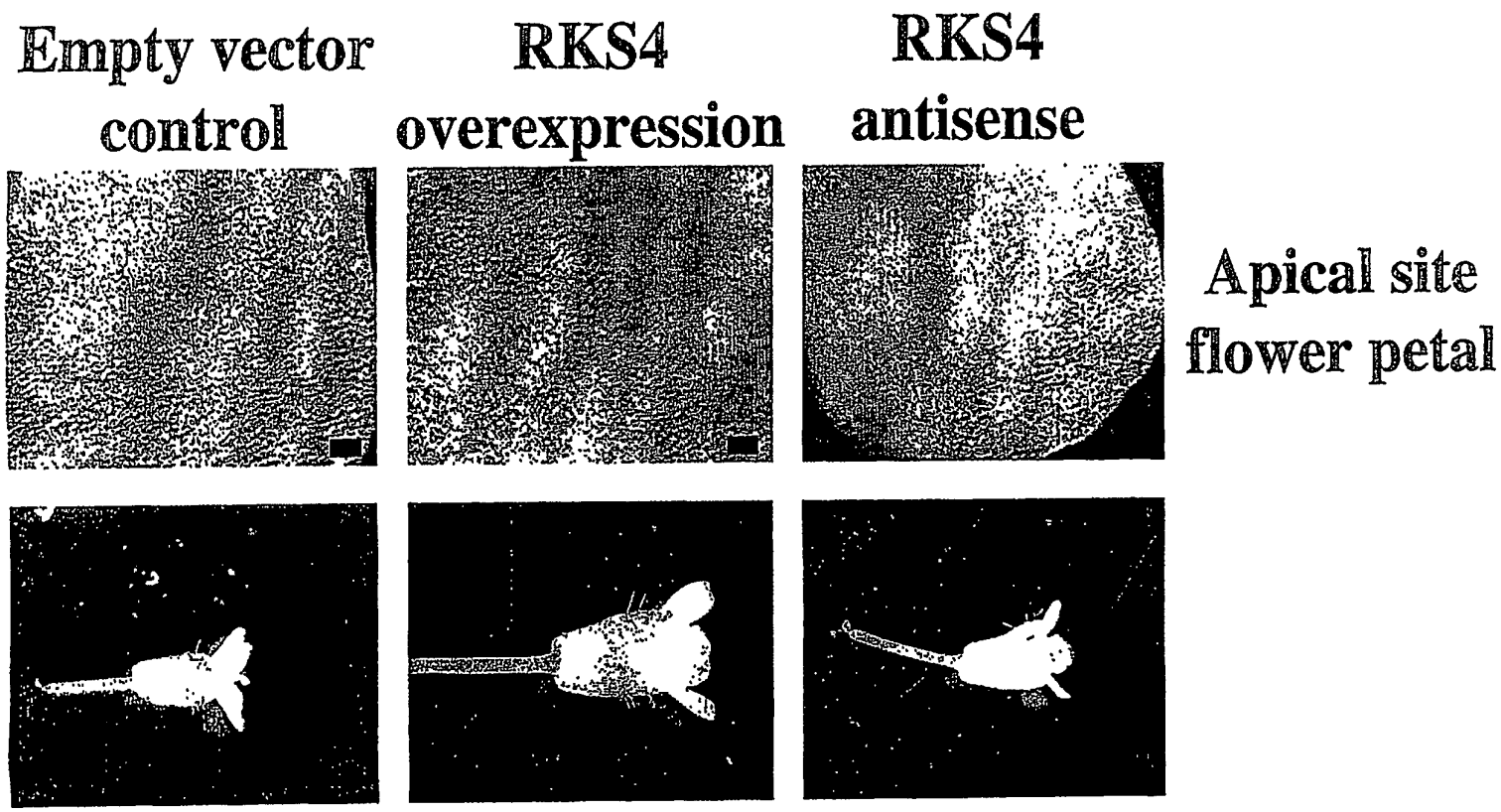
RKS4S T1-6;T25;T3



RKS4aT1-2;T2-6;T3

Fig. 13

RKS4 regulates cell number and cell size in *Arabidopsis thaliana.*



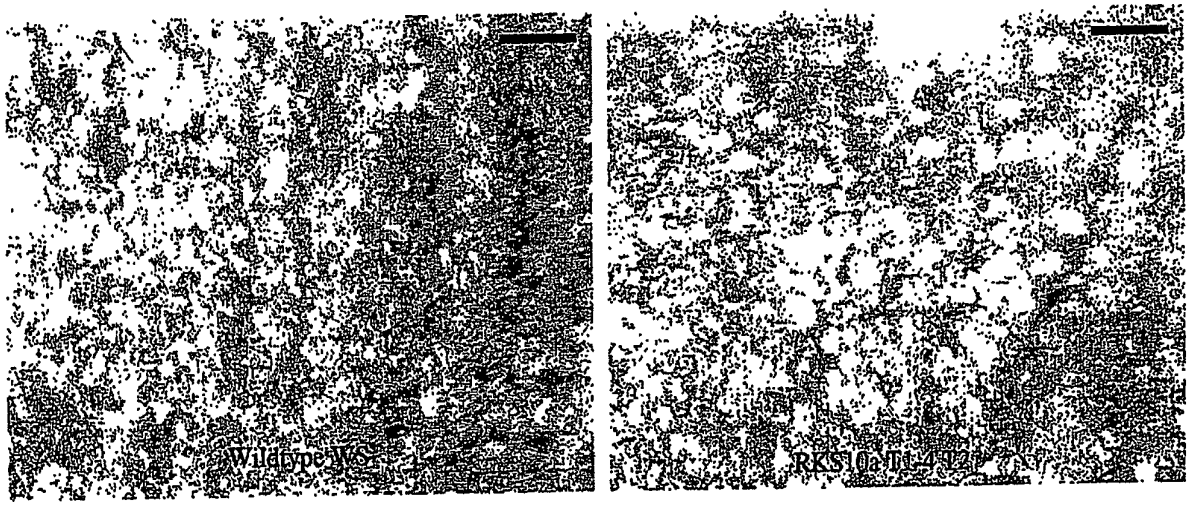
RKS10S T1-10
results in a decrease in size
of cotyl-like apical epidermal cells

pGreen 4K

RKS10S T1-10

Fig. 15

RKS10antisense T1-4
results in an increase in size
of the cotyl epidermal cells



Flower development from the same influorescense in transgenic *Arabidopsis thaliana*

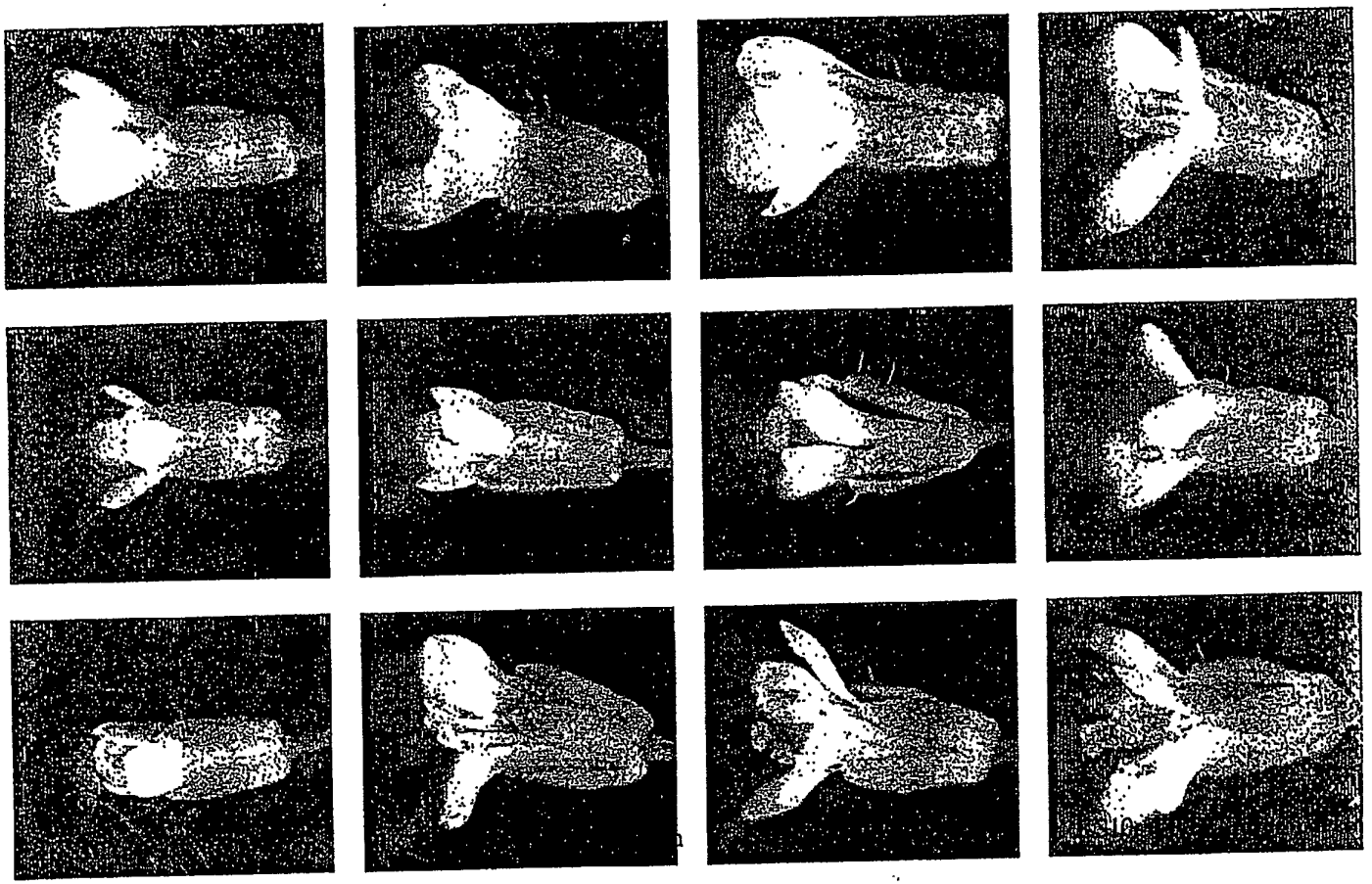
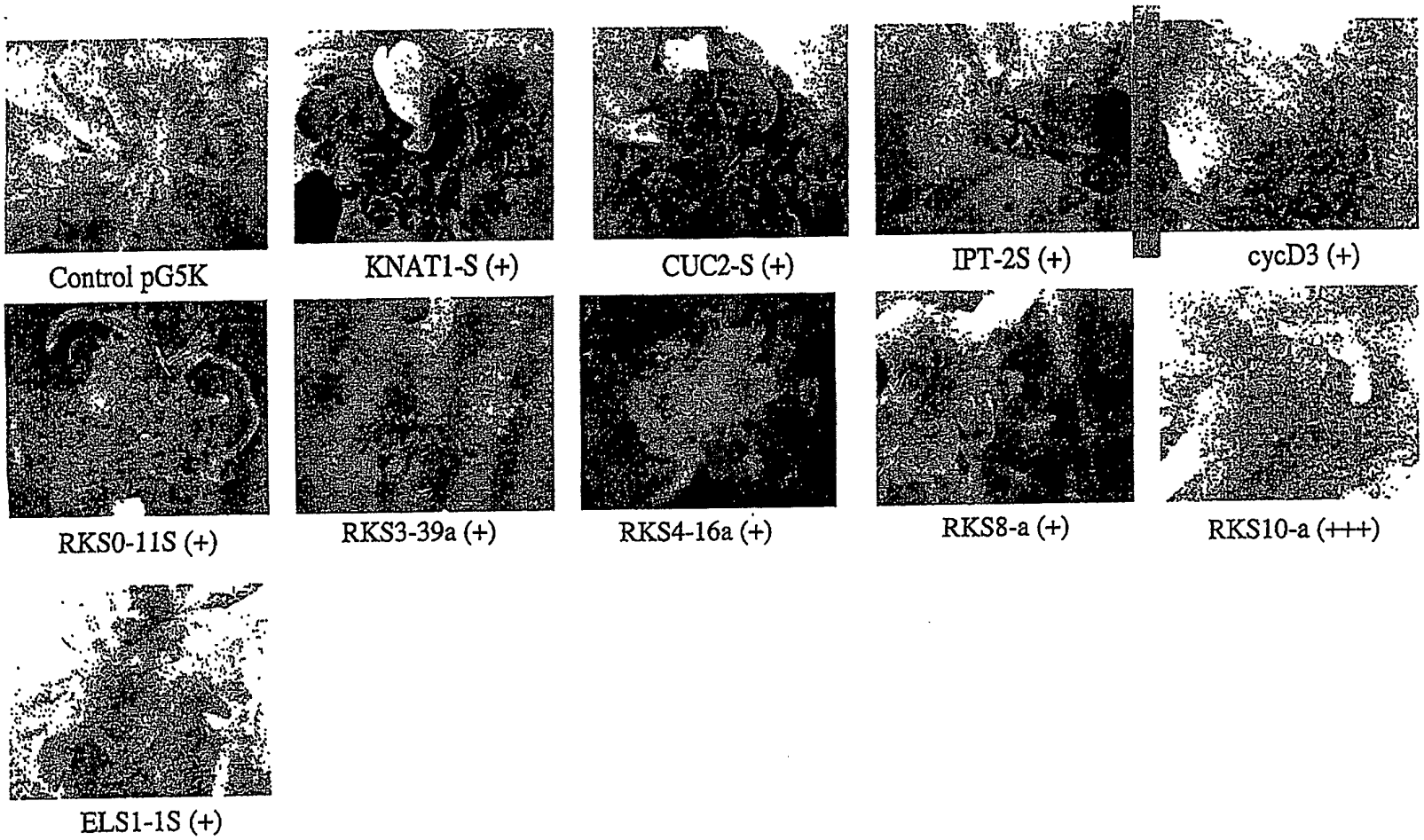


Fig. 17

Regeneration potential of *Arabidopsis* transgenic seedlings.



RKS0 stably transformed is able to induce a continuus regeneration of plants

GT-RKS0-23S transformation of tobacco leaf discs

Proliferating and regenerating RKS0 expressing tissue culture

Continuus regeneration of new plants

Fig. 19

Fasciation in transgenic *Arabidopsis thaliana*

RKS0-7S

RKS1-a

RKS3-39a

RKS8-S

pG5K

RKS10-S

RKS8-S

RKS8-S

Root growth of transgenic *Arabidopsis thaliana*

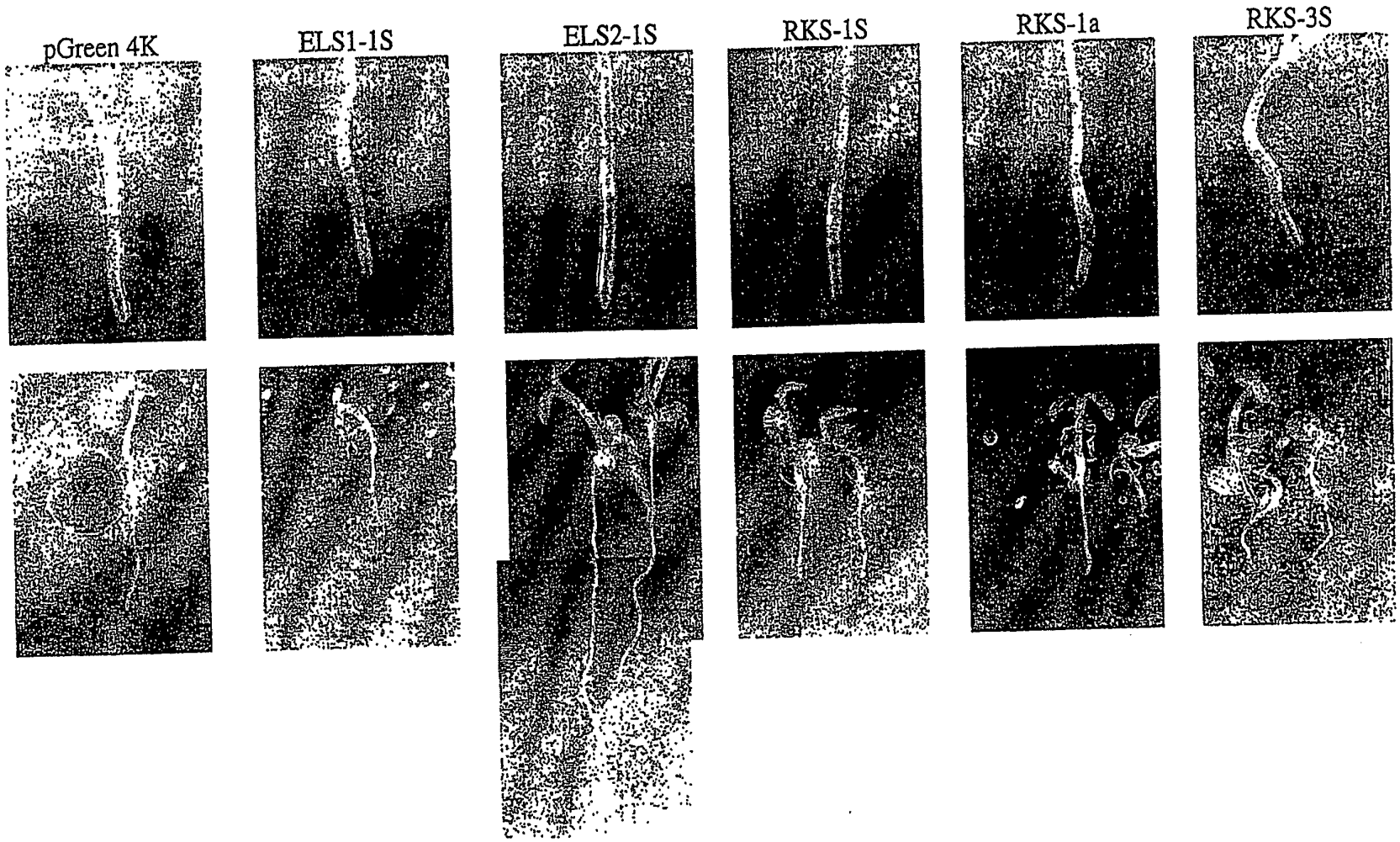
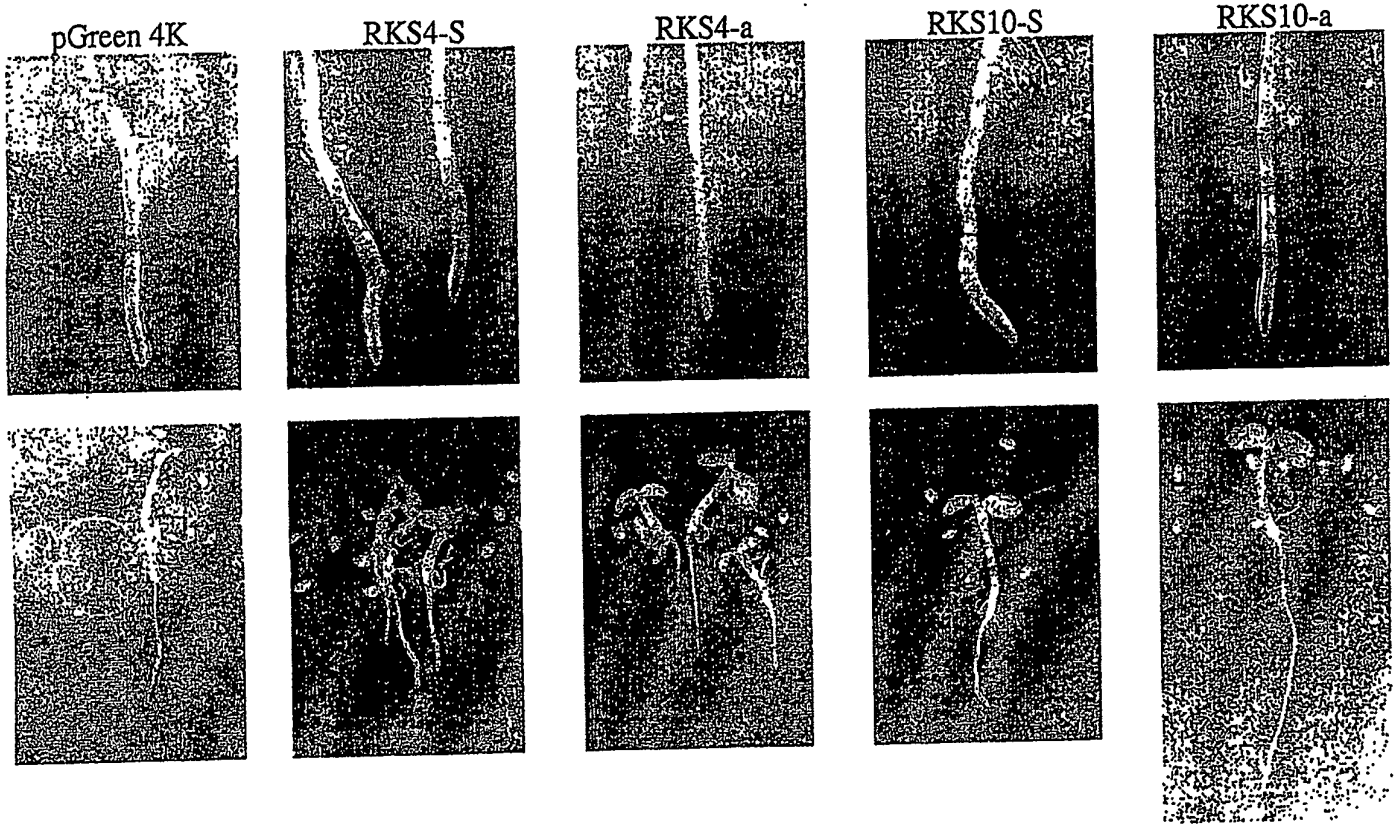


Fig. 21

Root growth of transgenic *Arabidopsis thaliana*



Root growth of transgenic *Arabidopsis thaliana*

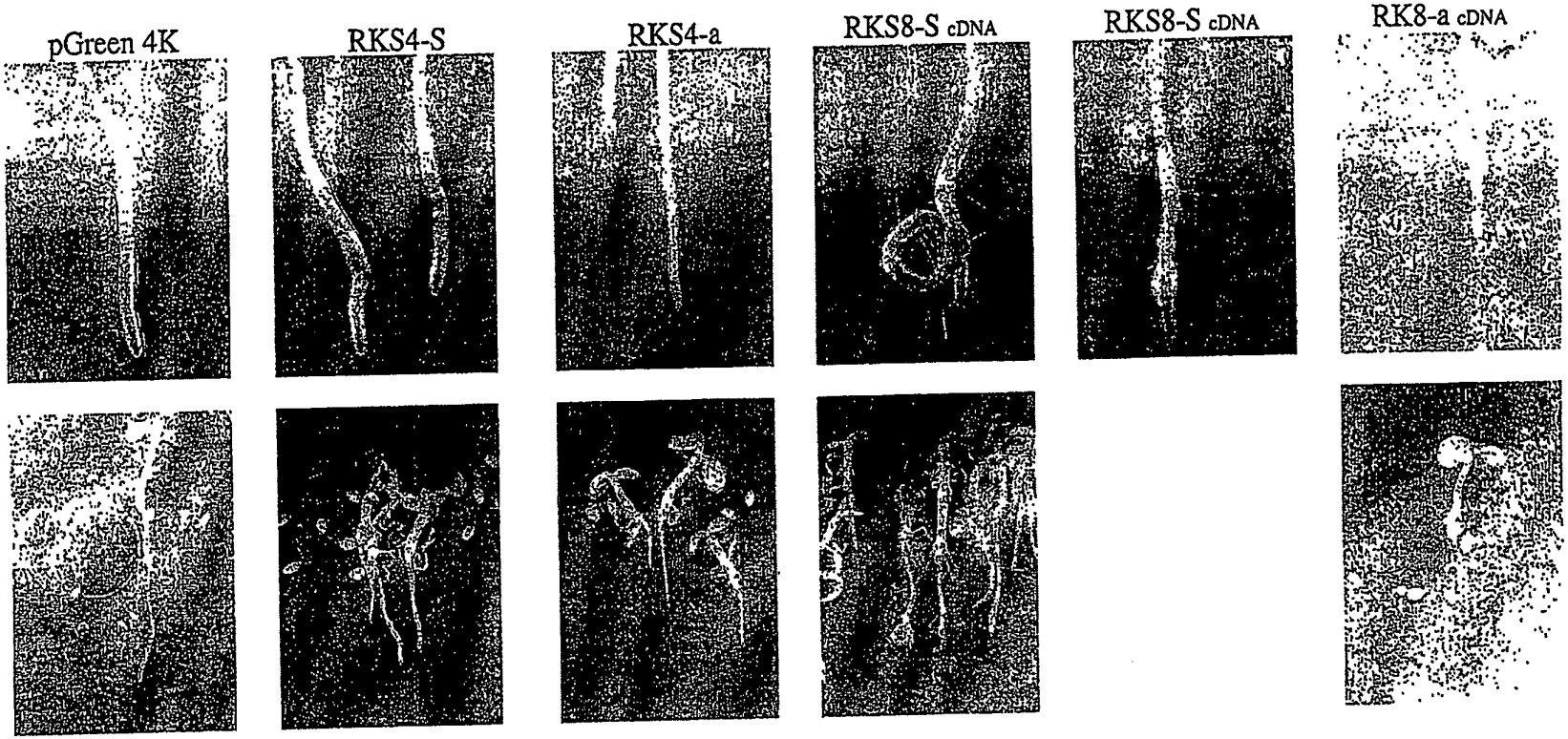
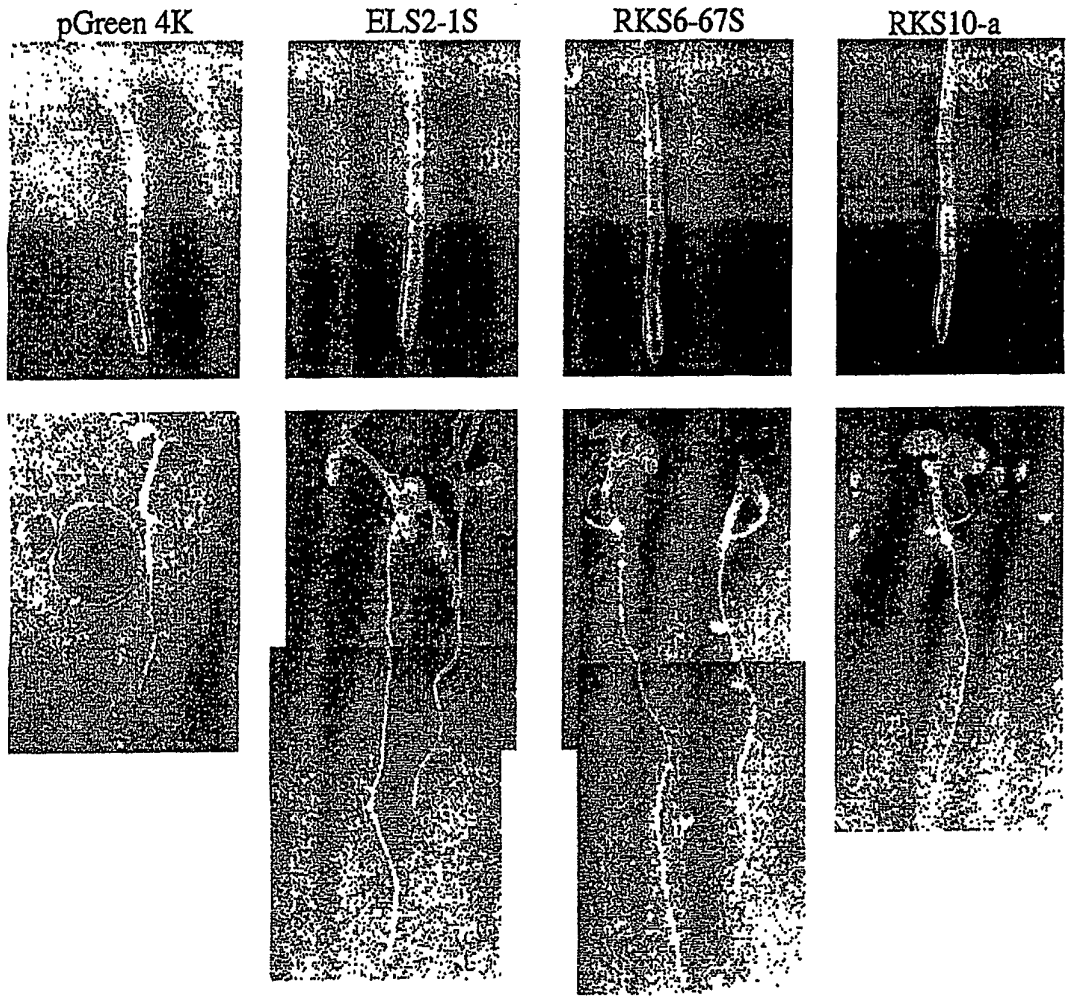


Fig. 23

Root growth of transgenic *Arabidopsis thaliana*



Transgenic *Arabidopsis thaliana* primary root length after 14 days of germination

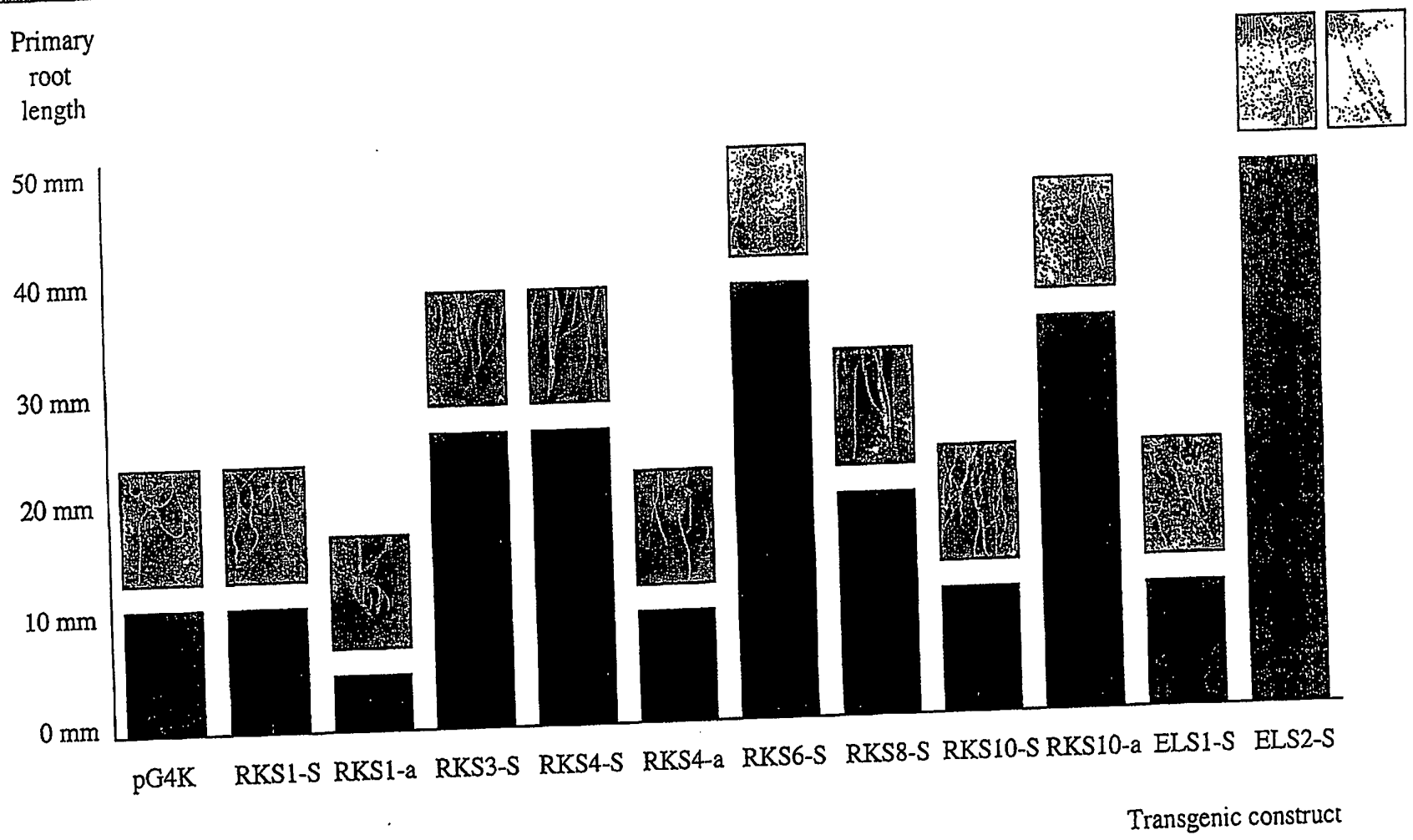
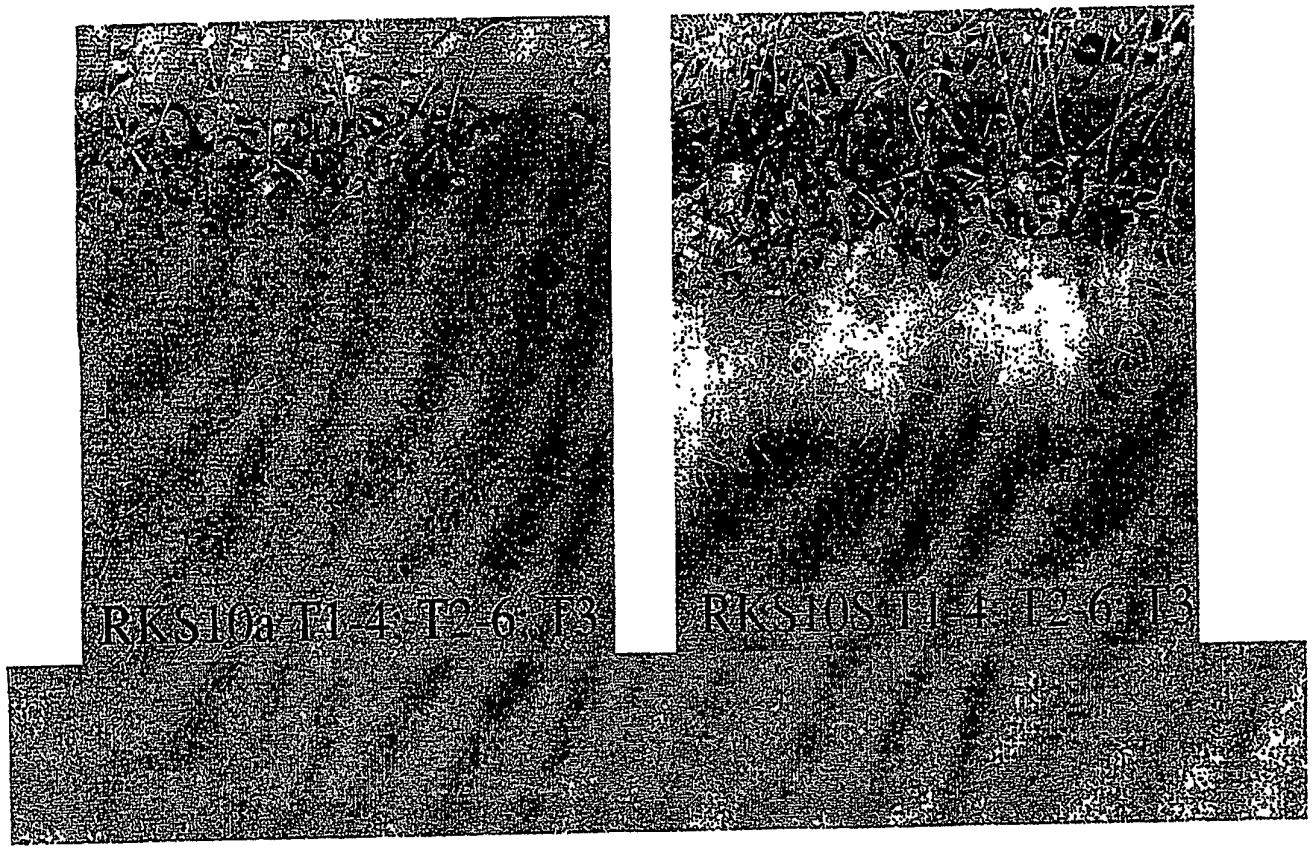


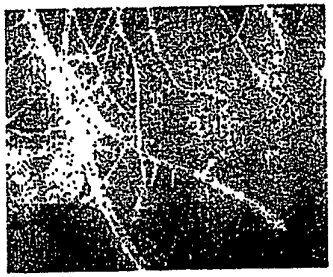
Fig. 25

Effects of RKS10 transgenic constructs on plant development of 45 days old *Arabidopsis* WS



Roots of Transgenic *Arabidopsis thaliana*

pG4K



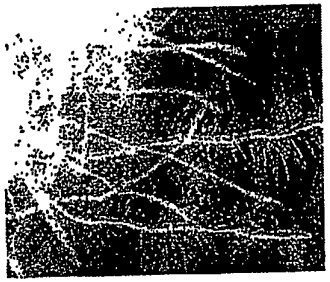
RKS8-S-cDNA

RKS3-39a

RKS10-S S

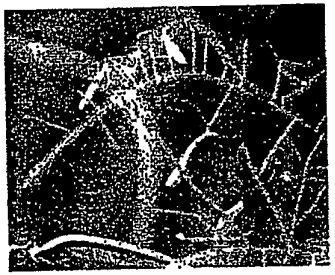
pG5K

RKS8-a

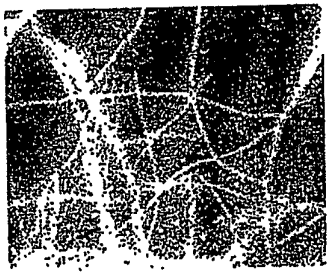


RKS3-S

RKS10-S R



RKS1-a

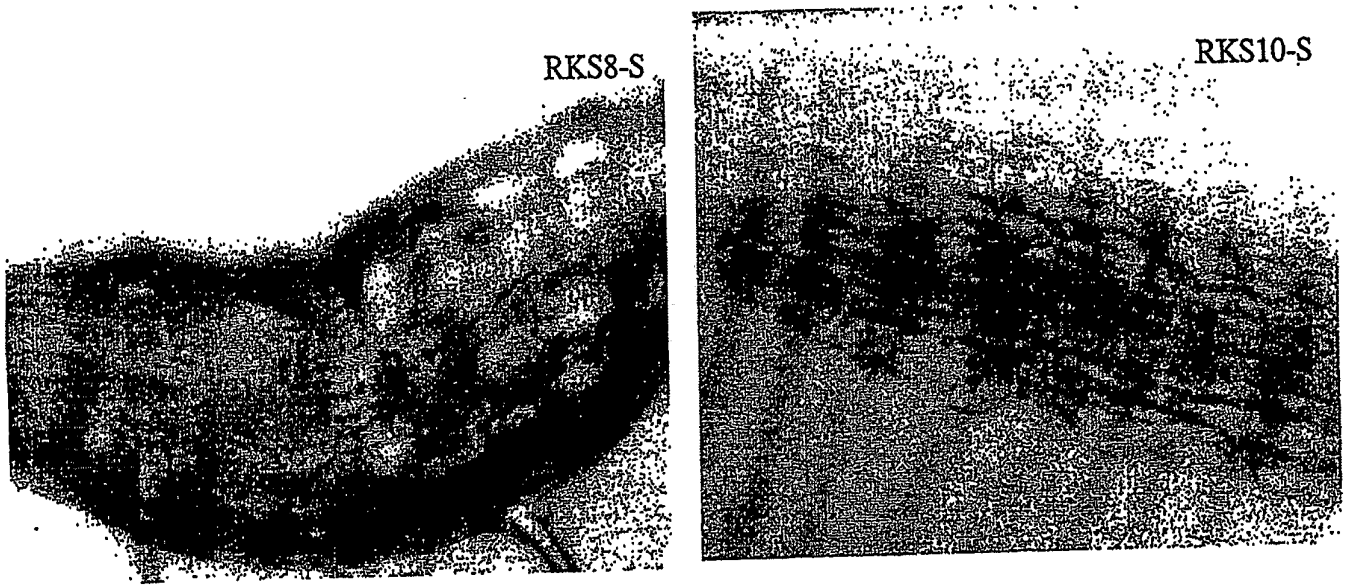


ELS1-S

RKS10-a

Fig. 27

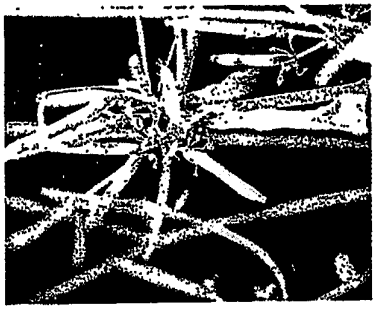
Root cells of transgenic
Arabidopsis thaliana



Influorescences of T1 transgenic *Arabidopsis* WS plants

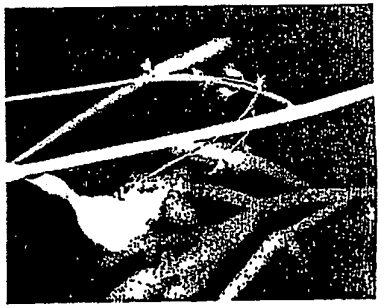
RKS10-S-T1-10

RKS10-a-T2



RKS8-a-T1-10

ELS-1-T1



Control pGreen4K

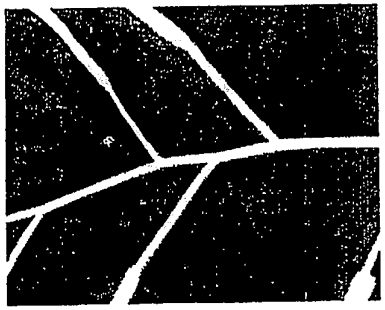
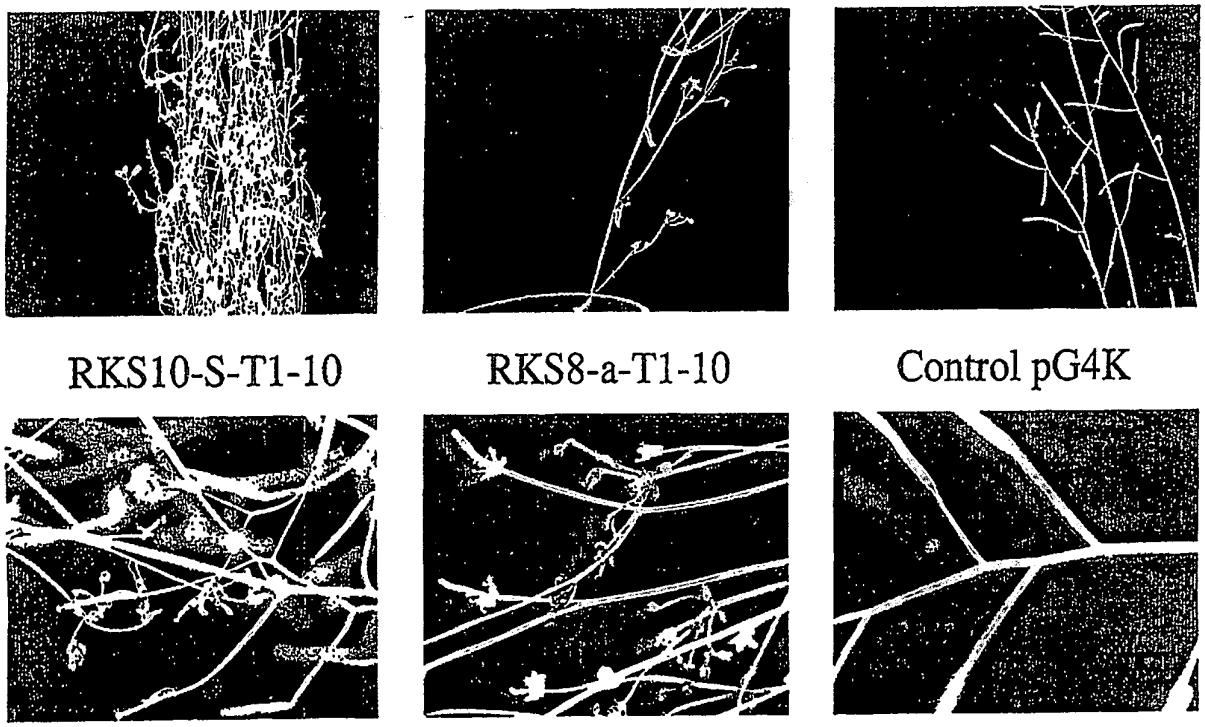
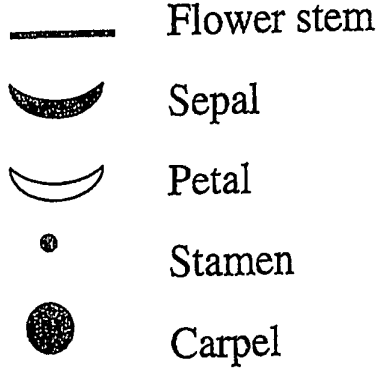


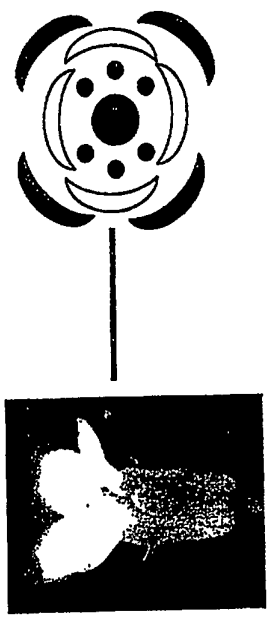
Fig. 29

Influorescences of T1 transgenic *Arabidopsis* WS plants



RKS10a T1 expression constructs in *Arabidopsis thalinana*





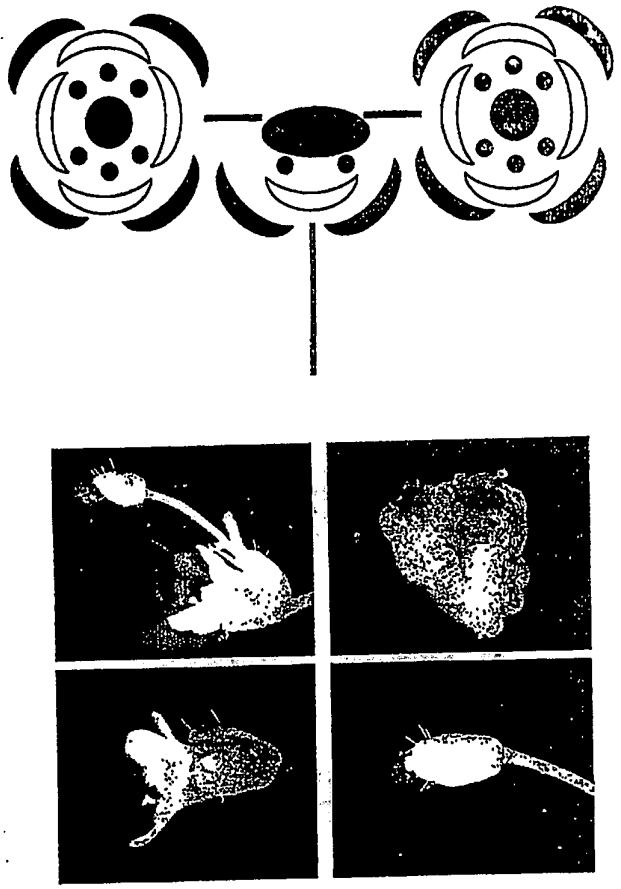
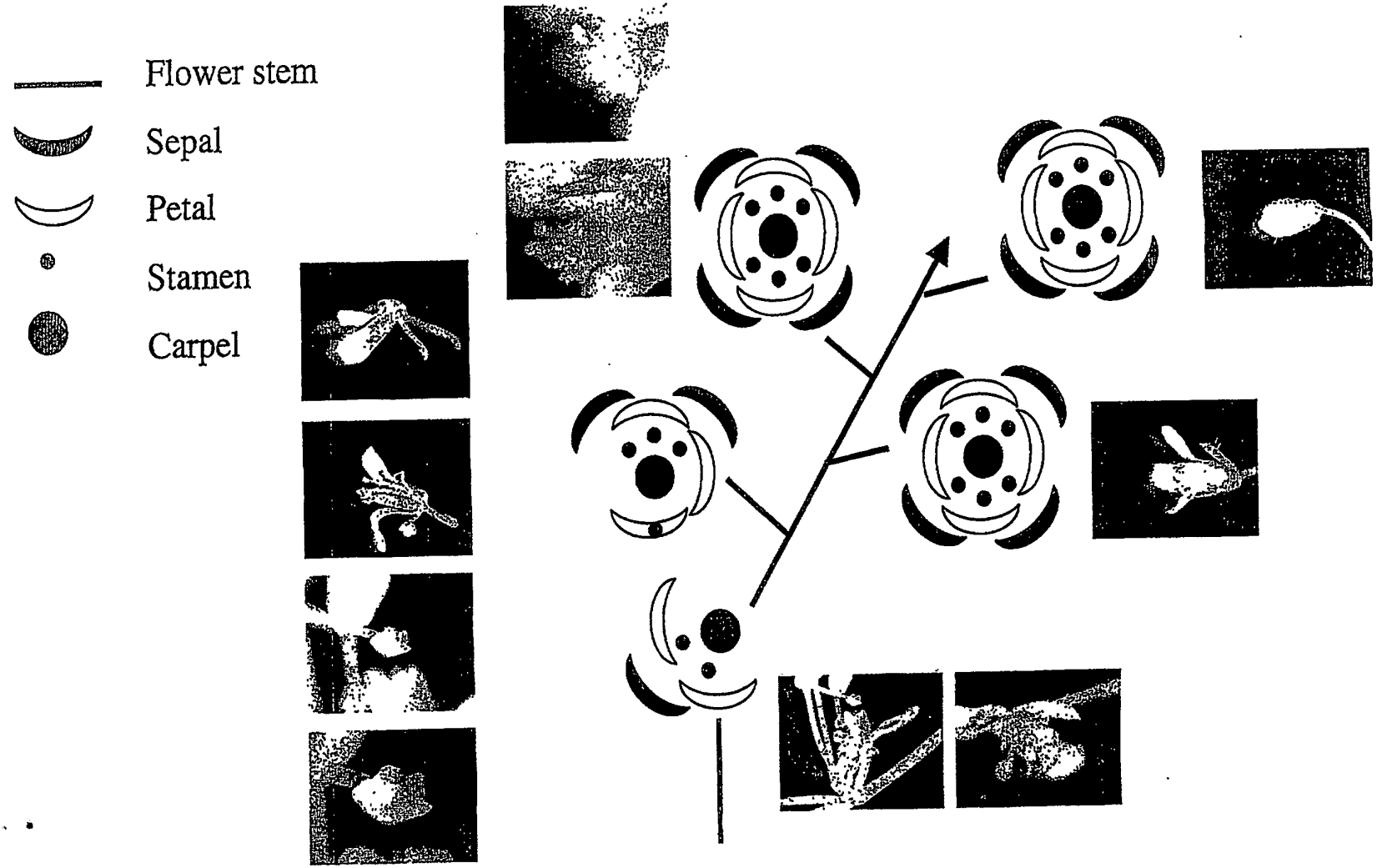
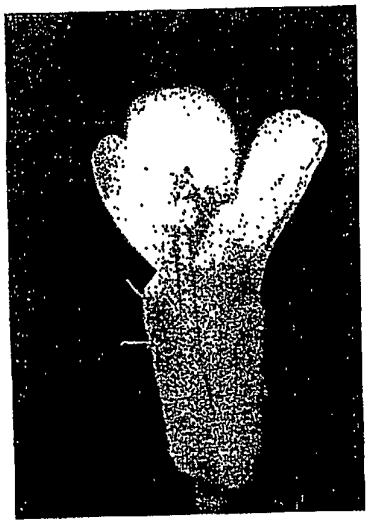


Fig. 31

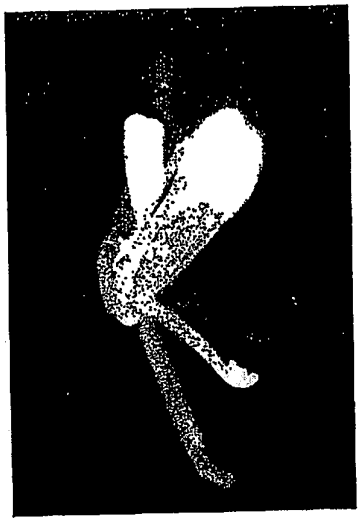
RKS10a T1-11 in *Arabidopsis thalinana*



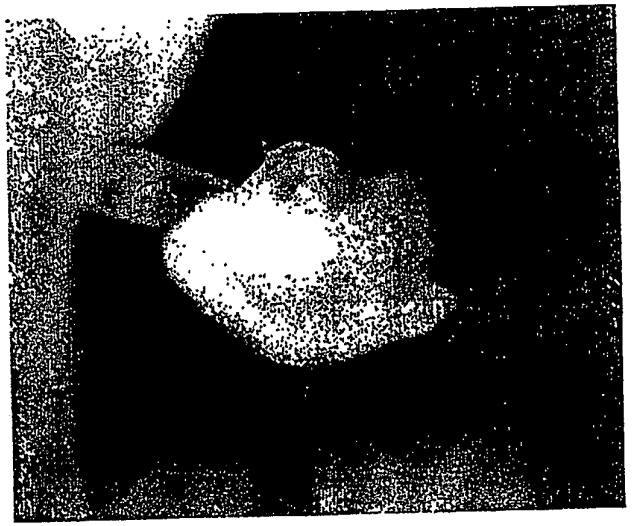
RKS10 antisense effects in *Arabidopsis thaliana*



pGreen 4K



RKS10a T1-11



detail flower RKS10a T1-11

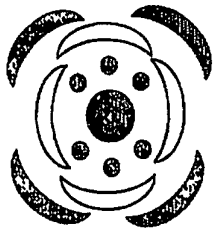
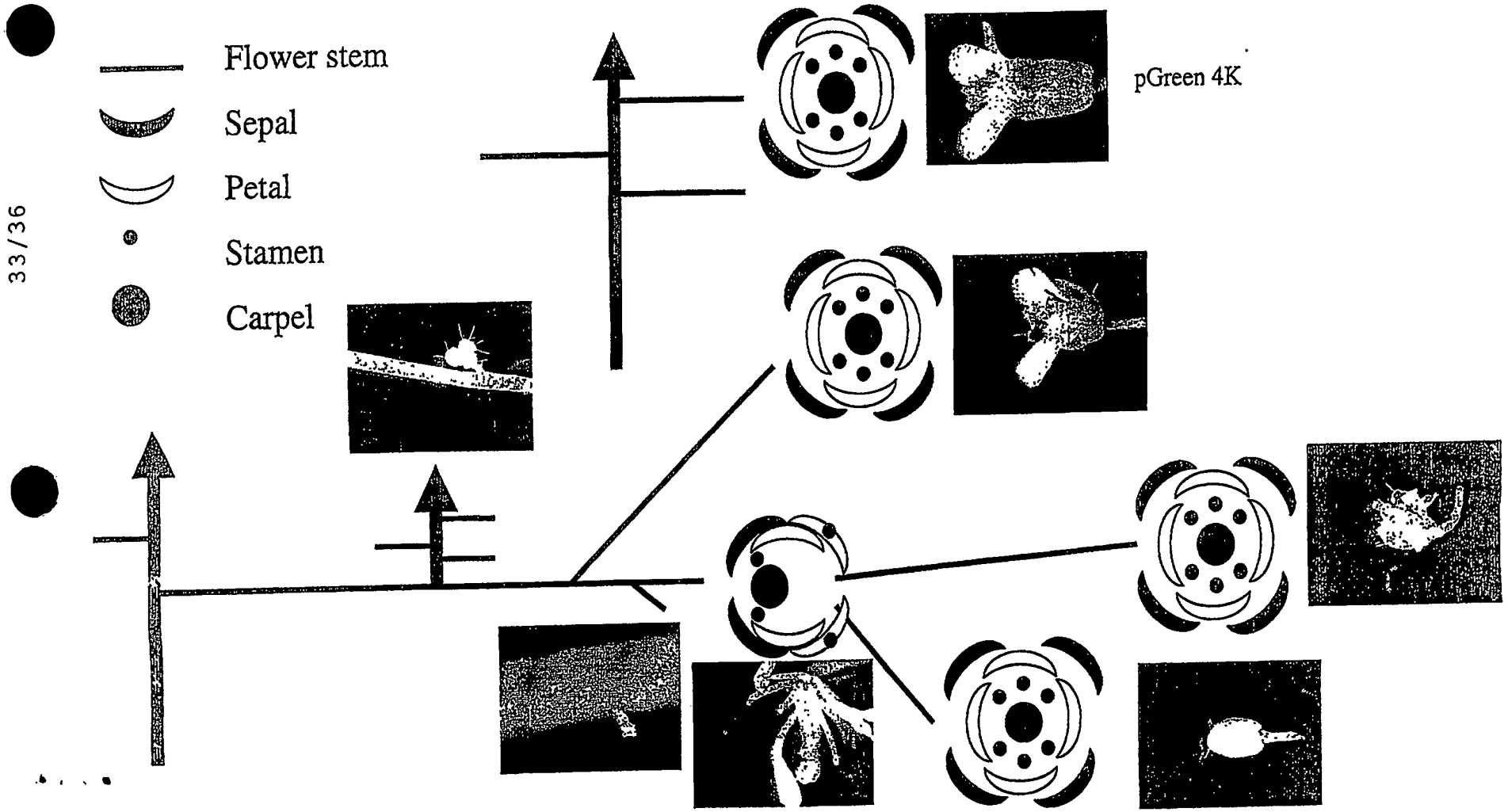


Fig. 33

RKS10a T1-12 in *Arabidopsis thalinana*



RKS10a T1-12 in *Arabidopsis thaliana*

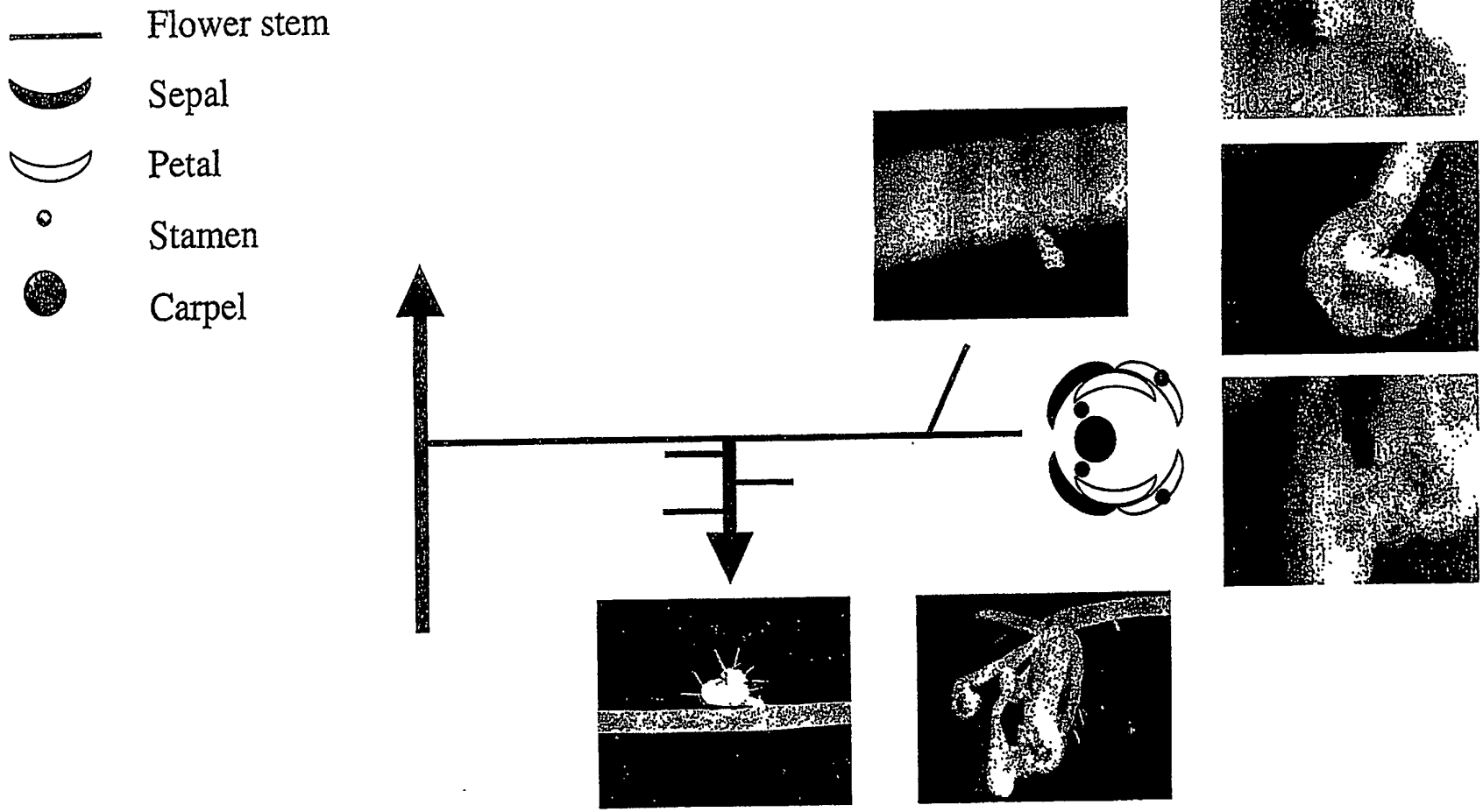
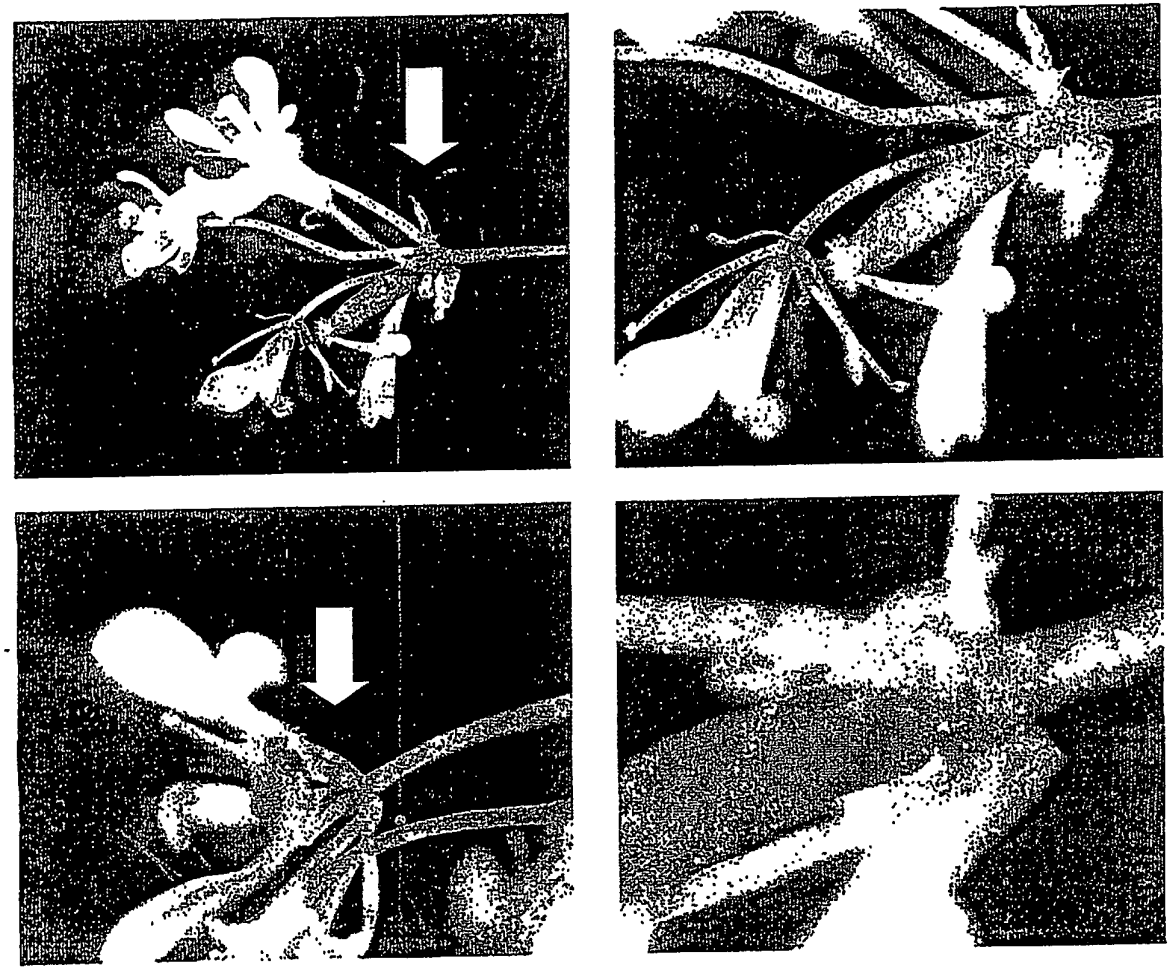
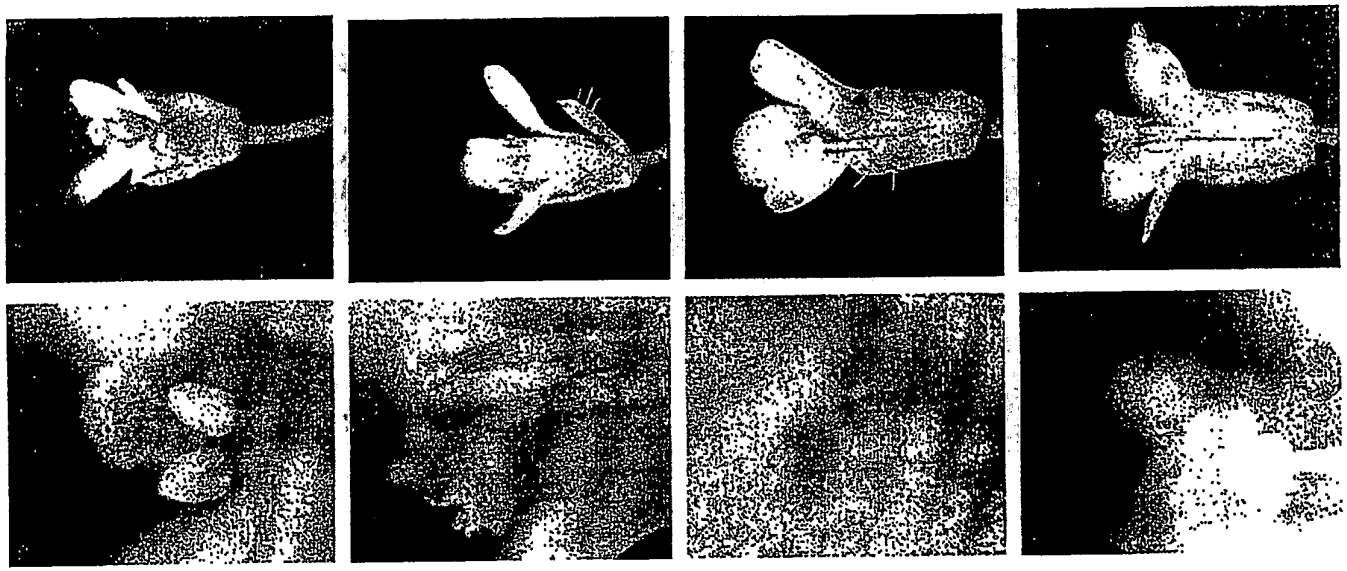


Fig. 35

RKS13 regulates flower meristem identity in *Arabidopsis thaliana*



Male sterile transgenes in *Arabidopsis thaliana*



RKS10S T1-10
no pollen formed

RKS10a T1-11
almost no pollen

pGreen4K
normal pollen

ELS 2 157.21S T1-11 T2-2
pollen development aborted